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(54) Title: PRODUCTION OF POLYUNSATURATED FATTY ACIDS BY EXPRESSION OF POLYKETIDE-LIKE SYNTHESIS GENES IN PLANTS

(57) Abstract

The present invention relates to compositions and methods for preparing polyunsaturated long chain fatty acids in plants, plant parts and plant cells, such as leaves, roots, fruits and seeds. Nucleic acid sequences and constructs encoding PKS-like genes required for the poly-unsaturated long chain fatty acid production, including the genes responsible for eicosapentenoic acid production of Shewanella putrefaciens and novel genes associated with the production of docosahexenoic acid in Vibrio marinus are used to generate transgenic plants, plant parts and cells which contain and express one or more transgenes encoding one or more of the PKS-like genes associated with such long chain polyunsaturated fatty acid production. Expression of the PKS-like genes in the plant system permits the large scale production of polyunsaturated long chain fatty acids such as eicosapentenoic acid and docosahexenoic acid for modification of the fatty acid profile of plants, plant parts and tissues. Manipulation of the fatty acid profiles allows for the production of commercial quantities of novel plant oils and products.

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PRODUCTION OF POLYUNSATURATED FATTY ACIDS BY EXPRESSION OF POLYKETIDE-LIKE SYNTHESIS GENES IN PLANTS

INTRODUCTION

5 Field of the Invention

This invention relates to modulating levels of enzymes and/or enzyme components capable of modifying long chain poly-unsaturated fatty acids (PUFAs) in a host cell, and constructs and methods for producing PUFAs in a host cell. The invention is exemplified by production of eicosapentenoic acid (EPA) using genes derived from Shewanella putrefaciens and Vibrio marinus.

Background

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Two main families of poly-unsaturated fatty acids (PUFAs) are the ω3 fatty acids, exemplified by eicosapentenoic acid, and the ω6 fatty acids, exemplified by arachidonic acid. PUFAs are important components of the plasma membrane of the cell, where they can be found in such forms as phospholipids, and also can be found in triglycerides. PUFAs also serve as precursors to other molecules of importance in human beings and animals, including the prostacyclins, leukotrienes and prostaglandins. Long chain PUFAs of importance include docosahexenoic acid (DHA) and eicosapentenoic acid (EPA), which are found primarily in different types of fish oil, gamma-linolenic acid (GLA), which is found in the seeds of a number of plants, including evening primrose (*Oenothera biennis*), borage (*Borago officinalis*) and black currants (*Ribes nigrum*), stearidonic acid (SDA), which is found in marine oils and plant seeds, and arachidonic acid (ARA), which along with GLA is found in filamentous fungi. ARA can be purified from animal tissues including liver and adrenal gland. Several genera of marine bacteria are known which synthesize either EPA or DHA. DHA is present in human milk along with ARA.

PUFAs are necessary for proper development, particularly in the developing infant brain, and for tissue formation and repair. As an example, DHA, is an important constituent of many human cell membranes, in particular nervous cells (gray matter), muscle cells, and spermatozoa and believed to affect the development of brain functions in general and to be essential for the development of eyesight. EPA and DHA have a number of nutritional and pharmacological uses. As an example adults affected by diabetes (especially non insulin-dependent) show deficiencies and imbalances in their

levels of DHA which are believed to contribute to later coronary conditions. Therefore a diet balanced in DHA may be beneficial to diabetics.

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For DHA, a number of sources exist for commercial production including a variety of marine organisms, oils obtained from cold water marine fish, and egg yolk fractions. The purification of DHA from fish sources is relatively expensive due to technical difficulties, making DHA expensive and in short supply. In algae such as Amphidinium and Schyzochytrium and marine fungi such as Thraustochytrium DHA may represent up to 48% of the fatty acid content of the cell. A few bacteria also are reported to produce DHA. These are generally deep sea bacteria such as Vibrio marinus. For ARA, microorganisms including the genera Mortierella, Entomophthora, Phytium and Porphyridium can be used for commercial production. Commercial sources of SDA include the genera Trichodesma and Echium. Commercial sources of GLA include evening primrose, black currants and borage. However, there are several disadvantages associated with commercial production of PUFAs from natural sources. Natural sources of PUFA, such as animals and plants, tend to have highly heterogeneous oil compositions. The oils obtained from these sources can require extensive purification to separate out one or more desired PUFA or to produce an oil which is enriched in one or more desired PUFA.

Natural sources also are subject to uncontrollable fluctuations in availability. Fish stocks may undergo natural variation or may be depleted by overfishing. Animal oils, and particularly fish oils, can accumulate environmental pollutants. Weather and disease can cause fluctuation in yields from both fish and plant sources. Cropland available for production of alternate oil-producing crops is subject to competition from the steady expansion of human populations and the associated increased need for food production on the remaining arable land. Crops which do produce PUFAs, such as borage, have not been adapted to commercial growth and may not perform well in monoculture. Growth of such crops is thus not economically competitive where more profitable and better established crops can be grown. Large -scale fermentation of organisms such as *Shewanella* also is expensive. Natural animal tissues contain low amounts of ARA and are difficult to process. Microorganisms such as *Porphyridium* and *Shewanella* are difficult to cultivate on a commercial scale.

Dietary supplements and pharmaceutical formulations containing PUFAs can retain the disadvantages of the PUFA source. Supplements such as fish oil capsules can

contain low levels of the particular desired component and thus require large dosages. High dosages result in ingestion of high levels of undesired components, including contaminants. Care must be taken in providing fatty acid supplements, as overaddition may result in suppression of endogenous biosynthetic pathways and lead to competition with other necessary fatty acids in various lipid fractions *in vivo*, leading to undesirable results. For example, Eskimos having a diet high in ω3 fatty acids have an increased tendency to bleed (U.S. Pat. No. 4,874,603). Fish oils have unpleasant tastes and odors, which may be impossible to economically separate from the desired product, such as a food supplements. Unpleasant tastes and odors of the supplements can make such regimens involving the supplement undesirable and may inhibit compliance by the patient.

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A number of enzymes have been identified as being involved in PUFA biosynthesis. Linoleic acid (LA, 18:2 \triangle 9, 12) is produced from oleic acid (18:1 \triangle 9) by a \triangle 12-desaturase. GLA (18:3 \triangle 6, 9, 12) is produced from linoleic acid (LA, 18:2 \triangle 9, 12) by a \triangle 6-desaturase. ARA (20:4 \triangle 5, 8, 11, 14) is produced from DGLA (20:3 \triangle 8, 11, 14), catalyzed by a \triangle 5-desaturase. Eicosapentenoic acid (EPA) is a 20 carbon, omega 3 fatty acid containing 5 double bonds (\triangle 5, 8, 11, 14, 17), all in the *cis* configuration. EPA, and the related DHA (\triangle 4, 7, 10, 13, 16, 19, C22:6) are produced from oleic acid by a series of elongation and desaturation reactions. Additionally, an elongase (or elongases) is required to extend the 18 carbon PUFAs out to 20 and 22 carbon chain lengths. However, animals cannot convert oleic acid (18:1 \triangle 9) into linoleic acid (18:2 \triangle 9, 12). Likewise, μ -linolenic acid (ALA, 18:3 \triangle 9, 12, 15) cannot be synthesized by mammals. Other eukaryotes, including fungi and plants, have enzymes which desaturate at positions \triangle 12 and \triangle 15. The major poly-unsaturated fatty acids of animals therefore are either derived from diet and/or from desaturation and elongation of linoleic acid (18:2 \triangle 9, 12) or μ -linolenic acid (18:3 \triangle 9, 12, 15).

Poly-unsaturated fatty acids are considered to be useful for nutritional, pharmaceutical, industrial, and other purposes. An expansive supply of poly-unsaturated fatty acids from natural sources and from chemical synthesis are not sufficient for commercial needs. Because a number of separate desaturase and elongase enzymes are required for fatty acid synthesis from linoleic acid (LA, 18:2 \triangle 9, 12), common in most plant species, to the more saturated and longer chain PUFAs, engineering plant host cells for the expression of EPA and DHA may require expression of five or six separate

enzyme activities to achieve expression, at least for EPA and DHA, and for production of quantities of such PUFAs additional engineering efforts may be required, for instance the down regulation of enzymes competing for substrate, engineering of higher enzyme activities such as by mutagenesis or targeting of enzymes to plastid organelles. Therefore it is of interest to obtain genetic material involved in PUFA biosynthesis from species that naturally produce these fatty acids and to express the isolated material alone or in combination in a heterologous system which can be manipulated to allow production of commercial quantities of PUFAs.

10 Relevant Literature

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Several genera of marine bacteria have been identified which synthesize either EPA or DHA (DeLong and Yayanos, Applied and Environmental Microbiology (1986) 51: 730-737). Researchers of the Sagami Chemical Research Institute have reported EPA production in E. coli which have been transformed with a gene cluster from the marine bacterium, Shewanella putrefaciens. A minimum of 5 open reading frames (ORFs) are required for fatty acid synthesis of EPA in E. coli. To date, extensive characterization of the functions of the proteins encoded by these genes has not been reported (Yazawa (1996) Lipids 31, S-297; WO 93/23545; WO 96/21735).

The protein sequence of open reading frame (ORF) 3 as published by Yazawa, USPN 5,683,898 is not a functional protein. Yazawa defines the protein as initiating at the methionine codon at nucleotides 9016-9014 of the *Shewanella* PKS-like cluster (Genbank accession U73935) and ending at the stop codon at nucleotides 8185-8183 of the *Shewanella* PKS-like cluster. However, when this ORF is expressed under control of a heterologous promoter in an *E. coli* strain containing the entire PKS-like cluster except ORF 3, the recombinant cells do not produce EPA.

Polyketides are secondary metabolites the synthesis of which involves a set of enzymatic reactions analogous to those of fatty acid synthesis (see reviews: Hopwood and Sherman, Annu. Rev. Genet. (1990) 24: 37-66, and Katz and Donadio, in Annual Review of Microbiology (1993) 47: 875-912). It has been proposed to use polyketide synthases to produce novel antibiotics (Hutchinson and Fujii, Annual Review of Microbiology (1995) 49:201-238).

SUMMARY OF THE INVENTION

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Novel compositions and methods are provided for preparation of long chain polyunsaturated fatty acids (PUFAs) using polyketide-like synthesis (PKS-like) genes in plants and plant cells. In contrast to the known and proposed methods for production of PUFAs by means of fatty acid synthesis genes, by the invention constructs and methods are provided for producing PUFAs by utilizing genes of a PKS-like system. The methods involve growing a host cell of interest transformed with an expression cassette functional in the host cell, the expression cassette comprising a transcriptional and translational initiation regulatory region, joined in reading frame 5' to a DNA sequence to a gene or component of a PKS-like system capable of modulating the production of PUFAs (PKSlike gene). An alteration in the PUFA profile of host cells is achieved by expression following introduction of a complete PKS-like system responsible for a PUFA biosynthesis into host cells. The invention finds use for example in the large scale production of DHA and EPA and for modification of the fatty acid profile of host cells and edible plant tissues and/or plant parts.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 provides designations for the ORFs of the EPA gene cluster of Shewanella. Figure 1A shows the organization of the genes; those ORFs essential for EPA production in E. coli are numbered. Figure 1B shows the designations given to subclones.

Figure 2 provides the *Shewanella* PKS-like domain structure, motifs and 'Blast' matches of ORF 6 (Figure 2A), ORF 7 (Figure 2B), ORF 8 (Figure 2C), ORF 9 (Figure 2D) and ORF 3 (Figure 2E). Figure 2F shows the structure of the region of the Anabeana chromosome that is related to domains present in *Shewanella* EPA ORFs.

Figure 3 shows results for pantethenylation - ORF 3 in E. coli strain SJ16.

Figure 4 is the sequence for the PKS-like cluster found in Shewanella, containing ORFs 3, 4, 5, 6, 7, 8 and 9. The start and last codons for each ORF are as follows:

ORF3 (published-inactive): 9016, 8186; ORF3 (active in EPA synthesis): 9157, 8186;

ORF 6: 13906, 22173; ORF 7: 22203, 24515; ORF 8: 24518, 30529; ORF 9: 30730, 32358.

Figure 5 shows the sequence for the PKS-like cluster in an approximately 40 kb DNA fragment of *Vibrio marinus*, containing ORFs 6, 7, 8 and 9. The start and last condons for each ORF are as follows: ORF 6: 17394, 25352; ORF 7: 25509, 28160; ORF 8: 28209, 34265; ORF 9: 34454, 36118.

Figure 6 shows the sequence for an approximately 19 kb portion of the PKS-like cluster of Figure 5 which contains the ORFs 6, 7, 8 and 9. The start and last condons for each ORF are as follows: ORF 6: 411, 8369; ORF 7: 8526, 11177; ORF 8: 11226, 17282; ORF 9: 17471, 19135.

Figure 7 shows a comparison of the PKS-like gene clusters of *Shewanella* putrefaciens and *Vibrio marinus*; Figure 7B is the *Vibrio marinus* operon sequence.

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Figure 8 is an expanded view of the PKS-like gene cluster portion of *Vibrio* marinus shown in Figure 7B showing that ORFs 6, 7 and 8 are in reading frame 2, while ORF 9 is in reading frame 3.

Figure 9 demonstrates sequence homology of ORF 6 of Shewanella putrefaciens and Vibrio marinus. The Shewanella ORF 6 is depicted on the vertical axis, and the Vibrio ORF 6 is depicted on the horizontal axis. Lines indicate regions of the proteins that have a 60% identity. The repeated lines in the middle correspond to the multiple ACP domains found in ORF 6.

Figure 10 demonstrates sequence homology of ORF 7 of Shewanella putrefaciens and Vibrio marinus. The Shewanella ORF 7 is depicted on the vertical axis, and the Vibrio ORF 7 is depicted on the horizontal axis. Lines indicate regions of the proteins that have a 60% identity.

Figure 11 demonstrates sequence homology of ORF 8 of Shewanella putrefaciens and Vibrio marinus. The Shewanella ORF 8 is depicted on the vertical axis, and the Vibro. ORF 8 is depicted on the horizontal axis. Lines indicate regions of the proteins that have a 60% identity.

Figure 12 demonstrates sequence homology of ORF 9 of Shewanella putrefaciens and Vibrio marinus. The Shewanella ORF 9 is depicted on the vertical axis, and the Vibrio ORF 9 is depicted on the horizontal axis. Lines indicate regions of the proteins that have a 60% identity.

Figure 13 is a depiction of various complementation experiments, and resulting PUFA production. On the right, is shown the longest PUFA made in the *E. coli* strain

containing the Vibrio and Shewanella genes depicted on the left. The hollow boxes indicate ORFs from Shewanella. The solid boxes indicate ORFs from Vibrio.

Figure 14 is a chromatogram showing fatty acid production from complementation of pEPAD8 from *Shewanella* (deletion ORF 8) with ORF 8 from *Shewanella*, in *E. coli* Fad E-. The chromatogram presents an EPA (20:5) peak.

Figure 15 is a chromatogram showing fatty acid production from complementation of pEPAD8 from *Shewanella* (deletion ORF 8) with ORF 8 from *Vibrio marinus*, in *E. coli* Fad E-. The chromatograph presents EPA (20:5) and DHA (22:6) peaks.

Figure 16 is a table of PUFA values from the ORF 8 complementation experiment, the chromatogram of which is shown in Figure 15.

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Figure 17 is a plasmid map showing the elements of pCGN7770.

Figure 18 is a plasmid map showing the elements of pCGN8535.

Figure 19 is a plasmid map showing the elements of pCGN8537.

Figure 20 is a plasmid map showing the elements of pCGN8525.

Figure 21 is a comparison of the *Shewanella* ORFs as defined by Yazawa and those disclosed in Figure 4. When a protein starting at the leucine (TTG) codon at nucleotides 9157-9155 and ending at the stop codon at nucleotides 8185-8183 is expressed under control of a heterologous promoter in an *E. coli* strain containing the entire PKS-like cluster except ORF 3, the recombinant cells do produce EPA. Thus, the published protein sequence is likely to be wrong, and the coding sequence for the protein may start at the TTG codon at nucleotides 9157-9155 or the TTG codon at nucleotides 9172-9170. This information is critical to the expression of a functional PKS-like cluster heterologous system.

Figure 22 is a plasmid map showing the elements of pCGN8560.

Figure 23 is plasmid map showing the elements of pCGN8556.

Figure 24 shows the translated DNA sequence upstream of the published ORF 3. The ATG start codon at position 9016 is the start codon for the protein described by Yazawa et al (1996) supra. The other arrows depict TTG or ATT codons that can also serve as start codons in bacteria. When ORF 3 is started from the published ATG codon at 9016, the protein is not functional in making EPA. When ORF 3 is initiated at the TTG codon at position 9157, the protein is capable of facilitating EPA synthesis.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

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In accordance with the subject invention, novel DNA sequences, DNA constructs and methods are provided, which include some or all of the polyketide-like synthesis (PKS-like) pathway genes from Shewanella, Vibrio or other microorganisms, for modifying the poly-unsaturated long chain fatty acid content of host cells, particularly host plant cells. The present invention demonstrates that EPA synthesis genes in Shewanella putrefaciens constitute a polyketide-like synthesis pathway. Functions are ascribed to the Shewanella and Vibrio genes and methods are provided for the production of EPA and DHA in host cells. The method includes the step of transforming cells with an expression cassette comprising a DNA encoding a polypeptide capable of increasing the amount of one or more PUFA in the host cell. Desirably, integration constructs are prepared which provide for integration of the expression cassette into the genome of a host cell. Host cells are manipulated to express a sense or antisense DNA encoding a polypeptide(s) that has PKS-like gene activity. By "PKS-like gene" is intended a polypeptide which is responsible for any one or more of the functions of a PKS-like activity of interest. By "polypeptide" is meant any chain of amino acids, regardless of length or post-translational modification, for example, glycosylation or phosphorylation. Depending upon the nature of the host cell, the substrate(s) for the expressed enzyme may be produced by the host cell or may be exogenously supplied. Of particular interest is the selective control of PUFA production in plant tissues and/or plant parts such as leaves, roots, fruits and seeds. The invention can be used to synthesize EPA, DHA, and other related PUFAs in host cells.

There are many advantages to transgenic production of PUFAs. As an example, in transgenic *E. coli* as in *Shewanella*, EPA accumulates in the phospholipid fraction, specifically in the *sn*-2 position. It may be possible to produce a structured lipid in a desired host cell which differs substantially from that produced in either *Shewanella* or *E. coli*. Additionally transgenic production of PUFAs in particular host cells offers several advantages over purification from natural sources such as fish or plants. In transgenic plants, by utilizing a PKS-like system, fatty acid synthesis of PUFAs is achieved in the cytoplasm by a system which produces the PUFAs through *de novo* production of the fatty acids utilizing malonyl Co-A and acetyl Co-A as substrates. In this fashion, potential problems, such as those associated with substrate competition and diversion of normal products of fatty acid synthesis in a host to PUFA production, are avoided.

Production of fatty acids from recombinant plants provides the ability to alter the naturally occurring plant fatty acid profile by providing new synthetic pathways in the host or by suppressing undesired pathways, thereby increasing levels of desired PUFAs, or conjugated forms thereof, and decreasing levels of undesired PUFAs. Production of fatty acids in transgenic plants also offers the advantage that expression of PKS-like genes in particular tissues and/or plant parts means that greatly increased levels of desired PUFAs in those tissues and/or parts can be achieved, making recovery from those tissues more economical. Expression in a plant tissue and/or plant part presents certain efficiencies, particularly where the tissue or part is one which is easily harvested, such as seed, leaves, fruits, flowers, roots, etc. For example, the desired PUFAs can be expressed in seed; methods of isolating seed oils are well established. In addition to providing a source for purification of desired PUFAs, seed oil components can be manipulated through expression of PKS-like genes, either alone or in combination with other genes such as elongases, to provide seed oils having a particular PUFA profile in concentrated form. The concentrated seed oils then can be added to animal milks and/or synthetic or semisynthetic milks to serve as infant formulas where human nursing is impossible or undesired, or in cases of malnourishment or disease in both adults and infants.

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Transgenic microbial production of fatty acids offers the advantages that many microbes are known with greatly simplified oil compositions as compared with those of higher organisms, making purification of desired components easier. Microbial production is not subject to fluctuations caused by external variables such as weather and food supply. Microbially produced oil is substantially free of contamination by environmental pollutants. Additionally, microbes can provide PUFAs in particular forms which may have specific uses. For example, Spirulina can provide PUFAs predominantly at the first and third positions of triglycerides; digestion by pancreatic lipases preferentially releases fatty acids from these positions. Following human or animal ingestion of triglycerides derived from Spirulina, thes PUFAs are released by pancreatic lipases as free fatty acids and thus are directly available, for example, for infant brain development. Additionally, microbial oil production can be manipulated by controlling culture conditions, notably by providing particular substrates for microbially expressed enzymes, or by addition of compounds which suppress undesired biochemical pathways. In addition to these advantages, production of fatty acids from recombinant microbes provides the ability to alter the naturally occurring microbial fatty acid profile by

providing new synthetic pathways in the host or by suppressing undesired pathways, thereby increasing levels of desired PUFAs, or conjugated forms thereof, and decreasing levels of undesired PUFAs.

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Production of fatty acids in animals also presents several advantages. Expression of desaturase genes in animals can produce greatly increased levels of desired PUFAs in animal tissues, making recovery from those tissues more economical. For example, where the desired PUFAs are expressed in the breast milk of animals, methods of isolating PUFAs from animal milk are well established. In addition to providing a source for purification of desired PUFAs, animal breast milk can be manipulated through expression of desaturase genes, either alone or in combination with other human genes, to provide animal milks with a PUFA composition substantially similar to human breast milk during the different stages of infant development. Humanized animal milks could serve as infant formulas where human nursing is impossible or undesired, or in the cases of malnourishment or disease.

DNAs encoding desired PKS-like genes can be identified in a variety of ways. In one method, a source of a desired PKS-like gene, for example genomic libraries from a Shewanella or Vibrio spp., is screened with detectable enzymatically- or chemicallysynthesized probes. Sources of ORFs having PKS-like genes are those organisms which produce a desired PUFA, including DHA-producing or EPA-producing deep sea bacteria growing preferentially under high pressure or at relatively low temperature. Microorgansims such as Shewanella which produce EPA or DHA also can be used as a source of PKS-like genes. The probes can be made from DNA, RNA, or non-naturally occurring nucleotides, or mixtures thereof. Probes can be enzymatically synthesized from DNAs of known PKS-like genes for normal or reduced-stringency hybridization methods. For discussions of nucleic acid probe design and annealing conditions, see, for example, Sambrook et al, Molecular Cloning: A Laboratory Manual (2nd ed.), Vols. 1-3, Cold Spring Harbor Laboratory, (1989) or Current Protocols in Molecular Biology, F. Ausubel et al, ed., Greene Publishing and Wiley-Interscience, New York (1987), each of which is incorporated herein by reference. Techniques for manipulation of nucleic acids encoding PUFA enzymes such as subcloning nucleic acid sequences encoding polypeptides into expression vectors, labelling probes, DNA hybridization, and the like are described generally in Sambrook, supra.

Oligonucleotide probes also can be used to screen sources and can be based on sequences of known PKS-like genes, including sequences conserved among known PKS-like genes, or on peptide sequences obtained from a desired purified protein.

Oligonucleotide probes based on amino acid sequences can be degenerate to encompass the degeneracy of the genetic code, or can be biased in favor of the preferred codons of the source organism. Alternatively, a desired protein can be entirely sequenced and total synthesis of a DNA encoding that polypeptide performed.

Once the desired DNA has been isolated, it can be sequenced by known methods. It is recognized in the art that such methods are subject to errors, such that multiple sequencing of the same region is routine and is still expected to lead to measurable rates of mistakes in the resulting deduced sequence, particularly in regions having repeated domains, extensive secondary structure, or unusual base compositions, such as regions with high GC base content. When discrepancies arise, resequencing can be done and can employ special methods. Special methods can include altering sequencing conditions by using: different temperatures; different enzymes; proteins which alter the ability of oligonucleotides to form higher order structures; altered nucleotides such as ITP or methylated dGTP; different gel compositions, for example adding formamide; different primers or primers located at different distances from the problem region; or different templates such as single stranded DNAs. Sequencing of mRNA can also be employed.

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For the most part, some or all of the coding sequences for the polypeptides having PKS-like gene activity are from a natural source. In some situations, however, it is desirable to modify all or a portion of the codons, for example, to enhance expression, by employing host preferred codons. Host preferred codons can be determined from the codons of highest frequency in the proteins expressed in the largest amount in a particular host species of interest. Thus, the coding sequence for a polypeptide having PKS-like gene activity can be synthesized in whole or in part. All or portions of the DNA also can be synthesized to remove any destabilizing sequences or regions of secondary structure which would be present in the transcribed mRNA. All or portions of the DNA also can be synthesized to alter the base composition to one more preferable to the desired host cell. Methods for synthesizing sequences and bringing sequences together are well established in the literature. *In vitro* mutagenesis and selection, site-directed mutagenesis, or other means can be employed to obtain mutations of naturally occurring PKS-like genes to produce a polypeptide having PKS-like gene activity *in vivo* with more desirable

physical and kinetic parameters for function in the host cell, such as a longer half-life or a higher rate of production of a desired polyunsaturated fatty acid.

Of particular interest are the Shewanella putrefaciens ORFs and the corresponding ORFs of Vibrio marinus. The Shewanella putrefaciens PKS-like genes can be expressed in transgenic plants to effect biosynthesis of EPA. Other DNAs which are substantially identical in sequence to the Shewanella putrefaciens PKS-like genes, or which encode polypeptides which are substantially similar to PKS-like genes of Shewanella putrefaciens can be used, such as those identified from Vibrio marinus. By substantially identical in sequence is intended an amino acid sequence or nucleic acid sequence exhibiting in order of increasing preference at least 60%, 80%, 90% or 95% homology to the DNA sequence of the Shewanella putrefaciens PKS-like genes or nucleic acid sequences encoding the amino acid sequences for such genes. For polypeptides, the length of comparison sequences generally is at least 16 amino acids, preferably at least 20 amino acids, and most preferably 35 amino acids. For nucleic acids, the length of comparison sequences generally is at least 50 nucleotides, preferably at least 60 nucleotides, and more preferably at least 75 nucleotides, and most preferably, 110 nucleotides.

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Homology typically is measured using sequence analysis software, for example, the Sequence Analysis software package of the Genetics Computer Group, University of Wisconsin Biotechnology Center, 1710 University Avenue, Madison, Wisconsin 53705, MEGAlign (DNAStar, Inc., 1228 S. Park St., Madison, Wisconsin 53715), and MacVector (Oxford Molecular Group, 2105 S. Bascom Avenue, Suite 200, Campbell, California 95008). BLAST (National Center for Biotechnology Information (WCBI) www.ncbi.nlm.gov; FASTA (Pearson and Lipman, *Science* (1985) 227:1435-1446). Such software matches similar sequences by assigning degrees of homology to various substitutions, deletions, and other modifications. Conservative substitutions typically include substitutions within the following groups: glycine and alanine; valine, isoleucine and leucine; aspartic acid, glutamic acid, asparagine, and glutamine; serine and threonine; lysine and arginine; and phenylalanine and tyrosine. Substitutions may also be made on the basis of conserved hydrophobicity or hydrophilicity (Kyte and Doolittle, *J. Mol. Biol.* (1982) 157: 105-132), or on the basis of the ability to assume similar polypeptide secondary structure (Chou and Fasman, *Adv. Enzymol.* (1978) 47: 45-148, 1978). A

related protein to the probing sequence is identified when $p \ge 0.01$, preferably $p \ge 10^{-7}$ or 10^{-8} .

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Encompassed by the present invention are related PKS-like genes from the same or other organisms. Such related PKS-like genes include variants of the disclosed PKSlike ORFs that occur naturally within the same or different species of Shewanella, as well as homologues of the disclosed PKS-like genes from other species and evolutionarily related proteins having analogous function and activity. Also included are PKS-like genes which, although not substantially identical to the Shewanella putrefaciens PKSlike genes, operate in a similar fashion to produce PUFAs as part of a PKS-like system. Related PKS-like genes can be identified by their ability to function substantially the same as the disclosed PKS-like genes; that is, they can be substituted for corresponding ORFs of Shewanella or Vibrio and still effectively produce EPA or DHA. Related PKSlike genes also can be identified by screening sequence databases for sequences homologous to the disclosed PKS-like genes, by hybridization of a probe based on the disclosed PKS-like genes to a library constructed from the source organism, or by RT-PCR using mRNA from the source organism and primers based on the disclosed PKS-like gene. Thus, the phrase "PKS-like genes" refers not only to the nucleotide sequences disclosed herein, but also to other nucleic acids that are allelic or species variants of these nucleotide sequences. It is also understood that these terms include nonnatural mutations introduced by deliberate mutation using recombinant technology such as single site mutation or by excising short sections of DNA open reading frames coding for PUFA enzymes or by substituting new codons or adding new codons. Such minor alterations substantially maintain the immunoidentity of the original expression product and/or its biological activity. The biological properties of the altered PUFA enzymes can be determined by expressing the enzymes in an appropriate cell line and by determining the ability of the enzymes to synthesize PUFAs. Particular enzyme modifications considered minor would include substitution of amino acids of similar chemical properties, e.g., glutamic acid for aspartic acid or glutamine for asparagine.

When utilizing a PUFA PKS-like system from another organism, the regions of a PKS-like gene polypeptide important for PKS-like gene activity can be determined through routine mutagenesis, expression of the resulting mutant polypeptides and determination of their activities. The coding region for the mutants can include deletions, insertions and point mutations, or combinations thereof. A typical functional analysis

begins with deletion mutagenesis to determine the N- and C-terminal limits of the protein necessary for function, and then internal deletions, insertions or point mutants are made in the open ready frame to further determine regions necessary for function. Other techniques such as cassette mutagenesis or total synthesis also can be used. Deletion mutagenesis is accomplished, for example, by using exonucleases to sequentially remove the 5' or 3' coding regions. Kits are available for such techniques. After deletion, the coding region is completed by ligating oligonucleotides containing start or stop codons to the deleted coding region after 5' or 3' deletion, respectively. Alternatively, oligonucleotides encoding start or stop codons are inserted into the coding region by a variety of methods including site-directed mutagenesis, mutagenic PCR or by ligation onto DNA digested at existing restriction sites. Internal deletions can similarly be made through a variety of methods including the use of existing restriction sites in the DNA, by use of mutagenic primers via site directed mutagenesis or mutagenic PCR. Insertions are made through methods such as linker-scanning mutagenesis, site-directed mutagenesis or mutagenic PCR. Point mutations are made through techniques such as site-directed mutagenesis or mutagenic PCR.

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Chemical mutagenesis also can be used for identifying regions of a PKS-like gene polypeptide important for activity. A mutated construct is expressed, and the ability of the resulting altered protein to function as a PKS-like gene is assayed. Such structurefunction analysis can determine which regions may be deleted, which regions tolerate insertions, and which point mutations allow the mutant protein to function in substantially the same way as the native PKS-like gene. All such mutant proteins and nucleotide sequences encoding them are within the scope of the present invention. EPA is produced in Shewanella as the product of a PKS-like system, such that the EPA genes encode components of this system. In Vibrio, DHA is produced by a similar system. The enzymes which synthesize these fatty acids are encoded by a cluster of genes which are distinct from the fatty acid synthesis genes encoding the enzymes involved in synthesis of the C16 and C18 fatty acids typically found in bacteria and in plants. As the Shewanella EPA genes represent a PKS-like gene cluster, EPA production is, at least to some extent, independent of the typical bacterial type II FAS system. Thus, production of EPA in the cytoplasm of plant cells can be achieved by expression of the PKS-like pathway genes in plant cells under the control of appropriate plant regulatory signals.

EPA production in *E. coli* transformed with the *Shewanella* EPA genes proceeds during anaerobic growth, indicating that O2-dependent desaturase reactions are not involved. Analyses of the proteins encoded by the ORFs essential for EPA production reveals the presence of domain structures characteristic of PKS-like systems. Fig. 2A shows a summary of the domains, motifs, and also key homologies detected by "BLAST" data bank searches. Because EPA is different from many of the other substances produced by PKS-like pathways, i.e., it contains 5, *cis* double bonds, spaced at 3 carbon intervals along the molecule, a PKS-like system for synthesis of EPA is not expected.

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Further, BLAST searches using the domains present in the Shewanella EPA ORFs reveal that several are related to proteins encoded by a PKS-like gene cluster found in Anabeana. The structure of that region of the Anabeana chromosome is shown in Fig. 2F. The Anabeana PKS-like genes have been linked to the synthesis of a long-chain (C26), hydroxy-fatty acid found in a glycolipid layer of heterocysts. The EPA protein domains with homology to the Anabeana proteins are indicated in Fig. 2F.

ORF 6 of Shewanella contains a KAS domain which includes an active site motif (DXAC*) as well as a "GFGG" motif which is present at the end of many Type II KAS proteins (see Fig. 2A). Extended motifs are present but not shown here. Next is a malonyl-CoA:ACP acyl transferase (AT) domain. Sequences near the active site motif (GHS*XG) suggest it transfers malonate rather than methylmalonate, i.e., it resembles the acetate-like ATs. Following a linker region, there is a cluster of 6 repeating domains, each ~100 amino acids in length, which are homologous to PKS-like ACP sequences. Each contains a pantetheine binding site motif (LGXDS*(L/I)). The presence of 6 such ACP domains has not been observed previously in fatty acid synthases (FAS) or PKS-like systems. Near the end of the protein is a region which shows homology to β-keto-ACP reductases (KR). It contains a pyridine nucleotide binding site motif "GXGXX(G/A/P)".

The Shewanella ORF 8 begins with a KAS domain, including active site and ending motifs (Fig. 2C). The best match in the data banks is with the Anabeana HglD. There is also a domain which has sequence homology to the N- terminal one half of the Anabeana HglC. This region also shows weak homology to KAS proteins although it lacks the active site and ending motifs. It has the characteristics of the so-called chain length factors (CLF) of Type II PKS-like systems. ORF 8 appears to direct the production of EPA versus DHA by the PKS-like system. ORF 8 also has two domains with homology to β-hydroxyacyl-ACP dehydrases (DH). The best match for both domains is

with *E. coli* FabA, a bi-functional enzyme which carries out both the dehydrase reaction and an isomerization (*trans* to *cis*) of the resulting double bond. The first DH domain contains both the active site histidine (H) and an adjacent cysteine (C) implicated in FabA catalysis. The second DH domain has the active site H but lacks the adjacent C (Fig. 2C). Blast searches with the second DH domain also show matches to FabZ, a second *E. coli* DH, which does not possess isomerase activity.

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The N-terminal half of ORF 7 (Fig. 2B) has no significant matches in the data banks. The best match of the C-terminal half is with a C-terminal portion of the Anabeana HglC. This domain contains an acyl-transferase (AT) motif (GXSXG). Comparison of the extended active site sequences, based on the crystal structure of the *E. coli* malonyl-CoA:ACP AT, reveals that ORF 7 lacks two residues essential for exclusion of water from the active site (*E. coli* nomenclature; Q11 and R117). These data suggest that ORF 7 may function as a thioesterase.

ORF 9 (Fig. 2D) is homologous to an ORF of unknown function in the Anabeana Hgl cluster. It also exhibits a very weak homology to NIFA, a regulatory protein in nitrogen fixing bacteria. A regulatory role for the ORF 9 protein has not been excluded. ORF 3 (Fig. 2E) is homologous to the Anabeana Hetl as well as EntD from *E. coli* and Sfp of *Bacillus*. Recently, a new enzyme family of phosphopantetheinyl transferases has been identified that includes Hetl, EntD and Sfp (Lamblot RH, *et al.* (1996) A new enzyme superfamily - the phophopantetheinyl transferases. *Chemistry & Biology*, Vol 3, #11, 923-936). The data of Fig. 3 demonstrates that the presence of ORF 3 is required for addition of \(\beta\)-alanine (i.e. pantetheine) to the ORF 6 protein. Thus, ORF 3 encodes the phosphopantetheinyl transferase specific for the ORF 6 ACP domains. (*See*, Haydock SF *et al.* (1995) Divergent sequence motifs correlated with the substrate specificity of (methyl)malonyl-CoA:acyl carrier protein transacylase domains in modular polyketide synthases, *FEBS Lett.*, 374, 246-248). Malonate is the source of the carbons utilized in the extension reactions of EPA synthesis. Additionally, malonyl-CoA rather than malonyl-ACP is the AT substrate, i.e., the AT region of ORF 6 uses malonyl Co-A.

Once the DNA sequences encoding the PKS-like genes of an organism responsible for PUFA production have been obtained, they are placed in a vector capable of replication in a host cell, or propagated *in vitro* by means of techniques such as PCR or long PCR. Replicating vectors can include plasmids, phage, viruses, cosmids and the like. Desirable vectors include those useful for mutagenesis of the gene of interest or for

expression of the gene of interest in host cells. A PUFA synthesis enzyme or a homologous protein can be expressed in a variety of recombinantly engineered cells. Numerous expression systems are available for expression of DNA encoding a PUFA enzyme. The expression of natural or synthetic nucleic acids encoding PUFA enzyme is typically achieved by operably linking the DNA to a promoter (which is either constitutive or inducible) within an expression vector. By expression vector is meant a DNA molecule, linear or circular, that comprises a segment encoding a PUFA enzyme, operably linked to additional segments that provide for its transcription. Such additional segments include promoter and terminator sequences. An expression vector also may include one or more origins of replication, one or more selectable markers, an enhancer, a polyadenylation signal, etc. Expression vectors generally are derived from plasmid or viral DNA, and can contain elements of both. The term "operably linked" indicates that the segments are arranged so that they function in concert for their intended purposes, for example, transcription initiates in the promoter and proceeds through the coding segment to the terminator. See Sambrook et al, supra.

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The technique of long PCR has made *in vitro* propagation of large constructs possible, so that modifications to the gene of interest; such as mutagenesis or addition of expression signals, and propagation of the resulting constructs can occur entirely *in vitro* without the use of a replicating vector or a host cell. *In vitro* expression can be accomplished, for example, by placing the coding region for the desaturase polypeptide in an expression vector designed for *in vitro* use and adding rabbit reticulocyte lysate and cofactors; labeled amino acids can be incorporated if desired. Such *in vitro* expression vectors may provide some or all of the expression signals necessary in the system used. These methods are well known in the art and the components of the system are commercially available. The reaction mixture can then be assayed directly for PKS-like enzymes for example by determining their activity, or the synthesized enzyme can be purified and then assayed.

Expression in a host cell can be accomplished in a transient or stable fashion. Transient expression can occur from introduced constructs which contain expression signals functional in the host cell, but which constructs do not replicate and rarely integrate in the host cell, or where the host cell is not proliferating. Transient expression also can be accomplished by inducing the activity of a regulatable promoter operably linked to the gene of interest, although such inducible systems frequently exhibit a low

basal level of expression. Stable expression can be achieved by introduction of a nucleic acid construct that can integrate into the host genome or that autonomously replicates in the host cell. Stable expression of the gene of interest can be selected for through the use of a selectable marker located on or transfected with the expression construct, followed by selection for cells expressing the marker. When stable expression results from integration, integration of constructs can occur randomly within the host genome or can be targeted through the use of constructs containing regions of homology with the host genome sufficient to target recombination with the host locus. Where constructs are targeted to an endogenous locus, all or some of the transcriptional and translational regulatory regions can be provided by the endogenous locus. To achieve expression in a host cell, the transformed DNA is operably associated with transcriptional and translational initiation and termination regulatory regions that are functional in the host cell.

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Transcriptional and translational initiation and termination regions are derived from a variety of nonexclusive sources, including the DNA to be expressed, genes known or suspected to be capable of expression in the desired system, expression vectors, chemical synthesis The termination region can be derived from the 3' region of the gene from which the initiation region was obtained or from a different gene. A large number of termination regions are known to and have been found to be satisfactory in a variety of hosts from the same and different genera and species. The termination region usually is selected more as a matter of convenience rather than because of any particular property. When expressing more than one PKS-like ORF in the same cell, appropriate regulatory regions and expression methods should be used. Introduced genes can be propagated in the host cell through use of replicating vectors or by integration into the host genome. Where two or more genes are expressed from separate replicating vectors, it is desirable that each vector has a different means of replication. Each introduced construct, whether integrated or not, should have a different means of selection and should lack homology to the other constructs to maintain stable expression and prevent reassortment of elements among constructs. Judicious choices of regulatory regions, selection means and method of propagation of the introduced construct can be experimentally determined so that all introduced genes are expressed at the necessary levels to provide for synthesis of the desired products.

A variety of procaryotic expression systems can be used to express PUFA enzyme. Expression vectors can be constructed which contain a promoter to direct transcription, a ribosome binding site, and a transcriptional terminator. Examples of regulatory regions suitable for this purpose in E. coli are the promoter and operator region of the E. coli tryptophan biosynthetic pathway as described by Yanofsky (1984) J. Bacteriol., 158:1018-1024 and the leftward promoter of phage lambda (Pλ) as described by Herskowitz and Hagen, (1980) Ann. Rev. Genet., 14:399-445. The inclusion of selection markers in DNA vectors transformed in E.coli is also useful. Examples of such markers include genes specifying resistance to ampicillin, tetracycline, or chloramphenicol. Vectors used for expressing foreign genes in bacterial hosts generally will contain a selectable marker, such as a gene for antibiotic resistance, and a promoter which functions in the host cell. Plasmids useful for transforming bacteria include pBR322 (Bolivar, et al, (1977) Gene 2:95-113), the pUC plasmids (Messing, (1983) Meth. Enzymol. 101:20-77, Vieira and Messing, (1982) Gene 19:259-268), pCQV2 (Queen, ibid.), and derivatives thereof. Plasmids may contain both viral and bacterial elements. Methods for the recovery of the proteins in biologically active form are discussed in U.S. Patent Nos. 4.966.963 and 4.999,422, which are incorporated herein by reference. See Sambrook, et al for a description of other prokaryotic expression systems.

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For expression in eukaryotes, host cells for use in practicing the present invention include mammalian, avian, plant, insect, and fungal cells. As an example, for plants, the choice of a promoter will depend in part upon whether constitutive or inducible expression is desired and whether it is desirable to produce the PUFAs at a particular stage of plant development and/or in a particular tissue. Considerations for choosing a specific tissue and/or developmental stage for expression of the ORFs may depend on competing substrates or the ability of the host cell to tolerate expression of a particular PUFA. Expression can be targeted to a particular location within a host plant such as seed, leaves, fruits, flowers, and roots, by using specific regulatory sequences, such as those described in USPN 5,463,174, USPN 4,943,674, USPN 5,106,739, USPN 5,175,095, USPN 5,420,034, USPN 5,188,958, and USPN 5,589,379. Where the host cell is a yeast, transcription and translational regions functional in yeast cells are provided, particularly from the host species. The transcriptional initiation regulatory regions can be obtained, for example from genes in the glycolytic pathway, such as alcohol dehydrogenase, glyceraldehyde-3-phosphate dehydrogenase (GPD),

phosphoglucoisomerase, phosphoglycerate kinase, etc. or regulatable genes such as acid phosphatase, lactase, metallothionein, glucoamylase, etc. Any one of a number of regulatory sequences can be used in a particular situation, depending upon whether constitutive or induced transcription is desired, the particular efficiency of the promoter in conjunction with the open-reading frame of interest, the ability to join a strong promoter with a control region from a different promoter which allows for inducible transcription, ease of construction, and the like. Of particular interest are promoters which are activated in the presence of galactose. Galactose-inducible promoters (GAL1, GAL7, and GAL10) have been extensively utilized for high level and regulated expression of protein in yeast (Lue et al. (1987) Mol. Cell. Biol. 7:3446; Johnston, (1987) Microbiol. Rev. 51:458). Transcription from the GAL promoters is activated by the GAL4 protein, which binds to the promoter region and activates transcription when galactose is present. In the absence of galactose, the antagonist GAL80 binds to GAL4 and prevents GAL4 from activating transcription. Addition of galactose prevents GAL80 from inhibiting activation by GAL4. Preferably, the termination region is derived from a yeast gene, particularly Saccharomyces, Schizosaccharomyces, Candida or Kluyveromyces. The 3' regions of two mammalian genes, γ interferon and α2 interferon, are also known to function in yeast.

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Nucleotide sequences surrounding the translational initiation codon ATG have been found to affect expression in yeast cells. If the desired polypeptide is poorly expressed in yeast, the nucleotide sequences of exogenous genes can be modified to include an efficient yeast translation initiation sequence to obtain optimal gene expression. For expression in *Saccharomyces*, this can be done by site-directed mutagenesis of an inefficiently expressed gene by fusing it in-frame to an endogenous *Saccharomyces* gene, preferably a highly expressed gene, such as the lactase gene.

As an alternative to expressing the PKS-like genes in the plant cell cytoplasm, is to target the enzymes to the chloroplast. One method to target proteins to the chloroplast entails use of leader peptides attached to the N-termini of the proteins. Commonly used leader peptides are derived from the small subunit of plant ribulose bis phosphate carboxylase. Leader sequences from other chloroplast proteins may also be used. Another method for targeting proteins to the chloroplast is to transform the chloroplast genome (Stable transformation of chloroplasts of *Chlamydomonas reinhardtii* (1 green alga) using bombardment of recipient cells with high-velocity tungsten microprojectiles coated with foreign DNA has been described. *See*, for example, Blowers *et al Plant Cell*

(1989) 1:123-132 and Debuchy et al EMBO J (1989) 8:2803-2809. The transformation technique, using tungsten microprojectiles, is described by Kline et al, Nature (London) (1987) 327:70-73). The most common method of transforming chloroplasts involves using biolistic techniques, but other techniques developed for the purpose may also be used. (Methods for targeting foreign gene products into chloroplasts (Shrier et al EMBO J. (1985) 4:25-32) or mitochnodria (Boutry et al, supra) have been described. See also Tomai et al Gen. Biol. Chem. (1988) 263:15104-15109 and US Patent No. 4,940,835 for the use of transit peptides for translocating nuclear gene products into the chloroplast. Methods for directing the transport of proteins to the chloroplast are reviewed in Kenauf TIBTECH (1987) 5:40-47.

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For producing PUFAs in avian species and cells, gene transfer can be performed by introducing a nucleic acid sequence encoding a PUFA enzyme into the cells following procedures known in the art. If a transgenic animal is desired, pluripotent stem cells of embryos can be provided with a vector carrying a PUFA enzyme encoding transgene and developed into adult animal (USPN 5,162,215; Ono et al. (1996) Comparative Biochemistry and Physiology A 113(3):287-292; WO 9612793; WO 9606160). In most cases, the transgene is modified to express high levels of the PKS-like enzymes in order to increase production of PUFAs. The transgenes can be modified, for example, by providing transcriptional and/or translational regulatory regions that function in avian cells, such as promoters which direct expression in particular tissues and egg parts such as yolk. The gene regulatory regions can be obtained from a variety of sources, including chicken anemia or avian leukosis viruses or avian genes such as a chicken ovalbumin gene.

Production of PUFAs in insect cells can be conducted using baculovirus expression vectors harboring PKS-like transgenes. Baculovirus expression vectors are available from several commercial sources such as Clonetech. Methods for producing hybrid and transgenic strains of algae, such as marine algae, which contain and express a desaturase transgene also are provided. For example, transgenic marine algae can be prepared as described in USPN 5,426,040. As with the other expression systems described above, the timing, extent of expression and activity of the desaturase transgene can be regulated by fitting the polypeptide coding sequence with the appropriate transcriptional and translational regulatory regions selected for a particular use. Of particular interest are promoter regions which can be induced under preselected growth

conditions. For example, introduction of temperature sensitive and/or metabolite responsive mutations into the desaturase transgene coding sequences, its regulatory regions, and/or the genome of cells into which the transgene is introduced can be used for this purpose.

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The transformed host cell is grown under appropriate conditions adapted for a desired end result. For host cells grown in culture, the conditions are typically optimized to produce the greatest or most economical yield of PUFAs, which relates to the selected desaturase activity. Media conditions which may be optimized include: carbon source, nitrogen source, addition of substrate, final concentration of added substrate, form of substrate added, aerobic or anaerobic growth, growth temperature, inducing agent, induction temperature, growth phase at induction, growth phase at harvest, pH, density, and maintenance of selection. Microorganisms such as yeast, for example, are preferably grown using selected media of interest, which include yeast peptone broth (YPD) and minimal media (contains amino acids, yeast nitrogen base, and ammonium sulfate, and lacks a component for selection, for example uracil). Desirably, substrates to be added are first dissolved in ethanol. Where necessary, expression of the polypeptide of interest may be induced, for example by including or adding galactose to induce expression from a GAL promoter.

When increased expression of the PKS-like gene polypeptide in a host cell which expresses PUFA from a PKS-like system is desired, several methods can be employed. Additional genes encoding the PKS-like gene polypeptide can be introduced into the host organism. Expression from the native PKS-like gene locus also can be increased through homologous recombination, for example by inserting a stronger promoter into the host genome to cause increased expression, by removing destabilizing sequences from either the mRNA or the encoded protein by deleting that information from the host genome, or by adding stabilizing sequences to the mRNA (*see* USPN 4,910,141 and USPN 5,500,365). Thus, the subject host will have at least have one copy of the expression construct and may have two or more, depending upon whether the gene is integrated into the genome, amplified, or is present on an extrachromosomal element having multiple copy numbers. Where the subject host is a yeast, four principal types of yeast plasmid vectors can be used: Yeast Integrating plasmids (YIps), Yeast Replicating plasmids (YRps), Yeast Centromere plasmids (YCps), and Yeast Episomal plasmids (YEps). YIps lack a yeast replication origin and must be propagated as integrated elements in the yeast

genome. YRps have a chromosomally derived autonomously replicating sequence and are propagated as medium copy number (20 to 40), autonomously replicating, unstably segregating plasmids. YCps have both a replication origin and a centromere sequence and propagate as low copy number (10-20), autonomously replicating, stably segregating plasmids. YEps have an origin of replication from the yeast 2µm plasmid and are propagated as high copy number, autonomously replicating, irregularly segregating plasmids. The presence of the plasmids in yeast can be ensured by maintaining selection for a marker on the plasmid. Of particular interest are the yeast vectors pYES2 (a YEp plasmid available from Invitrogen, confers uracil prototrophy and a GAL1 galactose-inducible promoter for expression), and pYX424 (a YEp plasmid having a constitutive TP1 promoter and conferring leucine prototrophy; (Alber and Kawasaki (1982). *J. Mol. & Appl. Genetics* 1: 419).

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The choice of a host cell is influenced in part by the desired PUFA profile of the transgenic cell, and the native profile of the host cell. Even where the host cell expresses PKS-like gene activity for one PUFA, expression of PKS-like genes of another PKS-like system can provide for production of a novel PUFA not produced by the host cell. In particular instances where expression of PKS-like gene activity is coupled with expression of an ORF 8 PKS-like gene of an organism which produces a different PUFA, it can be desirable that the host cell naturally have, or be mutated to have, low PKS-like gene activity for ORF 8. As an example, for production of EPA, the DNA sequence used encodes the polypeptide having PKS-like gene activity of an organism which produces EPA, while for production of DHA, the DNA sequences used are those from an organism which produces DHA. For use in a host cell which already expresses PKS-like gene activity it can be necessary to utilize an expression cassette which provides for overexpression of the desired PKS-like genes alone or with a construct to downregulate the activity of an existing ORF of the existing PKS-like system, such as by antisense or co-suppression. Similarly, a combination of ORFs derived from separate organisms which produce the same or different PUFAs using PKS-like systems may be used. For instance, the ORF 8 of Vibrio directs the expression of DHA in a host cell, even when ORFs 3, 6, 7 and 9 are from Shewanella, which produce EPA when coupled to ORF 8 of Shewanella. Therefore, for production of eicosapentanoic acid (EPA), the expression cassettes used generally include one or more cassettes which include ORFs 3, 6, 7, 8 and 9 from a PUFA-producing organism such as the marine bacterium Shewanella

putrefaciens (for EPA production) or Vibrio marinus (for DHA production). ORF 8 can be used for induction of DHA production, and ORF 8 of Vibrio can be used in conjunction with ORFs 3, 6, 7 and 9 of Shewanella to produce DHA. The organization and numbering scheme of the ORFs identified in the Shewanella gene cluster are shown in Fig 1A. Maps of several subclones referred to in this study are shown in Fig 1B. For expression of a PKS-like gene polypeptide, transcriptional and translational initiation and termination regions functional in the host cell are operably linked to the DNA encoding the PKS-like gene polypeptide.

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Constructs comprising the PKS-like ORFs of interest can be introduced into a host cell by any of a variety of standard techniques, depending in part upon the type of host cell. These techniques include transfection, infection, bolistic impact, electroporation, microinjection, scraping, or any other method which introduces the gene of interest into the host cell (see USPN 4,743,548, USPN 4,795,855, USPN 5,068,193, USPN 5,188,958, USPN 5,463,174, USPN 5,565,346 and USPN 5,565,347). Methods of transformation which are used include lithium acetate transformation (Methods in Enzymology, (1991) 194:186-187). For convenience, a host cell which has been manipulated by any method to take up a DNA sequence or construct will be referred to as "transformed" or "recombinant" herein. The subject host will have at least have one copy of the expression construct and may have two or more, depending upon whether the gene is integrated into the genome, amplified, or is present on an extrachromosomal element having multiple copy numbers.

For production of PUFAs, depending upon the host cell, the several polypeptides produced by pEPA, ORFs 3, 6, 7, 8 and 9, are introduced as individual expression constructs or can be combined into two or more cassettes which are introduced individually or co-transformed into a host cell. A standard transformation protocol is used. For plants, where less than all PKS-like genes required for PUFA synthesis have been inserted into a single plant, plants containing a complementing gene or genes can be crossed to obtain plants containing a full complement of PKS-like genes to synthesize a desired PUFA.

The PKS-like-mediated production of PUFAs can be performed in either prokaryotic or eukaryotic host cells. The cells can be cultured or formed as part or all of a host organism including an animal. Viruses and bacteriophage also can be used with appropriate cells in the production of PUFAs, particularly for gene transfer, cellular

targeting and selection. Any type of plant cell can be used for host cells, including dicotyledonous plants, monocotyledonous plants, and cereals. Of particular interest are crop plants such as *Brassica*, *Arabidopsis*, soybean, corn, and the like. Prokaryotic cells of interest include *Eschericia*, *Baccillus*, *Lactobaccillus*, *cyanobacteria* and the like.

Eukaryotic cells include plant cells, mammalian cells such as those of lactating animals, avian cells such as of chickens, and other cells amenable to genetic manipulation including insect, fungal, and algae cells. Examples of host animals include mice, rats, rabbits, chickens, quail, turkeys, cattle, sheep, pigs, goats, yaks, etc., which are amenable to genetic manipulation and cloning for rapid expansion of a transgene expressing population. For animals, PKS-like transgenes can be adapted for expression in target organelles, tissues and body fluids through modification of the gene regulatory regions. Of particular interest is the production of PUFAs in the breast milk of the host animal.

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Examples of host microorganisms include Saccharomyces cerevisiae, Saccharomyces carlsbergensis, or other yeast such as Candida, Kluyveromyces or other fungi, for example, filamentous fungi such as Aspergillus, Neurospora, Penicillium, etc. Desirable characteristics of a host microorganism are, for example, that it is genetically well characterized, can be used for high level expression of the product using ultra-high density fermentation, and is on the GRAS (generally recognized as safe) list since the proposed end product is intended for ingestion by humans. Of particular interest is use of a yeast, more particularly baker's yeast (S. cerevisiae), as a cell host in the subject invention. Strains of particular interest are SC334 (Mat α pep4-3 prbl-1122 ura3-52 leu2-3, 112 regl-501 gal1; (Hovland et al (1989) Gene 83:57-64); BJ1995 (Yeast Genetic Stock Centre, 1021 Donner Laboratory, Berkeley, CA 94720), INVSC1 (Mat α hiw3Δ1 leu2 trp1-289 ura3-52 (Invitrogen, 1600 Faraday Ave., Carlsbad, CA 92008) and INVSC2 (Mat α his 3 Δ 200 ura 3-167; (Invitrogen). Bacterial cells also may be used as hosts. This includes E. coli, which can be useful in fermentation processes. Alternatively, a host such as a Lactobacillus species can be used as a host for introducing the products of the PKSlike pathway into a product such as yogurt.

The transformed host cell can be identified by selection for a marker contained on the introduced construct. Alternatively, a separate marker construct can be introduced with the desired construct, as many transformation techniques introduce multiple DNA molecules into host cells. Typically, transformed hosts are selected for their ability to grow on selective media. Selective media can incorporate an antibiotic or lack a factor

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necessary for growth of the untransformed host, such as a nutrient or growth factor. An introduced marker gene therefor may confer antibiotic resistance, or encode an essential growth factor or enzyme, and permit growth on selective media when expressed in the transformed host cell. Desirably, resistance to kanamycin and the amino glycoside G418 are of particular interest (see USPN 5,034,322). For yeast transformants, any marker that functions in yeast can be used, such as the ability to grow on media lacking uracil, lencine, lysine or tryptophan.

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Selection of a transformed host also can occur when the expressed marker protein can be detected, either directly or indirectly. The marker protein can be expressed alone or as a fusion to another protein. The marker protein can be one which is detected by its enzymatic activity; for example \(\beta\)-galactosidase can convert the substrate X-gal to a colored product, and luciferase can convert luciferin to a light-emitting product. The marker protein can be one which is detected by its light-producing or modifying characteristics; for example, the green fluorescent protein of Aequorea victoria fluoresces when illuminated with blue light. Antibodies can be used to detect the marker protein or a molecular tag on, for example, a protein of interest. Cells expressing the marker protein or tag can be selected, for example, visually, or by techniques such as FACS or panning using antibodies.

The PUFAs produced using the subject methods and compositions are found in the host plant tissue and/or plant part as free fatty acids and/or in conjugated forms such as acylglycerols, phospholipids, sulfolipids or glycolipids, and can be extracted from the host cell through a variety of means well-known in the art. Such means include extraction with organic solvents, sonication, supercritical fluid extraction using for example carbon dioxide, and physical means such as presses, or combinations thereof. Of particular interest is extraction with methanol and chloroform. Where appropriate, the aqueous layer can be acidified to protonate negatively charged moieties and thereby increase partitioning of desired products into the organic layer. After extraction, the organic solvents can be removed by evaporation under a stream of nitrogen. When isolated in conjugated forms, the products are enzymatically or chemically cleaved to release the free fatty acid or a less complex conjugate of interest, and are then subjected to further manipulations to produce a desired end product. Desirably, conjugated forms of fatty acids are cleaved with potassium hydroxide.

If further purification is necessary, standard methods can be employed. Such methods include extraction, treatment with urea, fractional crystallization, HPLC, fractional distillation, silica gel chromatography, high speed centrifugation or distillation, or combinations of these techniques. Protection of reactive groups, such as the acid or alkenyl groups, can be done at any step through known techniques, for example alkylation or iodination. Methods used include methylation of the fatty acids to produce methyl esters. Similarly, protecting groups can be removed at any step. Desirably, purification of fractions containing DHA and EPA is accomplished by treatment with urea and/or fractional distillation.

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The uses of the subject invention are several. Probes based on the DNAs of the present invention find use in methods for isolating related molecules or in methods to detect organisms expressing PKS-like genes. When used as probes, the DNAs or oligonucleotides need to be detectable. This is usually accomplished by attaching a label either at an internal site, for example via incorporation of a modified residue, or at the 5' or 3' terminus. Such labels can be directly detectable, can bind to a secondary molecule that is detectably labeled, or can bind to an unlabelled secondary molecule and a detectably labeled tertiary molecule; this process can be extended as long as is practicable to achieve a satisfactorily detectable signal without unacceptable levels of background signal. Secondary, tertiary, or bridging systems can include use of antibodies directed against any other molecule, including labels or other antibodies, or can involve any molecules which bind to each other, for example a biotin-streptavidin/avidin system. Detectable labels typically include radioactive isotopes, molecules which chemically or enzymatically produce or alter light, enzymes which produce detectable reaction products, magnetic molecules, fluorescent molecules or molecules whose fluorescence or lightemitting characteristics change upon binding. Examples of labelling methods can be found in USPN 5,011,770. Alternatively, the binding of target molecules can be directly detected by measuring the change in heat of solution on binding of a probe to a target via isothermal titration calorimetry, or by coating the probe or target on a surface and detecting the change in scattering of light from the surface produced by binding of a target or a probe, respectively, is done with the BIAcore system.

PUFAs produced by recombinant means find applications in a wide variety of areas. Supplementation of humans or animals with PUFAs in various forms can result in increased levels not only of the added PUFAs, but of their metabolic progeny as well.

Complex regulatory mechanisms can make it desirable to combine various PUFAs, or to add different conjugates of PUFAs, in order to prevent, control or overcome such mechanisms to achieve the desired levels of specific PUFAs in an individual. In the present case, expression of PKS-like gene genes, or antisense PKS-like gene transcripts, can alter the levels of specific PUFAs, or derivatives thereof, found in plant parts and/or plant tissues. The PKS-like gene polypeptide coding region is expressed either by itself or with other genes, in order to produce tissues and/or plant parts containing higher proportions of desired PUFAs or containing a PUFA composition which more closely resembles that of human breast milk (Prieto et al., PCT publication WO 95/24494) than does the unmodified tissues and/or plant parts.

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PUFAs, or derivatives thereof, made by the disclosed method can be used as dietary supplements for patients undergoing intravenous feeding or for preventing or treating malnutrition. For dietary supplementation, the purified PUFAs, or derivatives thereof, can be incorporated into cooking oils, fats or margarines formulated so that in normal use the recipient receives a desired amount of PUFA. The PUFAs also can be incorporated into infant formulas, nutritional supplements or other food products, and find use as anti-inflammatory or cholesterol lowering agents.

Particular fatty acids such as EPA can be used to alter the composition of infant formulas to better replicate the PUFA composition of human breast milk. The predominant triglyceride in human milk is reported to be 1,3-di-oleoyl-2-palmitoyl, with 2-palmitoyl glycerides reported as better absorbed than 2-oleoyl or 2-lineoyl glycerides (see USPN 4,876,107). Typically, human breast milk has a fatty acid profile comprising from about 0.15 % to about 0.36 % as DHA, from about 0.03 % to about 0.13 % as EPA, from about 0.30 % to about 0.88 % as ARA, from about 0.22 % to about 0.67 % as DGLA, and from about 0.27 % to about 1.04 % as GLA. A preferred ratio of GLA:DGLA:ARA in infant formulas is from about 1:1:4 to about 1:1:1, respectively. Amounts of oils providing these ratios of PUFA can be determined without undue experimentation by one of skill in the art. PUFAs, or host cells containing them, also can be used as animal food supplements to alter an animal's tissue or milk fatty acid composition to one more desirable for human or animal consumption.

For pharmaceutical use (human or veterinary), the compositions generally are administered orally but can be administered by any route by which they may be successfully absorbed, e.g., parenterally (i.e. subcutaneously, intramuscularly or

intravenously), rectally or vaginally or topically, for example, as a skin ointment or lotion. Where available, gelatin capsules are the preferred form of oral administration. Dietary supplementation as set forth above also can provide an oral route of administration. The unsaturated acids of the present invention can be administered in conjugated forms, or as salts, esters, amides or prodrugs of the fatty acids. Any pharmaceutically acceptable salt is encompassed by the present invention; especially preferred are the sodium, potassium or lithium salts. Also encompassed are the N-alkylpolyhydroxamine salts, such as N-methyl glucamine, described in PCT publication WO 96/33155. Preferred esters are the ethyl esters.

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The PUFAs of the present invention can be administered alone or in combination with a pharmaceutically acceptable carrier or excipient. As solid salts, the PUFAs can also be administered in tablet form. For intravenous administration, the PUFAs or derivatives thereof can be incorporated into commercial formulations such as Intralipids. Where desired, the individual components of formulations can be individually provided in kit form, for single or multiple use. A typical dosage of a particular fatty acid is from 0.1 mg to 20 g, or even 100 g daily, and is preferably from 10 mg to 1, 2, 5 or 10 g daily as required, or molar equivalent amounts of derivative forms thereof. Parenteral nutrition compositions comprising from about 2 to about 30 weight percent fatty acids calculated as triglycerides are encompassed by the present invention. Other vitamins, and particularly fat-soluble vitamins such as vitamin A, D, E and L-carnitine optionally can be included. Where desired, a preservative such as a tocopherol can be added, typically at about 0.1% by weight.

The following examples are presented by way of illustration, not of limitation.

EXAMPLES

Example 1

The Identity of ORFs Derived from Vibrio marinus

Using polymerase chain reaction (PCR) with primers based on ORF 6 of Shewanella (Sp ORF 6) sequences (FW 5' primers CUACUACUACUACCAAGCT AAAGCACTTAACCGTG, and CUACUACUACUAACAGCGAAATGCTTATCAAG for Vibrio and SS9 respectively and 3' BW primers: CAUCAUCAUCAUCAGCGACC

AAAACCAAATGAGCTAATAC for both *Vibrio* and SS9) and genomic DNAs templates from *Vibrio* and a borophyllic *photobacter* producing EPA (provided by Dr. Bartlett, UC San Diego), resulted in PCR products of *ca*.400 bases for *Vibrio marinus* (*Vibrio*) and *ca*.900 bases for SS9 presenting more than 75% homology with corresponding fragments of Sp ORF 6 (*see* Figure 25) as determined by direct counting of homologous amino acids.

A *Vibrio* cosmid library was then prepared and using the *Vibrio* ORF 6 PCR product as a probe (*see* Figure 26); clones containing at least ORF 6 were selected by colony hybridization.

Through additional sequences of the selected cosmids such as cosmid #9 and cosmid #21, a *Vibrio* cluster (Figure 5) with ORFs homologous to, and organized in the same sequential order (ORFs 6-9) as ORFs 6-9 of *Shewanella*, was obtained (Figure 7). The *Vibrio* ORFs from this sequence are found at 17394 to 36115 and comprehend ORFs 6-9.

15 <u>Table</u>

<u>Vibrio operon figures</u>

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	17394 to 25349	length = 7956 nt
	25509 to 28157	length = 2649 nt
20	28209 to 34262	length = 6054 nt
	34454 to 36115	length = 1662 nt

The ORF designations for the *Shewanella* genes are based on those disclosed in Figure 4, and differ from those published for the *Shewanella* cluster (Yazawa et al, USPN 5,683,898). For instance, ORF 3 of Figure 4 is read in the opposite direction from the other ORFs and is not disclosed in Yazawa et al USPN 5,683,898 (See Fig. 24) for comparison with Yazawa et al USPN 5,683,898).

Sequences homologous to ORF 3, were not found in the proximity of ORF 6 (17000 bases upstream of ORF 6) or of ORF 9 (ca.4000 bases downstream of ORF 9). Motifs characteristic of phosphopantethenyl transferases (Lambalot et al (1996) Current Biology 3:923-936) were absent from the Vibrio sequences screened for these motifs. In addition, there was no match to Sp ORF 3 derived probes in genomic digests of Vibrio and of SC2A Shewanella (another bacterium provided by the University of San Diego and

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also capable of producing EPA). Although ORF 3 may exist in *Vibrio*, its DNA may not be homologous to that of Sp ORF 3 and/or could be located in portions of the genome that were not sequenced.

Figure 6 provides the sequence of an approximately 19 kb *Vibrio* clone comprising ORFs 6-9. Figures 7 and 8 compare the gene cluster organizations of the PKS-like systems of *Vibrio marinus* and *Shewanella putrefacians*. Figures 9 through 12 show the levels of sequence homology between the corresponding ORFs 6, 7, 8 and 9, respectively.

Example 2

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ORF 8 Directs DHA Production

As described in example 1, DNA homologous to *Sp* ORF 6 was found in an unrelated species, SS9 *Photobacter*, which also is capable of producing EPA.

Additionally, ORFs homologous to *Sp* ORF 6-9 were found in the DHA producing V*brio marinus* (*Vibrio*). From these ORFs a series of experiments was designed in which deletions in each of *Sp* ORFs 6-9 that suppressed EPA synthesis in *E. coli* (Yazawa (1996) *supra*) were complemented by the corresponding homologous genes from *Vibrio*.

The Sp EPA cluster was used to determine if any of the Vibrio ORFs 6-9 was responsible for the production of DHA. Deletion mutants provided for each of the Sp ORFs are EPA and DHA null. Each deletion was then complemented by the corresponding Vibrio ORF expressed behind a lac promoter (Figure 13).

The complementation of a Sp ORF 6 deletion by a Vibrio ORF 6 reestablished the production of EPA. Similar results were obtained by complementing the Sp ORF 7 and ORF 9 deletions. By contrast, the complementation of a Sp ORF 8 deletion resulted in the production of C22:6. Vibrio ORF 8 therefore appears to be a key element in the synthesis of DHA. Figures 14 and 15 show chromatograms of fatty acid profiles from the respective complementations of Sp del ORF 6 with Vibrio ORF 6 (EPA and no DHA) and Sp del ORF 8 with Vibrio ORF 8 (DHA). Figure 16 shows the fatty acid percentages for the ORF 8 complementation, again demonstrating that ORF 8 is responsible for DHA production.

These data show that polyketide-like synthesis genes with related or similar ORFs can be combined and expressed in a heterologous system and used to produce a distinct PUFA species in the host system, and that ORF 8 has a role in determining the ultimate chain length. The *Vibrio* ORFs 6, 7, 8, and 9 reestablish EPA synthesis. In the case of

Vibrio ORF 8, DHA is also present (ca. 0.7%) along with EPA (ca. 0.6%) indicating that this gene plays a significant role in directing synthesis of DHA vs EPA for these systems.

Example 3

Requirements for Production of DHA

To determine how *Vibrio* ORFs of the cluster ORF 6-9 are used in combination with *Vibrio* ORF 8, some combinations of *Vibrio* ORF 8 with some or all of the other *Vibrio* ORFS 6-9 cluster were created to explain the synthesis of DHA.

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Vibrio ORFs 6-9 were complemented with Sp ORF 3. The results of this complementation are presented in Figures 16b and 16c. The significant amounts of DHA measured (greater than about 9%) and the absence of EPA suggest that no ORFs other than those of Vibrio ORFs 6-9 are required for DHA synthesis when combined with Sp ORF 3. This suggests that Sp ORF 3 plays a general function in the synthesis of bacterial PUFAs.

With respect to the DHA vs EPA production, it may be necessary to combine *Vibrio* ORF 8 with other *Vibrio* ORFs of the 6-9 cluster in order to specifically produce DHA. The roles of *Vibrio* ORF 9 and each of the combinations of *Vibrio* ORFs (6,8), (7, 8), (8, 9), etc in the synthesis of DHA are being studied.

Example 4

Plant Expression Constructs

A cloning vector with very few restriction sites was designed to facilitate the cloning of large fragments and their subsequent manipulation. An adapter was assembled by annealing oligonucleotides with the sequences AAGCCCGGGCTT and GTACAAGCCCGGGCTTAGCT. This adapter was ligated to the vector pBluescript II SK+ (Stratagene) after digestion of the vector with the restriction endonucleases Asp718 and SstI. The resulting vector, pCGN7769 had a single SrfI (and embedded SmaI) cloning site for the cloning of blunt ended DNA fragments.

A plasmid containing the napin cassette from pCGN3223, (USPN 5,639,790) was modified to make it more useful for cloning large DNA fragments containing multiple restriction sites, and to allow the cloning of multiple napin fusion genes into plant binary transformation vectors. An adapter comprised of the self annealed oligonucleotide of sequence CGCGATTTAAATGGCGCGCCCTGCAGGCGCCCCTGCAGGGCGC

GCCATTTAAAT was ligated into the vector pBC SK+ (Stratagene) after digestion of the vector with the restriction endonuclease *Bss*HII to construct vector pCGN7765. Plamids pCGN3223 and pCGN7765 were digested with *Not*I and ligated together. The resultant vector, pCGN7770 (Figure 17), contains the pCGN7765 backbone and the napin seed specific expression cassette from pCGN3223.

Shewanella constructs

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Genes encoding the Shewanella proteins were mutagenized to introduce suitable cloning sites 5' and 3' ORFs using PCR. The template for the PCR reactions was DNA of the cosmid pEPA (Yazawa et al, supra). PCR reactions were performed using Pfu DNA polymerase according to the manufacturers' protocols. The PCR products were cloned into Srfl digested pCGN7769. The primers CTGCAGCTCGAGACAATGTTGATT TCCTTATACTTCTGTCC and GGATCCAGATCTCTAGCTAGTCTTAGCTGAAGC TCGA were used to amplify ORF 3, and to generate plasmid pCGN8520. The primers TCTAGACTCGAGACATGAGCCAGACCTCTAAACCTACA and CCCGGGCTC GAGCTAATTCGCCTCACTGTCGTTTGCT were used to amplify ORF 6, and generate plasmid pCGN7776. The primers GAATTCCTCGAGACAATGCCGCTGCGCATCG CACTTATC and GGTACCAGATCTTTAGACTTCCCCTTGAAGTAAATGG were used to amplify ORF 7, and generate plasmid pCGN7771. The primers GAATTCGTCG ACACAATGTCATTACCAGACAATGCTTCT and TCTAGAGTCGACTTATAC AGATTCTTCGATGCTGATAG were used to amplify ORF 8, and generate plasmid pCGN7775. The primers GAATTCGTCGACACAATGAATCCTACAGCAA CTAACGAA and TCTAGAGGATCCTTAGGCCATTCTTTGGTTTGGCTTC were used to amplify ORF 9, and generate plasmid pCGN7773.

The integrity of the PCR products was verified by DNA sequencing of the inserts of pCGN7771, PCGN8520, and pCGN7773. ORF 6 and ORF 8 were quite large in size. In order to avoid sequencing the entire clones, the center portions of the ORFs were replaced with restriction fragments of pEPA. The 6.6 kilobase *Pacl/Bam*HI fragment of pEPA containing the central portion of ORF 6 was ligated into *Pacl/Bam*HI digested pCGN7776 to yield pCGN7776B4. The 4.4 kilobase *Bam*HI/*Bgl*II fragment of pEPA containing the central portion of ORF 8 was ligated into *Bam*HI/*Bgl*II digested pCGN7775 to yield pCGN7775A. The regions flanking the pEPA fragment and the cloning junctions were verified by DNA sequencing.

Plasmid pCGN7771 was cut with XhoI and BglII and ligated to pCGN7770 after digestion with SalI and BglII. The resultant napin/ORF 7 gene fusion plasmid was designated pCGN7783. Plasmid pCGN8520 was cut with XhoI and BglII and ligated to pCGN7770 after digestion with SalI and BglII. The resultant napin/ORF 3 gene fusion plasmid was designated pCGN8528. Plasmid pCGN7773 was cut with SalI and BamHI and ligated to pCGN7770 after digestion with SalI and BglII. The resultant napin/ORF 9 gene fusion plasmid was designated pCGN7785. Plasmid pCGN7775A was cut with SalI and ligated to pCGN7770 after digestion with SalI. The resultant napin/ORF 8 gene fusion plasmid was designated pCGN7782. Plasmid pCGN7776B4 was cut with XhoI and ligated to pCGN7770 after digestion with SalI. The resultant napin/ORF 6 gene fusion plasmid was designated pCGN7786B4.

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A binary vector for plant transformation, pCGN5139, was constructed from pCGN1558 (McBride and Summerfelt (1990) Plant Molecular Biology, 14:269-276). The polylinker of pCGN1558 was replaced as a HindIII/Asp718 fragment with a polylinker containing unique restriction endonuclease sites, AscI, PacI, XbaI, SwaI, BamHI, andNotI. The Asp718 and HindIII restriction endonuclease sites are retained in pCGN5139. PCGN5139 was digested with NotI and ligated with NotI digested pCGN7786B4. The resultant binary vector containing the napin/ORF 6 gene fusion was designated pCGN8533. Plasmid pCGN8533 was digested with Sse8387I and ligated with Sse8387I digested pCGN7782. The resultant binary vector containing the napin/ORF 6 gene fusion and the napin/ORF 8 gene fusion was designated pCGN8535 (Figure 18).

The plant binary transformation vector, pCGN5139, was digested with Asp718 and ligated with Asp718 digested pCGN8528. The resultant binary vector containing the napin/ORF 3 gene fusion was designated pCGN8532. Plasmid pCGN8532 was digested with Not1 and ligated with Not1 digested pCGN7783. The resultant binary vector containing the napin/ORF 3 gene fusion and the napin/ORF 7 gene fusion was designated pCGN8534. Plasmid pCGN8534 was digested with Sse8387I and ligated with Sse8387I digested pCGN7785. The resultant binary vector containing the napin/ORF 3 gene fusion, the napin/ORF 7 gene fusion and the napin/ORF 9 gene fusion was designated pCGN8537 (Figure 19).

Vibrio constructs

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The Vibrio ORFs for plant expression were all obtained using Vibrio cosmid #9 as a starting molecule. Vibrio cosmid #9 was one of the cosmids isolated from the Vibrio cosmid library using the Vibrio ORF 6 PCR product described in Example 1.

A gene encoding *Vibrio* ORF 7 (Figure 6) was mutagenized to introduce a *Sal*I site upstream of the open reading frame and *Bam*HI site downstream of the open reading frame using the PCR primers: TCTAGAGTCGACACAATGGCGGAATTAGCTG
TTATTGGT and GTCGACGGATCCCTATTTGTTCGTGTTTTGCTATATG. A gene encoding *Vibrio* ORF 9 (Figure 6) was mutagenized to introduce a *Bam*HI site upstream of the open reading frame and an *Xho*HI site downstream of the open reading frame using the PCR primers: GTCGACGGATCCACAATGAATATAGTAAGTAATCATTCGGCA and GTCGACCTCGAGTTAATCACTCGTACGATAACTTGCC. The restriction sites were introduced using PCR, and the integrity of the mutagenized plasmids was verified by DNA sequence. The *Vibrio* ORF 7 gene was cloned as a *Sal*I-*Bam*HI fragment into the napin cassette of *Sal-Bgl*I digested pCGN7770 (Figure 17) to yield pCGN8539. The *Vibrio* ORF 9 gene was cloned as a *Sal*I-*Bam*HI fragment into the napin cassette of *Sal-Bal*I digested pCGN7770 (Figure 17) to yield pCGN8543.

Genes encoding the *Vibrio* ORF 6 and ORF 8 were mutagenized to introduce *Sall* sites flanking the open reading frames. The *Sall* sites flanking ORF 6 were introduced using PCR. The primers used were: CCCGGGTCGACACAATGGCTAAAAAGAACA CCACATCGA and CCCGGGTCGACTCATGACATATCGTTCAAAATGTCACTGA. The central 7.3 kb *BamHI-XhoI* fragment of the PCR product was replaced with the corresponding fragment from *Vibrio* cosmid #9. The mutagenized ORF 6 were cloned into the *SalI* site of the napin cassette of pCGN7770 to yield plasmid pCGN8554.

The mutagenesis of ORF 8 used a different strategy. A BamHI fragment containing ORF 8 was subcloned into plasmid pHC79 to yield cosmid #9". A SalI site upstream of the coding region was introduced on and adapter comprised of the oligonucleotides TCGACATGGAAAATATTGCAGTAGTAGGTATTGCTAATTT GTTC and CCGGGAACAAATTAGCAATACCTACTACTGCAATATTTTCCATG. The adapter was ligated to cosmid #9" after digestion with SalI and XmaI. A SalI site was introduced downstream of the stop codon by using PCR for mutagenesis. A DNA fragment containing the stop codon was generated using cosmid #9" as a template with the primers TCAGATGAACTTTATCGATAC and TCATGAGACGTCGTCGACTTA

CGCTTCAACAATACT. The PCR product was digested with the restriction endonucleases *ClaI* and *AatII* and was cloned into the cosmid 9" derivative digested with the same enzymes to yield plasmid 8P3. The *SaII* fragment from 8P3 was cloned into *SaII* digested pCGN7770 to yield pCGN8515.

PCGN8532, a binary plant transformation vector that contains a *Shewannella* ORF 3 under control of the napin promoter was digested with *Not*I, and a *Not*I fragment of pCGN8539 containing a napin *Vibrio* ORF 7 gene fusion was inserted to yield pCGN8552. Plasmid pCGN8556 (Figure 23), which contains *Shewannella* ORF 3, and *Vibrio* ORFs 7 and 9 under control of the napin promoter was constructed by cloning the *Sse*8357 fragment from pCGN8543 into *Sse*8387 digested pCGN8552.

The NotI digested napin/ORF 8 gene from plasmid pCGN8515 was cloned into a NotI digested plant binary transformation vector pCGN5139 to yield pCGN8548. The Sse8387 digested napin/ORF 6 gene from pCGN8554 was subsequently cloned into the Sse8387 site of pCGN8566. The resultant binary vector containing the napin/ORF 6 gene fusion and napin/ORF 8 gene fusion was designated pCGN8560 (Figure 22).

Example 5 Plant Transformation and PUFA Production

EPA production

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The Shewanella constructs pCGN8535 and pCGN8537 can be transformed into the same or separate plants. If separate plants are used, the transgenic plants can be crossed resulting in heterozygous seed which contains both constructs.

pCGN8535 and pCGN8537 are separately transformed into *Brassica napus*. Plants are selected on media containing kanamycin and transformation by full length inserts of the constructs is verified by Southern analysis. Immature seeds also can be tested for protein expression of the enzyme encoded by ORFs 3, 6, 7, 8, or 9 using western analysis, in which case, the best expressing pCGNE8535 and pCGN8537 T₁ transformed plants are chosen and are grown out for further experimentation and crossing. Alternatively, the T₁ transformed plants showing insertion by Southern are crossed to one another producing T₂ seed which has both insertions. In this seed, half seeds may be analyzed directly from expression of EPA in the fatty acid fraction. Remaining half-seed

of events with the best EPA production are grown out and developed through conventional breeding techniques to provide *Brassica* lines for production of EPA.

Plasmids pCGN7792 and pCGN7795 also are simultaneously introduced into *Brassica napus* host cells. A standard transformation protocol is used (*see* for example USPN 5,463,174 and USPN 5,750,871, however *Agrobacteria* containing both plasmids are mixed together and incubated with *Brassica* cotyledons during the cocultivation step. Many of the resultant plants are transformed with both plasmids.

DHA production

A plant is transformed for production of DHA by introducing pCGN8556 and pCGN8560, either into separate plants or simultaneously into the same plants as described for EPA production.

Alternatively, the *Shewanella* ORFs can be used in a concerted fashion with ORFs 6 and 8 of *Vibrio*, such as by transforming with a plant the constructs pCGN8560 and pCGN7795, allowing expression of the corresponding ORFs in a plant cell. This combination provides a PKS-like gene arrangement comprising ORFs 3, 7 and 9 of *Shewanella*, with an ORF 6 derived from *Vibrio* and also an OFR 8 derived from *Vibrio*. As described above, ORF 8 is the PKS-like gene which controls the identity of the final PUFA product. Thus, the resulting transformed plants produce DHA in plant oil.

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Example 6

Transgenic plants containing the Shewanella PUFA genes

Brassica plants

Fifty-two plants cotransformed with plasmids pCGN8535 andpCGN8537 were analyzed using PCR to determine if the *Shewanella* ORFs were present in the transgenic plants. Forty-one plants contained plasmid pCGN8537, and thirty-five plants contained pCGN8535. 11 of the plants contained all five ORFs required for the synthesis of EPA. Several plants contained genes from both of the binary plasmids but appeared to be missing at least one of the ORFs. Analysis is currently being performed on approximately twenty additional plants.

Twenty-three plants transformed with pCGN8535 alone were analyzed using PCR to determine if the *Shewanella* ORFs were present in the transgenic plants. Thirteen of

these plants contained both *Shewanella* ORF 6 and *Shewanella* ORF 8. Six of the plants contained only one ORF.

Nineteen plants transformed with pCGN8537 were alone analyzed using PCR to determine if the *Shewanella* ORFs were present in the transgenic plants. Eighteen of the plants contained *Shewanella* ORF 3, *Shewanella* ORF 7, and *Shewanella* ORF 9. One plant contained *Shewanella* ORFs 3 and 7.

<u>Arabidopsis</u>

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More than 40 transgenic Arabidopsis plants cotransformed with plasmids pCGN8535 and pCGN8537 are growing in our growth chambers. PCR analysis to determine which of the ORFs are present in the plants is currently underway.

By the present invention PKS-like genes from various organisms can now be used to transform plant cells and modify the fatty acid compositions of plant cell membranes or plant seed oils through the biosynthesis of PUFAs in the transformed plant cells. Due to the nature of the PKS-like systems, fatty acid end-products produced in the plant cells can be selected or designed to contain a number of specific chemical structures. For example, the fatty acids can comprise the following variants: Variations in the numbers of keto or hydroxyl groups at various positions along the carbon chain; variations in the numbers and types (cis or trans) of double bonds; variations in the numbers and types of branches off of the linear carbon chain (methyl, ethyl, or longer branched moieties); and variations in saturated carbons. In addition, the particular length of the end-product fatty acid can be controlled by the particular PKS-like genes utilized.

All publications and patent applications mentioned in this specification are indicative of the level of skill of those skilled in the art to which this invention pertains. All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.

The invention now being fully described, it will be apparent to one of ordinary skill in the art that many changes and modifications can be made thereto without departing from the spirit or scope of the appended claims.

What is claimed is:

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- 1. An isolated nucleic acid comprising:
- a Vibrio marinus nucleotide sequence selected from the group consisting of the ORF 6, ORF 7, ORF 8 and ORF 9 as shown in Figure 6.
- 2. An isolated nucleic acid comprising: a nucleotide sequence which encodes a polypeptide of a polyketide-like synthesis system, wherein said system produces a docosahexenoic acid when expressed in a host cell.
- 3. The isolated nucleic acid according to Claim 2, wherein said nucleotide sequence is derived from a marine bacterium.
- 4. The isolated nucleic acid according to Claim 2, wherein said nucleotide sequence is a *Vibrio marinus* ORF 8 as shown in Figure 6.
- 5. An isolated nucleic acid comprising:
 a nucleotide sequence which is substantially identical to a sequence of at least 50
 nucleotides of a *Vibrio marinus* nucleotide sequence selected from the group consisting of ORF 6, ORF 7, ORF 8 and ORF 9 as shown in Figure 6.
- 6. A recombinant microbial cell comprising at least one copy of an isolated nucleic acid according to Claim 1 or Claim 2.
- 7. The recombinant microbial cell according to Claim 6, wherein said cell comprises each element of a polyketide-like synthesis system required to produce a long chain polyunsaturated fatty acid.
 - 8. The recombinant microbial cell according to Claim 7, wherein said cell is a eukaryotic cell.
 - 9. The recombinant microbial cell according to Claim 8, wherein said eukaryotic cell is a fungal cell, an algae cell or an animal cell.

- 10. The recombinant microbial cell according to Claim 9, wherein said fungal cell is a yeast cell and said algae cell is a marine algae cell.
- 11. The recombinant microbial cell according to Claim 6, wherein said cell is a prokaryotic cell.
 - 12. The recombinant microbial cell according to Claim 11, wherein said cell is a bacterial cell or a cyanobacterial cell.
- 13. The microbial cell according to Claim 6, wherein said recombinant microbial cell is enriched for 22:6 fatty acids as compared to a non-recombinant microbial cell which is devoid of said isolated nucleic acid.
 - 14. A method for production of docosahexenoic acid in a microbial cell culture, said method comprising:

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growing a microbial cell culture having a plurality of microbial cells, wherein said microbial cells or ancestors of said microbial cells were transformed with a vector comprising one or more nucleic acids having a nucleotide sequence which encodes a polypeptide of a polyketide synthesizing system, wherein said one or more nucleic acids are operably linked to a promoter, under conditions whereby said one or more nucleic acids are expressed and docosahexenoic acid is produced in said microbial cell culture.

15. A method for production of a long chain polyunsaturated fatty acid in a plant cell, said method comprising:

growing a plant having a plurality of plant cells, wherein said plant cells or ancestors of said plant cells were transformed with a vector comprising one or more nucleic acids having a nucleotide sequence which encodes one or more polypeptides of a polyketide synthesizing system which produces a long chain polyunsaturated fatty acid, wherein each of said nucleic acids are operably linked to a promoter functional in a plant cell, under conditions whereby said polypeptides are expressed and a long chain polyunsaturated fatty acid is produced in said plant cells.

- 16. The method according to Claim 15, wherein said long chain polyunsaturated fatty acid produced in said plant cells is a 20:5 and 22:6 fatty acid.
- 17. The method according to Claim 15, wherein said nucleic acids comprise nucleotide sequences encoding any one of the polypeptides selected from the group consisting of *Vibrio marinus* ORF 6, ORF 7, ORF 8 and ORF 9 as shown in Figure 6 and *Shewanella putrefaciens* ORF 3, ORF 6, ORF 7, ORF 8 and ORF 9 as shown in Figure 4.

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- 18. The method according to Claim 15, wherein said nucleic acid constructs are derived from two or more polyketide synthesizing systems.
 - 19. A recombinant plant cell which produces an long chain polyunsaturated fatty acid exogenous to said plant cell, wherein said recombinant plant cell is produced according to a method comprising:

transforming a plant cell or an ancestor or said plant cell with a vector comprising one or more nucleic acids having a nucleotide sequence which encodes one or more polypeptides of a polyketide synthesizing system which produces a long chain polyunsaturated fatty acid, wherein each of said nucleic acids are operably linked to a promoter functional in said plant cell whereby a recombinant plant cell is obtained; and

growing said recombinant plant cell under conditions whereby said polypeptides are expressed and a long chain polyunsaturated fatty acid is produced in said plant cell.

- 20. The recombinant plant cell according to Claim 19, wherein said recombinant plant cell is a recombinant seed cell.
- 21. The recombinant plant cell according to Claim 20, wherein said recombinant seed cell is a recombinant embryo cell.
- 22. The method according to Claim 15, wherein said long chain polyunsaturated fatty acid produced in said plant cells is eicosapentenoic acid.
 - 23. The method according to Claim 15, wherein said long chain polyunsaturated fatty acid produced in said plant cells is docosahexenoic acid.

- 24. The recombinant plant cell according to Claim 19, wherein said recombinant plant cell is from a plant selected from the group consisting of *Brassica*, soybean, safflower, and sunflower.
- 5 25. A plant oil produced by a recombinant plant cell according to Claim 19, wherein said plant oil comprises eicosapentenoic acid.
 - 26. A plant oil produced by a recombinant plant cell according to Claim 19, wherein said plant oil comprises docosahexenoic acid.
 - 27. The plant oil according to Claim 25 or Claim 26, wherein said plant oil is encapsulated.
 - 28. A dietary supplement comprising a plant oil according to Claim 27.
 - 29. A recombinant E. coli cell which produces docosahexenoic acid.
 - 30. A plant oil comprising eicosapentenoic acid.

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31. A plant oil comprising docosahexenoic acid.

32. The recombinant microbial cell according to Claim 12, wherein said bacterial cell is a lactobacillus cell.

Fig. 1 Organization of Shewanella EPA Genes and Clones Obtained from the Sagami Chemical Institute.

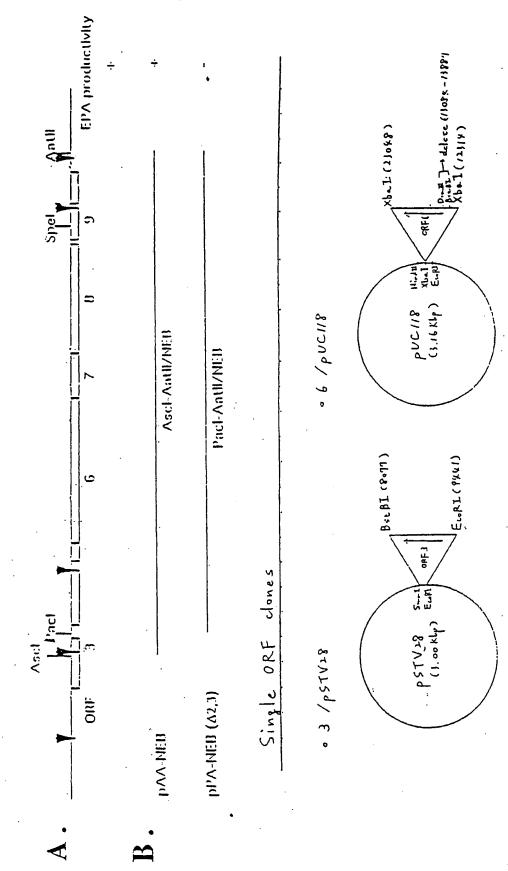
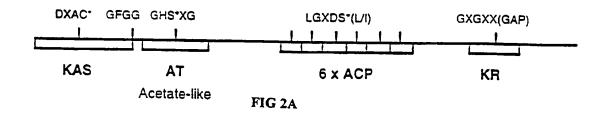
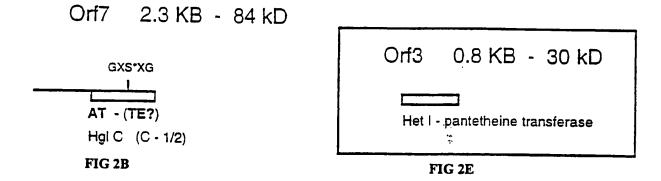


Fig. 2 S. IEWANELLA EPA C ?Fs

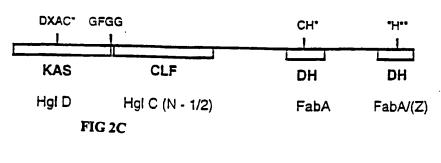
Motifs - Domains - Homologies

Orf6 8.3 KB - 293 kD



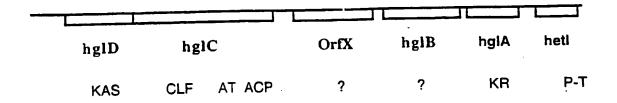


Orf8 6.0 KB - 217 kD



Orf9 1.6 KB - 59 kD

Anabeana - OrfX homolog
FIG 2D



Anabeana "PKS" Genes Involved in Heterocyst Glycolipid Synthesis**

Orf3 Encodes a Phosphopantetheine Transferase

= 30000 1. pUC19 3. pAA-Neb (EPA +)2. $pPA-NEB (\triangle Orf3)$ 4. Orf6 subclone 5. Orf6 + Orf3 subclones 6. Orf3 subclone

Autoradiograph of [C14] \(\beta \)-Alanine labelled proteins from \(E \). coli (strain SJ16) cells transformed with the above listed plasmids. Cells were grown in the presence of [C14] B-alanine and the appropriate antibiotics. Proteins were extracted, separated by SDS-PAGE and transferred to a PVDF membrane prior to autoradiography. ACP and an unknown (but previously observed) 35 kD protein were labelled in all of the samples. The high molecular mass proteins detected in lanes 2 and 5 are full-'length (largest band) and truncated products of the Shewanella Orf6 gene (confirmed by Western analysis - data not shown). E. coli strain SJ16 is conditionally blocked in B-alanine synthesis.

Sequence Range: 1 to 37895

nge: 1 to 37	895						
	20	•	40	•	60	•	80
GATCTCTTAC	AAAGAAACTA	TCTCAATGTG	AATTTAACCT	TAATTCCGTT	TAATTACGGC	CTGATAGAGC	ATCACCCAAT
	100		120	. •	140	•	160
CAGCCATAAA	actgtaaagt	GGGTACTCAA	AGGTGGCTGG	GCGATTCTTC	TCAAATACAA	AGTGCCCAAC	CCAAGCAAAT
	180		200	•	220		240
CCATATCCGA	TAACAGGTAA	AAGTAGCAAT	AAACCCCAGC	GCTGAGTTAG	TAATACATAA	GCGAATAATA	GGATCACTAA
	260		280		300		320
ACTACTGCCG	AAATAGTGTA	ATATTCGACA	GTTTCTATGC	TGATGTTGAG	AAAATAAATA	AGGGTAAAAT	TCAGCALLAG
	340		360		380		400
A A C GATAGCG	CTTACTCATT	ACTEACACET	CGGTAAAAA	GCAACTCGCC	ATTAACTTGG	CCAATCGTCA	GTTGTTCTAT
	420		440		460	•	480
	•	*****************************	TATATGTGCA	TTATCATTAG	CAAAAACTCC	GATACCATCA	AGATGAAGTT
CGTCTCAAAG		IMMINICIC			540		560
•	500	•	.520		•	-	•
GTTCATCACA		ACTGCGTCGA	TAAGCTTACT	CCATAGCCC		·	640
•	. 580	•	600	•	620		•
AACTGTAAAA	TGCCACATTG	GCCACTTGGT	AAGCTCTCTA	TAATCTGATT		ATAAGTGCCT	
•	660		680		700	•	720
CCAACCAGTA	CTTAACAACA	TCTTTAAACG	CCAATGCCAA	AAACCCGCTT	CACCTAAGGG	AACCTGCTGA	GTCACTATGC
•	740	•	760		780	•	800
AGGCTACGCC	TATCAATCTA	TCCCCAACGA	ACATACCAAT	AAGTGCTTGC	TOCTGTTGCC	AGAGCTCATT	GAGTTCTTCT
	820	_	840		860		880
CGAATAGCCC	CGCGAAGCTT	TTGCTCATAC	TGCGCTTGAT	CACCACTAAA	AAGTGTTTCG	ATAAAAAAGG	GATCATCATG
	900		920		940	_	960
ATAGGCGTTA	TAGAGAATAG	AGGCTGCTAT	GCGTAAATCT	TCTGCCGTGA	GATAAACTGC	ACGACACTCT	TCCATGGCTT
	980		1000		1020		1040
GATCTTCCAT	TGTTATTGTC	CTTGACCTTG	ATCACACAAC	ACCAATGTAA	CAAGACTGTA	TAGAAGTGCA	ATTAATAATC
0/1101110	1060		1080		1100		1120
	•	- 	TTGCTAAACA	• ብርርዊዊዊ	GCTTTGACAA	AACTTTGCCT	AGACTITAAC
AATTCGTGCA		CHOCKITICI	1160		1180		1200
•	1140		•		•	**************************************	ATAATGTAAT
GATAGAAATC		AGAAAAGCTA	CAACCTAGAG	GGGAATAATC		IMMATCING	1280
•	1220	•	1240		1260	•	•
AAACACCGAG	TTTATCGACC	ATACTTAGAT	AGAGTCATAG	CAACGAGAAT	AGTTATGGAT	ACAACGCCGC	AAGATCTATC
•	1300	•	1320	•	1340	•	1360
ACACCTGTTT	TTACAGCTAG	GATTAGCAAA	TGATCAACCC	GCAATTGAAC	AGTTTATCAA	TGACCATCAA	TTAGCGGACA
	1380	•	1400	•	1420		. 1440
ATATATTGCT	ACATCAAGCA	AGCTTTTGGA	GCCCATCGCA	AAAGCACTTC	TTAATTGAGT	CATTTAATGA	AGATGCCCAG
_	1460		1480		1500	•	1520
TGGACCGAAG	TCATCGACCA	CTTAGACACC	TTATTAAGAA	AAAACTAACC	ATTACAACAG	CARCTTTAAA	TTTTGCCGTA
•	1540)	1560	,	1580		1600
AGCCATCTCC	CCCCACCCCA	CAACAGCGTI	GTTGCTTATG	ACCACTGGAG	TACATTCGT	TTTAGTCGTT	TTACCATCAC
	1620		1640		1660		1680
CATGGGTACS	TTGAGTGCGA	TAAAAAAGC	CATABACTTO	TTTATCGGC	TGAATATAGO	CITCGITAN	ATCAGCTGTT
	1700		1720		1740		1760
- c-c-s-mm - s - c		, ,	,			, ,	CTTGAGTTTT
CCCATTAAAC		. c.c.,,mci					

Fig. 4 1/30

		1800		1820		1840
1780 CTCCCAAGCA CCGTGATTAT		•	•	•	- - -	•
	CCCAGTUAGA T		CAACATIGA	1900		1920
1860		1880		•	*	•
TCCCAAAACA TGCTAAACCT	AATAATTTAT		MCTFCC#G1	1980	1111100111	2000
1940	•	1960		•		•
AACTTACATG CCAAAACACA	AGCTGTTGTT	•	ATTATTATT	2060	AGGATATOCC	2080
2020	•	2040	*	•		•
ATAATTACCA ATGTTTAAGG	AATTTGACTA A		GATTGAGCA		GCIGCIAAA	2160
2100	•	2120		2140	*	•
ACAAGGTAGA GAACCAACAT	TAGCATTGAT		CTTGGTAATA		GCGCGAG1.A	2240
2180	•	2200	•	2220		•
TGCAACAGTT TAAGTCTATG	AGTGCAGAAG		AATACCTAGC		CAGCAAAAGA	
2260		2280		2300	•	2320
GGTCAATCAA GCTTATCTCA	ATCTGAACAA	GCTGATAGGA	TCCTCCAGCT		CTCAATGAAT	
2340		2360	•	2380	•	2400
ATTTAATGGG CTAAAAAGTC	AATTTGATAA	CTTACAACAA	AACCTGATGA			
2420	•	2440		2460	•	2480
TGAACTACGA TTTGAATGT1	TTGATAACAC	CACGATTACT	GCAGCAGAAA	AAGCCATTAA	TGGTTTGCTT	
2500	•	2520	•	2540	•	2560
GAGCCAATGG CCAGGTTCTA	GGTCGTGAAT	TTGCCGTTGC	ATTTAACGAT	GGTGAGTTTA	AAGCACGCAI	
2580		2600	•	2620	•	2640
GAAAAAAGCA GCTTATCTA	ACGCTTTAAT	AGTCCTTGGG	TAAATAGTGC	ACTCGAAGAG	CTAACCGAAC	
2666		2680	•	2700		2720
TGCGCCACGT GAAAAGTAT	A TTGGCCAAGA	TATTAATTCT	GAAGCATCTA	GCCAAGACAC	ACCAAGTTG	
274		2760		2780	,	2800
ACACAAGTTA TGTGCACAT	G TGCTCACCAC	TAAGAAATGG	CGACACCTTG	CAGCCTATTO	CACTGTATC	
282		2840	•	2860	•	2880
ACTGCCAACG GCGATCATA	A ACGAATGATC	CGTTGGCAAA	CAGAATGGCA	AGCTTGTGA	r GAATTGCAA	A TGGCCGCAGC
290	•	2920	•	2940	•	2960
TACTAAAGCT GAATTTGCC	G CACTTGAAGA	GCTAACCAGT	CATCAGAGTO	ATCTATTTA	GCGTGGTTG	G GACTTACGTG
298		3000		302	•	3040
GCAGAGTCGA ATACTTGAC	G AAAATTCCGA	CCTATTACTA	TTTATACCGT	r GTTGGCGGT	G AAAGCTTAG	C AGTAGAAAAG
306		3080	•	-	•	3120
CAGCGCTCTT GTCCTAAGT	G TGGCAGTCAA	GAATGGCTGC	TCGATAAAC	C ATTATTGGA	T ATGTTCCAT	T TTCCCTGTGA
314	0	3160	,	318	0	3200
CACCTGCCGC ATCGTATCT	A ATATCTCTTG	GGACCATTTA	TAACTCTTC	C GAGTCTTAT	C ACACTAGAG	T TTAGTCAGCA
322	:0	3240		326	•	3280
TAAAAATGGC GCTTATATT		AAATATAAGO	GCCATTTTC.	A TCGATACTA	T ATATCAGC	G ACTATTTCC
330	00	3320)	. 334	10	3360
GCGTAAATTA GCCCACAT	ATTTCATTC	T TTGCCAGATO	CCTGGATGA	T CTAGTTGTO	G CATCGACTO	T TCAATAGGTT
33	30	3400	•	342	20	3440
TAACCGCAGG TGTAACCC	IT GGAGTCAAT	r CGTTTATAA	A CTCGTTTAA	A CTGTCACT	CA ATTTAACG	CT TTGTACTTCA
34	60	348	0	350	00	3520
CCTGGAATTT CAATCCAT	AC GCTGCCATC	A CTATTATTA	A CCGTCAACA	TTTATCTT	CA TCATCAAG	AA TACCAATAAA

Fig. 4 2/30

	•					
3540		3560	•	3580		3600
CCAAGTCGCC TCTTGCTTAA	GCTTTCTCTT CAT	CATTAAA TO	ACCAATGA	TGTTTTGTTG	TAAGTATTCA	AAATCAGTTT
3620	•	3640	•	3660	•	3680
GATCCCACAC TTGGATTAGC	TCACCTTGGC CCC	CATTGTGA G	CAAAAAAT	AGCGGTGCAG	AAAAATGACT	GCCAAAAAAT
3700	•	3720	•	3740	•	3760
GGATTAATTT CTGCAGATAA	TGTCATTTCA AG	TGCTGTTT C	AACATTAGC	AAATTCACCA	GGTTGTTGAC	GTACAACCGA
3780	_	3800		3820	•	3840
TTGCCAAAAC ACTGCGCCAT	CGGAGCCCGC TT	CGGCGACA A	CACACTCAG	ACTITTCTCC	TTGCGCATAA	TATCTTGGCT
3860		3880		3900		3920
GTTCACCAAG CTTATCCATG	TAGGCTTGTT GA	TATTTAGA T.	AAAAAAAGA	TCTAAAGCAG	GTAAAGAAGA	CACTTAAGCC
3940		3960	•	3980		4000
AGTTCCAAAA TCAGTTATAA	TAGGGGTCTA TT	TTGACATG G	AAACCGTAT	TGATGACACA	ACATCATGAT	CCCTACAGTA
4020		4040		4060		4080
ACGCCCCGA ACTTTCTGAA	TTAACTTTAG GA	AAGTCGAC C	GGTTATCAA	GAGCAGTATG	ATGCATCTTT	ACTACAAGCG
. 4100		4120		4140		4160
TGCCGCGTAA ATTAAACCGT	GATGCTATCG GT	CTAACCAA T	GAGCTACCT	TTTCATGGCT	CTGATATTTG	GACTGGCTAC
4180)	4200		4220		4240
GAACTGTCTT GGCTAAATGC	TAAAGGCAAG CC	AATGATTG C	TATTGCAGA	CTTTAACCTA	AGTTTTGATA	GTAAAAATCT
4260)	4280		4300		4320
GATCGAGTCT AAGTCGTTT	A AGCTGTATTT AJ	AACAGCTAT A	ACCAAACAC	GATTTGATAG	CGTTCAAGCO	GTTCAAGAAC
4340)	4360		. 4380		4400
GTTTAACTGA AGACTTAAG	GCCTGTGCCC A	AGGCACAGT	racggtaaaa	GTGATTGAAC	CTAAGCAAT	TAACCACCTG
442	0 .	4440		4460)	4480
AGAGTGGTTG ATATGCCAG	G TACCTGCATT G	ACGATTTAG	ATATTGAAGT	TGATGACTAT	AGCTTTAAC	T CTGACTATCT
450	0	4520		4540)	4560
CACCGACAGT GTTGATGAC	A AAGTCATGGT T	GCTGAAACG	CTAACGTCAA	ACTTATTGA	ATCAAACTG	C CTAATCACTT
458	0	4600	•	4620		. 4640
CTCAGCCTGA CTGGGGTAC	A GTGATGATCC G	TTATCAAGG	GCCTAAGATA	GACCGTGAA	A AGCTACTTA	G ATATCTGATT
466	0	4680		. 470	o •	4720
TCATTTAGAC AGCACAATG	A ATTTCATGAG C	AGTGTGTTG	AGCGTATAT	TGTTGATTT	A AAGCACTAT	T GCCAATGTGC
474	0	4760		478	0	4800
CAAACTTACT GTCTATGCA	C GTTATACCCG C	CCTCCTCCT	TTAGATATC	A ACCCATATO	G TAGCGACTT	T GAAAACCCTG
482	:0	4840		486	0	4880
CAGAAAATCA GCGCCTAGG	G AGACAGTAAT 1	IGATTGCAGT	ACCTACAAA	A AACAATGCC	T ATAAGCCAJ	G CTTATGGGCA
490		4920		494	•	4960
TTTTTATATT ATCAACTT	T CATCAAACCT (CAGCCGCCAA	GCCTTTTAG	T TTTATCGCT	A AATTAAGCO	CG CTCTCTCAGC
. 491	90	5000		502	10	5040
CAAATATTTG CAGGATTT	IG CTGTAATTTA '	TGGCTCCACA	CCATGAAAT	A CTCTATCGC	SC TCTACCGC	AA AAGGTAAGTC
50	60	5080		510	00	5120
AAATACCTGT AAGCCAAA	CA GCTTGGCATA	TTCGTCAGTG	TGGGCTTTT	G ACGCGATA	GC TAACGCAT	CA CTTTTTGAGG
51	40	5160	•	511	BO .	5200
CAACCGACAT CATACTTA	AT ATTGATGATT	GCTCGCTGTG	CATTTGCCT	TT GCCGGTAA	CA CCTGTTTA	GT CAGCAAGTCG
•	20	5240		52	•	5280
GCAACACTTA AATTGTAG	CG GCGCATCTTA	PATAATAAAA	GCTTTTCA	TT AAAGTATT	GC TCTTGCGT	
53	000	5320)	53	40	5360

Fig 4 3/30

GATCC	ተጥርርር	TGI	ACC A T	• •	GTG	CAC	ACA J	AACT!	LATT?	T AT	CTG	ATTA	CT	rttt(ACT	CTT	TAAA	GCC (GCAG	ATTC	TG	
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GCAGC	CAAA	TATO			AAA	TCCAG	· ccr	rrrc:	ragt'	· TG TA	AGGT	CATO	TG	CAAC	CTI	CTT	CAAT	GAG	CGGC	GGCT	CY	
				5460					54			٠			5500						20	
CGAAA	TACA	A TAT	rtaa'	TTGC	AGT	GCCC'	rgt :	AACA	CTTG	· CT C	AATT'	TGAT	TT	GCAAG	CAGT	TGT.	attg	CCG	ACTC	GCTG	GC	
				5540					55						5580						00	
ATACA	CATA	· A AA	AGTT	• CGCT	CAC	TTGA	AGT	GGGG	AADT	AT G	CTTC	AAAG	TA	GTCG	CAAC	TTG	CTCA	ATT	GTTG	ACAT	AG	
				5620					56	40				:	5660					56	80	
CGCCC	GCGA	· G CT	CTTG	• A ATA	AGC	GTCA	TCG	CACT	TGCG	GT A	GGTT	TAAC	T CC	CCTA	CCCA	CTC	GAGT	AAA'	CAAC	TCTI	rc t	
				5700					57	20					5740					57	760	
CCAAC	ATA:	C TT	TTTA	GCCT	CGA	AATC	GCA	TTAC	TAAC	CG A	CGAC	TGAG	T CA	AATC	CAGC	TCT	TCTC	CCC	CCCG	GCT	AAA	
				5780					50	00					5820					58	840	
AGATO	SAGGT	c cc	ATAC	ACCG	CAC	AAAT	AAC	GCGA	ATAA	L AA	TAAG	ATCA	AA A	GCTT.	TTTG	CTC	CGAC	ATA	AATO	CAGC	TAT	
				5860						980	٠				5900						920	
CTCC	TATC	C TT	ATCC	TATE	CCI	ATATT	AAA	AGTT	'AGC'	rcc r	GAGC	ACTO	T AC	CTCA	AAAA	CAZ	CTC	\GCG	TATT	DAAT	CCA	
				5940						60					5980						000	
ATAT'	TTTGC	G AA	CTC	ATTA	(TA	ATTC?	AATA	AAAT	ACT	TT	LATA	ATATA	CA AJ	ROCA	LAGTC	ATA	'TTA	ragc	CCT			
				6020						040					6060			•		_	080	
AATC	OTTAA	A AC	TTAC	CTAT	r AC	rggco	ADTC	ATTA	AGC	AAA 1	rgtc:	CATO	:A G:	rctcc	CTGC			ATGC	AAT	ATTG	AGA	
		•		6100						120				•		61	•			•		
CATA	AAGC"	T T	SAAC	rgat?	r ca	ATCT	TACG	¥ĊĊſ	CTAA	CTT	M M	AAA (Q Q	T (L L	M M	A A	I	S	I	M>	0465
	616	0					6	180						62	00							
	CTT			TTC A	AAT	GCG (CTA	GCA (ccc	CAA	CAT	GAA (E	CAT	GAC (CAC I	ATC .	ACT T	G T T V	GAT D	TAC Y	GAA E>	
_	L	F	S	F		A 240	ь	Α.	^	Q	п	62		-		•	-		628			
6220	AAA	CCC /	ecs *	404	_	•	۸۲۲	ATA .	CCT	CAC	AAC		•	GTA	GCT i	AAA	ACA	CTT	AAC	* TTT	GCC	
G				T	E	Н	T	I	A	н	N	Q	A	V	A	ĸ	T	L	N	F	A>	
			_	300				•			20			•			634	•			•	
GAC D	ACG T	CGT R		TTT F	GAG E	CAA	TCG S	TCT S	AAA K	TAA N	CTA L	GTC V	GCC A	AAG K	TTT F	GAT D	AAA K	GCA A	ACT T	GCC A	GAT D>	
	- 6	360				-		63	80						640	0						
A TA	mm n	*	GCC	GAA	TTT	* GCT	TTT	TTA	• AGC	GAT	GAA	ATC	ССТ	GAC	TCG	• GTT	AAC	ccG	TCT	CTC	TAC	
I	Ľ	R	A	E	F	A	F	I	s	D	E	1	P	D	s	V	N	P	3	-		
6420							40			•			646	•			•			6480		
CGT R	CAG Q	GCT A	CAG Q	CTT L	AAT N	ATG M	OTC V	CCT P	taa N	GGT G	CTG L	TAT Y	AAA K	GTG V	AGC S	GAT D	GGC	ATT	Y	Q	V>	
					500						652							6540				
CGC	GCT	* ACC	GAC	TTA	TCT	AAC	CTT	ACA	CTT	ATC	CGC	AGT	GAT	AAC	CGT	TGG	ATA	GCA	TAC	GAT	GTT	
R	Ğ.			L	s	N	L	T			R	Ş	D	N				^	•	,	**	
•			560			•			65	•				mm s		•		CCT		CAT	· ccc	
TTG L	TTA L	ACC T	K	GAA E	GCA A	GCA A	K	A	S	L	Q	F	A	L	K	N	L	P	K	D	G>	
6	620						66	40						6660						6	680	Ein 4
GAT	TTA	ccc	GTT	GTT	GCG	ATG	TTA	TAC	TCC	CAT	AGC	CAT	GCG	GAC	CAC	TTT	GGC	GGA G	GCT A	CGC	GGT G>	114.1
D	'n	۲	V	٧		.00 M	•	•	3	^		6720		,	••	•	•		5740	••	-	4/20
- Come	n	GAC	እጥ ር	- Transc		•	ርጥር		• 	` TAC		٠		י אאר	ልጥና	ACT	LAA!		•	r GTC	GAT	-1/20
GT	. CAA	UAU	~10		1	JA I	910	~~~											T .	17	n.	

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6780
GAG AAC GTA CTT GCC GGT AAC GCC ATG AGC CGC CGC GCA GCT TAT CAA TAC GGC GCA ACA CTG GGC E N V L A G N A N S R R A A Y Q Y G A T L G>
                                           6840
AAA CAT GAC CAC GGT ATT GTT GAT GCT GCG CTA GGT AAA GGT CTA TCA AAA GGT GAA ATC ACT TAC K H D H G I V D A A L G K G L S K G E I T Y>
GTC GCC CCA GAC TAC ACC TTA AAC AGT GAA GGC AAA TGG GAA ACG CTG ACG ATT GAT GGT CTA GAG V A P D Y T L N S E G K W E T L T I D G L E>
                                                           6980
ATG GTG TTT ATG GAT GCC TCG GGC ACC GAA GCT GAG TCA GAA ATG ATC ACT TAT ATT CCC TCT AAA 'M V F H D A S G T E A E S E M I T Y I P S K>
AAA GCG CTC TGG ACG GCG GAG CTT ACC TAT CAA GGT ATG CAC AAC ATT TAT ACG CTG CGC GGC GCT K A L W T A E L T Y Q G H H N I Y T L R G A>
                                                                           7120
 AAA GTA CGT GAT GCG CTC AAG TGG TCA AAA GAT ATC AAC GAA ATG ATC AAT GCC TTT GGT CAA GAT K V R D A L K W S K D I N E M I N A F G Q D>
                                                                7180
 GTC GAA GTG CTG TTT GCC TCG CAC TCT GCG CCA GTG TGG GGT AAC CAA GCG ATC AAC GAT TTC TTA V E V L F A S H S A P V W G N Q A I N D F L>
                                                     7240
 CGC CTA CAG CGT GAT AAC TAC GGC CTA GTG CAC AAT CAA ACC TTG AGA CTT GCC AAC GAT GGT GTC R L Q R D N Y G L V H N Q T L R L A N D G V>
 GGT ATA CAA GAT ATT GGC GAT GCG ATT CAA GAC ACG ATT CCA GAG TCT ATC TAC AAG ACG TGG CAT G I Q D I G D A I Q D T I P E S I Y K T W H>
 ACC AAT GGT TAC CAC GGC ACT TAT AGC CAT AAC GCT AAA GCG GTT TAT AAC AAG TAT CTA GGC TAC T N G Y H G T Y S H N A K A V Y N K Y L G Y>
  TTC GAT ATG AAC CCA GCC AAC CTT AAT CCG CTG CCA ACC AAG CAA GAA TCT GCC AAG TTT GTC GAA F D M N P A N L N P L P T K Q E S A K F V E>
  TAC ATG GGC GGC GCA GAT GCC GCA ATT AAG CGC GCT AAA GAT GAT TAC GCT CAA GGT GAA TAC CGC Y M G C A D A A I K R A K D D Y A Q G E Y R>
  TTT GTT GCA ACG GCA TTA AAT AAG GTG GTG ATG GCC GAG CCA GAA AAT GAC TCC GCT CGA TTG
       V A T A L N K V V M A E P E N D S A R Q L>
  CTA GCC GAT ACC TAT GAG CAA CTT GGT TAT CAA GCA GAA GGG GCT GGC TGG AGA AAC ATT TAC TTA
L A D T Y E Q L G Y Q A E G A G W R N I Y L>
   ACT GGC GCA CAA GAG CTA CGA GTA GGT ATT CAA GCT GGC GCG CCT AAA ACC GCA TCG GCA GAT GTC
T G A Q E L R V G I Q A G A P K T A S A D V>
  7740
                                                                              7780
   ATC AGT GAA ATG GAC ATG CCG ACT CTA TTT GAC TTC CTC GCG GTG AAG ATT GAT AGT CAA CAG GCG I S E M D M P T L F D F L A V K I D S Q Q A>
                                                                  7840
   GCT AAG CAC GGC TTA GTT AAG ATG AAT GTT ATC ACC CCT GAT ACT AAA GAT ATT CTC TAT ATT GAG A K H G L V K M N V I T P D T K D I L Y I E>
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    CTA AGC AAC GGT AAC TTA AGC AAC GCA GTG GTC GAC AAA GAG CAA GCA GCT GAC GCA AAC CTT ATG
L S N G N L S N A V V D K E Q A A D A N L M>
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Fig. 4 5/30

7960 GTT AAT AAA GCT GAC GTT AAC CGC ATC TTA CTT GGC CAA GTA ACC CTA AAA GCG TTA TTA GCC AGC V N K A D V N R I L L G Q V T L K A L L A S> 8040 GGC GAT GCC AAG CTC ACT GGT GAT AAA ACG GCA TTT AGT AAA ATA GCC GAT AGC ATG GTC GAG TTT G D A K L T G D K T A F S K 1 A D S M V E F> 8100 8120 ACA CCT GAC TTC GAA ATC GTA CCA ACG CCT GTT AAA TGAGGCA TTAATCTCAA CAAGTGCAAG CTAGACATAA 8180 AAATGGGGCG ATTAGACGCC CCATTTTTA TGCAATTTTG AACTA GCT AGT CTT AGC TGA AGC TCG AAC AAC <S T K A S A R V V 8240 AGC TTT AAA ATT CAC TTC TTC TGC TGC AAT ACT TAT TTG CTG ACA CTG ACC AAT ACT CAG TGC AAA A K F N V E E A A I S I Q Q C Q G I S L A F
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80 8320 B280 ACG ATA ACT ATC ATC AAG ATG GCC CAG TAA ACA ATG CCA ATT ATC AGC AGC GTT CAT TTG CTG TTC < R $\,$ Y $\,$ S $\,$ D $\,$ D $\,$ L $\,$ H $\,$ G $\,$ L $\,$ C $\,$ H $\,$ W $\,$ N $\,$ D $\,$ A $\,$ N $\,$ M $\,$ Q $\,$ Q $\,$ E 8380 TTT AGC CTC AAT CAA ACC TAA ACC AGA CTT TTG TGG CTC AGC GTT AGG CTT ATT AGA ACT CGA CTC <K A E I L G L G S K Q P E A N P K N S S S E 8440 TAG TAA AGC AAG ACC AAT ATC TTG TTT TAA CAA AAC CTG TCG CTG ATT AAG TTG ATG CTC AAC CTT
L L A L G I D Q K L L V Q R Q N L Q H E V K
8500
8540 GTG ATC CGC AAT AGC ATC GGA AAT ATC AAC ACA ATG GCT CAA GCT TTT AGG TGC ATT AAC TCC AAG
<H D A I A D S I D V C H S L S K P A N V G L 8560 8580 AAA AGT TTC GCT CAG TGC AGA GAA GTC AAA CGC AAA AGA TTT TAG CGA TAA TGC CAG CCC AAG TCC <F T E S L λ S F D F λ F S K L S L λ L G L G 8660 8640 TTT CGC TTT AAT GTA AGA CTC CTT GAG CGC CCA CAA ATC AAA AAA GCG GTC TCG CTG CAA GGC CTC <K A K I Y S E K L A W L D F F R D R Q L A E 8720 8700 TGG TAA CGC TAA CAA GGC TCG CTT TTC TGA TTC AGA GAA ATA ATG ACT AAG AAT AGA GTG GAT ATT

<P L A L L A R K E S E S F Y H S L I S H I N 8780 GGT GCT GTT ACG GCA ACG CTC AAT GTC GAC GCC AAA CTC AAT ACT AGC AGA GTC AGT TTC CTC CTT S N R C R E I D V G F E I S A S D T E E K 8860 8840 GCT TGC CTG ACT GGC GCC TTT ATT ATC AGC AGT GCA AAT GCC TAC TAA TAG CCA ATC TCC ACT ATG <S A Q S A G K N D A T C I G V L L W D G S H 8920 ACT CAC ATT AAA GTG GAC CCC GGT TTG AGC AAA TTG CGC ATC ACT CAA TCT AGG CTT ACC TTT GTC <S V N F H V G T Q A F Q A D S L R P K G K D 8980 8960 GCC ATA TTC AAA GCG CCA TTC ATT GGG GCG TAT TTC ACT ATG TTG TGA CAA TAA AGC GCG CAA ATA CGC GC GCC TCT TAC CAT TAAA CCTTGAGTTT TAGCTTCTTG TTTAATGTAG CGATTAACCT TAATTAACTC ATCTTCAGGC 9100 9120 9140

AGCCATGACT TAACCAACTC TGTAGTCTGG TTATCGCACT CTTGTATTGT TAACGGACAG AAGTATAAGG AAATCAATCG

6/30

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   GCA TTT GGT GGT GGA ATT GGT CGC TCA ACC TTT GAC AGT AAC GGC AAT CCT ATT GCA CAA CAA GAA A F G G G I G R S T F D S N G N P I A Q Q E>
                                                                    10620
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   CGT GAT GGG ACT AAC AGC TTT GCA TTT GGT TCA TTC CCT AAT GGC TGT GAC ACA TGT TTC AAC ACT R D G T N S F A F G S F P N G C D T C F N T>
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   GAA GCA TAC GAA AAC TAT ATT CCA GGG GTA GAA AGA ATA AAC GTT GGC TCA TCA TTC AAC TTT GAT E A Y E N Y I P G V E R I N V G S S F N F D>
                                                  10740
   TTT ACC GAT AAC ATT CAA TTT TAC ACT GAC TTC AGA TAT GTA AAG TCA GAT ATT CAG CAA CAA TTT F T D N I Q F Y T D F R Y V K S D I Q Q Q F>
                                           10800
    CAG CCT TCA TTC CGT TTT GGT AAC ATT AAT ATC AAT GTT GAA GAT AAC GCC TTT TTG AAT GAC GAC Q P S F R F G N I N I N V E D N A F L N D D>
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    TTG CGT CAG CAA ATG CTC GAT GCG GGT CAA ACC AAT GCT AGT TTT GCC AAG TTT TTT GAT GAA TTA
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    GGA AAT CGC TCA GCA GAA AAT AAA CGC GAA CTT TTC CGT TAC GTA GGT GGC TTT AAA GGT GGC TTT G N R S A E N K R E L F R Y V G G F K G G F>
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    GAT ATT AGC GAA ACC ATA TTT GAT TAC GAC CTT TAC TAT GTT TAT GGC GAG ACT AAT AAC CGT CGT D I S E T I F D Y D L Y Y V Y G E T N N R R>
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    AAA ACC CTT AAT GAC CTA ATT CCT GAT AAC TTT GTC GCA GCT GTC GAC TCT GTT ATT GAT CCT GAT K T L N D L I P D N F V A A V D S V I D P D>
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    ACT GGC TTA GCA GCG TGT CGC TCA CAA GTA GCA AGC GCT CAA GGC GAT GAC TAT ACA GAT CCC GCG T G L A A C R S Q V A S A Q G D D Y T D P A>
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     TOT GTA AAT GGT AGC GAC TGT GTT GCT TAT AAC CCA TTT GGC ATG GGT CAA GCT TCA GCA GAA GCC
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     CGC GAC TGG GTT TCT GCT GAT GTG ACT CGT GAA GAC AAA ATA ACT CAA CAA GTG ATT GGT GGT ACT R D W V S A D V T R E D K I T Q Q V I G G T>
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     CTC GGT ACC GAT TCT GAA GAA CTA TTT GAG CTT CAA GGT GGT GCA ATC GCT ATG GTT GTT GGT TTT
      L G T D S E E L F E L Q G G A I A M V V G F>
                                   11400
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     GAA TAC CGT GAA GAA ACG TCT GGT TCA ACA ACC GAT GAA TTT ACT AAA GCA GGT TTC TTG ACA AGC E Y R E E T S G S T T D E F T K A G F L T S>
                                          11460
     GCT GCA ACG CCA GAT TCT TAT GGC GAA TAC GAC GTG ACT GAG TAT TTT GTT GAG GTG AAC ATC CCA
               T P D S Y G E Y D V T E Y F V E V N I P>
                                    11520
     GTA CTA AAA GAA TTA CCT TTT GCA CAT GAG TTG AGC TTT GAC GGT GCA TAC CGT AAT GCT GAT TAC V L K E L P F A H E L S F D G A Y R N A D Y>
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                           11580
      TCA CAT GCC GGT AAG ACT GAA GCA TGG AAA GCT GGT ATG TTC TAC TCA CCA TTA GAG CAA CTT GCA
       S H A G K T E A W K A G M F Y S P L E Q L A>
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      TTA CGT GGT ACG GTA GGT GAA GCA GTA CGA GCA CCA AAC ATT GCA GAA GCC TTT AGT CCA CGC TCT
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Fig. 4 8/30

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TTG AAG CGT TAT GGC GTT TCA CGG GTG AGT GAT ATC CAA AGC CCA CAC GTA CCA ATG CGC TAC TTT L K R Y G V S R V S D I Q S P H V P M R Y F>
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      13560
 AAA ACC TTA GCC TTA CCT AGT ACC GAT GGT GAC AAT GTG GTG CAA GTG GTG TCT CTC ACC ATT CCA
 K T L A L P S T D G D N V V Q V V S L T I P>
                                                      13660
 CCA AAG TTA ACC GAA GAA GCA CCC AGT TCA ATT TTG CTC GGC ATT GAT CCT CAT AGC GAC TGG ATC P K L T E E A P S S I L L G I D P H S D W I>
                                               13720
 TAT CTC GAC ATA TAC CAA GAT GGC AAT ACA CAA GCC ACG AAT AGA TAT ATG GCT TAT GTG CTA AAA Y L D I Y Q D G N T Q A T N R Y M A Y V L K>
                                                                     13800
                                      13780
 CAC GGG CCA TTC CAT TTA CGA AAG TTA CTC GTG CGT AAC TAT CAC ACC TTT TTA CAG CGC TTT CCT H G P F H L R K L L V R N Y H T F L Q R F P>
                                                 13860
                            13840
 GGA GCG ACG CAA AAT CGC CGC CCC TCT AAA GAT ATG CCT GAA ACA ATC AAG ACG CCT GAA ACA
  G A T Q N R R P S K D M P E T I N K T P E
                    13900
                                                                                 13940
                                                  13920
 CAG GCA CCC AGT GGA GAC TCA TA ATG AGC CAG ACC TCT AAA CCT ACA AAC TCA GCA ACT GAG CAA
                                                                                                            0-f6
  Q A P S G D S>
                                           13980
 GCA CAA GAC TCA CAA GCT GAC TCT CGT TTA AAT AAA CGA CTA AAA GAT ATG CCA ATT GCT ATT GTT A Q D S Q A D S R L N K R L K D M P I A I V>
                                   14040
                                                                 14060
  GGC ATG GCG AGT ATT TTT GCA AAC TCT CGC TAT TTG AAT AAG TTT TGG GAC TTA ATC AGC GAA AAA G M A S I F A N S R Y L N K F W D L I S E K>
                                                        14120
                          14100
  ATT GAT GCG ATT ACT GAA TTA CCA TCA ACT CAC TGG CAG CCT GAA GAA TAT TAC GAC GCA GAT AAA I D A I T E L P S T H W Q P E E Y Y D A D K>
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14180
ACC GCA GCA GAC AAA AGC TAC TGT AAA CGT GGT GGC TTT TTG CCA GAT GTA GAC TTC AAC CCA ATG T A A D K S Y C K R G G F L P D V D F N P M>
                                           14240
GAG TIT GGC CTG CCG CCA AAC ATT TTG GAA CTG ACC GA\dagger TCA TCG CAA CTA TTA TCA CTC ATC GTT E F G L P P N 1 L E L T D S S Q L L S L I V>
                                  14300
GCT AAA GAA GTG TTG GCT GAT GCT AAC TTA CCT GAG AAT TAC GAC CGC GAT AAA ATT GGT ATC ACC A K E V L A D A N L P E N Y D R D K I G I T>
                                                                                             14400
 TTA GGT GTC GGC GGT GGT CAA AAA ATT AGC CAC AGC CTA ACA GCG CGT CTG CAA TAC CCA GTA TTG L G V G G G O K I S H S L T A R L Q Y P V L>
            14420
 AAG AAA GTA TTC GCC AAT AGC GGC ATT AGT GAC ACC GAC AGC GAA ATG CTT ATC AAG AAA TTC CAA K V F A N S G I S D T D S E M L I K K F Q>
                                                                        14520
 GAC CAA TAT GTA CAC TGG GAA GAA AAC TCG TTC CCA GGT TCA CTT GGT AAC GTT ATT GCG GGC CGT D Q Y V H W E E N S F P G S L G N V I A G R> ^\circ
                                                              14580
                            14560
  ATC GCC AAC CGC TTC GAT TTT GGC GGC ATG AAC TGT GTG GTT GAT GCT GCC TGT GCT GGA TCA CTT

I A N R F D F G G M N C V V D A A C A G S L>
                                                   14640
  GCT GCT ATG CGT ATG GCG CTA ACA GAG CTA ACT GAA GGT CGC TCT GAA ATG ATG ATC ACC GGT GGT A A M R M A L T E L T E G R S E M M I T G G>
                                          14700
        14680
  GTG TGT ACT GAT AAC TCA CCC TCT ATG TAT ATG AGC TTT TCA AAA ACG CCC GCC TTT ACC ACT AAC V C T D N S P S M Y M S F S K T P A F T T N>
                                                                   14780
   GAA ACC ATT CAG CCA TTT GAT ATC GAC TCA AAA GGC ATG ATG ATT GGT GAA GGT ATT GGC ATG GTG E T I Q P F D I D S K G M M I G E G I G M V>
   GCG CTA AAG CGT CTT GAA GAT GCA GAG CGC GAT GGC GAC CGC ATT TAC TCT GTA ATT AAA GGT GTG A L K R L E D A E R D G D R I Y S V I K G V>
                                               14900
    GGT GCA TCA TCT GAC GGT AAG TTT AAA TCA ATC TAT GCC CCT CGC CCA TCA GGC CAA GCT AAA GCA
     G A S S D G K F K S I Y A P R P S G Q A K A>
                                     14960
    CTT AAC CGT GCC TAT GAT GAC GCA GGT TTT GCG CCG CAT ACC TTA GGT CTA ATT GAA GCT CAC GGA L N R A Y D D A G F A P H T L G L I E A H G>
                                                               15040
     15100
     GAT ACC AAG CAA CAC ATT GCG CTA GGT TCA GTT AAA TCA CAA ATT GGT CAT ACT AAA TCA ACT GCA
D T K Q H I A L G S V K S Q I G H T K S T A>
                                           15160
      GGT ACA GCA GGT TTA ATT AAA GCT GCT CTT GCT TTG CAT CAC AAG GTA CTG CCG CCG ACC ATT AAC G T A G L I K A A L A L H H K V L P P T I N>
      GTT AGT CAG CCA AGC CCT AAA CTT GAT ATC GAA AAC TCA CCG TTT TAT CTA AAC ACT GAG ACT CGT V S Q P S P K L D I E N S P F Y L N T E T R>
       CCA TGG TTA CCA CGT GTT GAT GGT ACG CCG CGC CGC GCG GGT ATT AGC TCA TTT GGT TTT GGT GGC P W L P R V D G T P R R A G I S S F G F G G>
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rig.4

16 / 106 ACT AAC TTC CAT TTT GTA CTA GAA GAG TAC AAC CAA GAA CAC AGC CGT ACT GAT AGC GAA AAA GCT T N F H F V L E E Y N Q E H S R T D S E K A> ANG TAT COT CAA CGC CAA GTG GCG CAA AGC TTC CTT GTT AGC GCA AGC GAT AAA GCA TCG CTA ATT Y R Q R Q V A Q S F L V S A S D K A S L I> AAC GAG TTA AAC GTA CTA GCA GCA TCT GCA AGC CAA GCT GAG TTT ATC CTC AAA GAT GCA GCA GCA ELN V L A A S A S Q A E F I L K D A A A> AAC TAT GGC GTA CGT GAG CTT GAT AAA AAT GCA CCA CGG ATC GGT TTA GTT GCA AAC ACA GCT GAA N Y G V R E L D K N A P R I G L V A N T A E> GAG TTA GCA GGC CTA ATT AAG CAA GCA CTT GCC AAA CTA GCA GCT AGC GAT GAT AAC GCA TGG CAG E L A G L I K Q A L A K L A A S D D N A W Q> CAA GGT TCA CAA TAT CTC AAT ATG GGC CGT GAC CTT ACT TGT TAT TAC CCA GAG ATG CGT CAG CAA Q G S Q Y L N M G R D L T C Y Y P E M R Q Q> TTT GTA ACT GCA GAT AAA GTA TTT GCC GCA AAT GAT AAA ACG CCG TTA TCG CAA ACT CTG TAT CCA F V. T A D K V F A A N D K T P L S Q T L Y P> AAG CCT GTA TTT AAT AAA GAT GAA TTA AAG GCT CAA GAA GCC ATT TTG ACC AAT ACC GCC AAT GCC K P V F N K D E L K A Q E A I L T N T A N A> GAC ATG GTT GCA GGC CAT AGC TTT GGT GAG CTA AGT GCA CTG TGT GCT GCA GGT GTT ATT TCA GCT M V A G H S F G E L S A L C A A G V I S A> GAT GAC TAC TAC AAG CTG GCT TTT GCT CGT GGT GAG GCT ATG GCA ACA AAA GCA CCG GCT AAA GAC D D Y Y K L A F A R G E A M A T K A P A K D> GTA ATT GCA GGC CCA ACA GCA ACT ACC CCT GAT GCG GCT AAA GCG CTA ACT GAG CTT GGT TAC AAA I A G P T A T T A D A A K A L T E L G Y GCG ATT AAC CTG CCA GTA TCA GGT GCA TTC CAC ACT GAA CTT GGT GGT CAC GCT CAA GCG CCA TTT A I N L P V S G A F H T E L V G H A Q A P F> GCT AAA GCG ATT GAC GCA GCC AAA TTT ACT AAA ACA AGC CGA GCA CTT TAC TCA AAT GCA ACT GGC

GGA CTT TAT GAA AGC ACT GCT GCA AAG ATT AAA GCC TCG TTT AAG AAA CAT ATG CTT CAA TCA GTG

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CGC TTT ACT AGC CAG CTA GAA GCC ATG TAC AAC GAC GGC GCC CGT GTA TTT GTT GAA TTT GGT CCA
    F T S Q L E A M Y N D G A R V F V E
                                                                        16640
AAG AAC ATC TTA CAA AAA TTA GTT CAA GGC ACG CTT GTC AAC ACT GAA AAT GAA GTT TGC ACT ATC K N I L Q K L V Q G T L V N T E N E V C T I>
                                                                16700
TCT ATC AAC CCT AAT CCT AAA GTT GAT AGT GAT CTG CAG CTT AAG CAA GCA GCA ATG CAG CTA GCG S I N P N P K V D S D L Q L K Q A A M Q L A>
                                                       16760
GTT ACT GGT GTG GTA CTC AGT GAA ATT GAC CCA TAC CAA GCC GAT ATT GCC GCA CCA GCG AAA AAG V T G V V L S E I D P \dot{\rm Y} Q A D I A A P A K K>
TCG CCA ATG AGC ATT TCG CTT AAT GCT GCT AAC CAT ATC AGC AAA GCA ACT CGC GCT AAG ATG GCC S P M S I S L N A A N H I S K A T R A K M A>
                                                                 16900
AAG TCT TTA GAG ACA GGT ATC GTC ACC TCG CAA ATA GAA CAT GTT ATT GAA GAA AAA ATC GTT GAA K S L E T G I V T S Q I E H V I E E K I V E>
                                                           16960
V E K L V E V E K I V E K V V E V E K V
                                                 17020
GTT GAA GCT CCT GTT AAT TCA GTG CAA GCC AAT GCA ATT CAA ACC CGT TCA GTT GTC GCT CCA GTA V E A P V N S V Q A N A I Q T R S V V A P V>
                                         17080
ATA GAG AAC CAA GTC GTG TCT AAA AAC AGT AAG CCA GCA GTC CAG AGC ATT AGT GGT GAT GCA CTC I E N Q V V S K N S K P A V Q S I S G D A L>
                                17140
                                                           17160
 AGC AAC TTT TTT GCT GCA CAG CAG CAA ACC GCA CAG TTG CAT CAG CAG TTC TTA GCT ATT CCG CAG S N F F A A Q Q Q T A Q L H Q Q F L A I P Q>
                                                   17220
 CAA TAT GGT GAG ACG TTC ACT ACG CTG ATG ACC GAG CAA GCT AAA CTG GCA AGT TCT GGT CTT GCA Q Y G E T F T T L M T E Q A K L A S S G V A>
                                           17280
 ATT CCA GAG AGT CTG CAA CGC TCA ATG GAG CAA TTC CAC CAA CTA CAA GCG CAA ACA CTA CAA AGC I P E S L Q R S M E Q F H Q L Q A Q T L Q S>
                                  17340
 CAC ACC CAG TTC CTT GAG ATG CAA GCG GGT AGC AAC ATT GCA GCG TTA AAC CTA CTC AAT AGC AGC H T Q F L E M Q A G S N I A A L N L L N S S>
                                                        17420
                         17400
 CAA GCA ACT TAC GCT CCA GCC ATT CAC AAT GAA GCG ATT CAA AGC CAA GTG GTT CAA AGC CAA ACT Q A T Y A P A I H N E A I Q S Q V V Q S Q T>
 17540
 CCA AAA GCG CAG CCA GCA CCT GTG ACA ACT GCA GTT CAA ACT GCT CCG GCA CAA GTT GTT CGT CAA
  17600
  GCC GCA CCA GTT CAA GCC GCT ATT GAA CCG ATT AAT ACA AGT GTT GCG ACT ACA ACG CCT TCA GCC A A P V Q A A I E P I N T S V A T T T P S A>
                                                                                17700
                                                  17680
                     17660
```

Fig. 4

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TTC AGC GCC GAA ACA GCC CTG AGC GCA ACA AAA GTC CAA GCC ACT ATG CTT GAA GTG GTT GCT GAG
F S A E T A L S A T K V Q A T M L E V V A E>
                                                              17760
                                    17740
AAA ACC GGT TAC CCA ACT GAA ATG CTA GAG CTT GAA ATG GAT ATG GAA GCC GAT TTA GGC ATC GAT K T G Y P T E M L E L E M D M E A D L G I D>
                           17800
                                                       17820
TOT ATC AAG COT GTA GAA ATT CTT GGC ACA GTA CAA GAT GAG CTA CCG GGT CTA CCT GAG CTT AGC
 S I K R V E I L G T V Q D E L P G L P E L S>
                                              17880
                   17860
CCT GAA GAT CTA GCT GAG TGT CGA ACG CTA GGC GAA ATC GTT GAC TAT ATG GGC AGT AAA CTG CCG P E D L A E C R T L G E I V D Y M G S K L P>
           17920
                                      17940
GCT GAA GGC TCT ATG AAT TCT CAG CTG TCT ACA GGT TCC GCA GCT GCG ACT CCT GCA GCG AAT GGT
 AEGSMNSQLSTGSAAATPAANG>
   17980
                             18000
CTT TCT GCG GAG AAA GTT CAA GCG ACT ATG ATG TCT GTG GTT GCC GAA AAG ACT GGC TAC CCA ACT L S A E K V Q A T M M S V V A E K T G Y P T>
                    18060
GAA ATG CTA GAG CTT GAA ATG GAT ATG GAA GCC GAT TTA GGC ATA GAT TCT ATC AAG CGC GTT GAA E M L E L E M D M E A D L G I D S I K R V E>
ATT CTT GGC ACA GTA CAA GAT GAG CTA CCG GGT CTA CCT GAG CTT AGC CCT GAA GAT CTA GCT GAG
I L G T V Q D E L P G L P E L S P E D L A E>
TGT CGT ACT CTA GGC GAA ATC GTT GAC TAT ATG AAC TCT AAA CTC GCT GAC GGC TCT AAG CTG CCG C R T L G E I V D Y M N S K L A D G S K L P>
                                                     18280
GCT GAA GGC TCT ATG AAT TCT CAG CTG TCT ACA AGT GCC GCA GCT GCG ACT CCT GCA GCG AAT GGT A E G S M N S Q L S T S A A A A T P A A N G>
                                             18340
 CTC TCT GCG GAG AAA GTT CAA GCG ACT ATG ATG TCT GTG GTT GCC GAA AAG ACT GGC TAC CCA ACT
 L S A E K V Q A T M M S V V A E K T G Y P T>
                                     18400
 GAA ATG CTA GAA CTT GAA ATG GAT ATG GAA GCT GAC CTT GGC ATC GAT TCA ATC AAG CGC GTT GAA
 ATT CTT GGC ACA GTA CAA GAT GAG CTA CCG GGT TTA CCT GAG CTA AAT CCA GAA GAT TTG GCA GAG I L G T V Q D E L P G L P E L N P E D L A E>
          18520
                                             18540
 TGT CGT ACT CTT GGC GAA ATC GTG ACT TAT ATG AAC TCT AAA CTC GCT GAC GGC TCT AAG CTG CCA
C R T L G E I V T Y M N S K L A D G S K L P>
                                       18600
 GCT GAA GGC TCT ATG CAC TAT CAG CTG TCT ACA AGT ACC GCT GCT GCG ACT CCT GTA GCG AAT GGT
  A E G S M H Y Q L S T S T A A A T P V A N G>
                               18660
  CTC TCT GCA GAA AAA GTT CAA GCG ACC ATG ATG TCT GTA GTT GCA GAT AAA ACT GGC TAC CCA ACT
  L S A E K V Q A T M M S V V A D K T G Y P
                                                   18740
                                                                               18760
                       18720
  GAA ATG CTT GAA CTT GAA ATG GAT ATG GAA GCC GAT TTA EGT ATC GAT TCT ATC AAG CGC GTT GAA
  EMLELEMDNEADLGIDSIKRVE>
                           18800
  ATT CTT GGC ACA GTA CAA GAT GAG CTA CCG GGT TTA CCT GAG CTA AAT CCA GAA GAT CTA GCA GAG I L G T V Q D E L P G L P E L N P E D L A E>
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18840

Fig. 4 14/4:30

18880

WO 98/55625 PCT/US98/11639

19/106

MISSING AT THE TIME OF PUBLICATION

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20060
            20040
20020
   GTT AGC AAT GCG TTC TTG TGG GCC AAA TTA TTG CAA CCA AAG CTC GTT GCT GGA GCA GAT GCG CGT V S N A F L W A K L L Q P K L V A G A D A R>
                                             20120
    CGC TGT TTT GTA ACA GTA AGC CGT ATC GAC GGT GGC TTT GGT TAC CTA AAT ACT GAC GCC CTA AAA R C F V T V S R I D G G F G Y L N T D A L K>
                                      20180
    GAT GCT GAG CTA AAC CAA GCA GCA TTA GCT GGT TTA ACT AAA ACC TTA AGC CAT GAA TGG CCA CAA D A E L N Q A A L A G L T K T L S H E W P Q>
                                                         20260
  20220
                             20240
    GTG TTC TGT CGC GCG CTA GAT ATT GCA ACA GAT GTT GAT GCA ACC CAT CTT GCT GAT GCA ATC ACC
         FCRALDIATOVDATHLADAITS
                                               20320
                     20300
    AGT GAA CTA TTT GAT AGC CAA GCT CAG CTA CCT GAA GTG GGC TTA AGC TTA ATT GAT GGC AAA GTT S E L F D S Q A Q L P E V G L S L I D G K V>
                                        20380
    AAC CGC GTA ACT CTA GTT GCT GCT GAA GCT GCA GAT AAA ACA GCA AAA GCA GAG CTT AAC AGC ACA
                 T L V A A E A A D K T A K A E L N S T>
                                 20440
    GAT AAA ATC TTA GTG ACT GGT GGG GCA AAA GGG GTG ACA TTT GAA TGT GCA CTG GCA TTA GCA TCT
D K I L V T G G A K G V T F E C A L A L A S>
                        20500
    CGC AGC CAG TCT CAC TTT ATC TTA GCT GGG CGC AGT GAA TTA CAA GCT TTA CCA AGC TGG GCT GAG R S Q S H F I L A G R S E L Q A L P S W A E>
                                                          20600
                           20580
    GGT AAG CAA ACT AGC GAG CTA AAA TCA GCT GCA ATC GCA CAT ATT ATT TCT ACT GGT CAA AAG CCA
G K Q T S E L K S A A I A H I I S T G Q K P>
                                                               20660
     ACG CCT AAG CAA GTT GAA GCC GCT GTG TGG CCA GTG CAA AGC AGC ATT GAA ATT AAT GCC GCC CTA
T P K Q V E A A V W P V Q S S I E I N A A L>
                                                       20720
                           20700
     GCC GCC TTT AAC AAA GTT GGC GCC TCA GCT GAA TAC GTC AGC ATG GAT GTT ACC GAT AGC GCC GCA A A F N K V G A S A E Y V S M D V T D S A A>
                                                                         20800
                                              20780
                  20760
     ATC ACA GCA GCA CTT AAT GGT CGC TCA AAT GAG ATC ACC GGT CTT ATT CAT GGC GCA GGT GTA CTA
      I T A A L N G R S N E I T G L I H G A G V L>
                       20840
                                                    20860
     GCC GAC AAG CAT ATT CAA GAC AAG ACT CTT GCT GAA CTT GCT AAA GTT TAT GGC ACT AAA GTC AAC
                                                        20920 20940
                              20900
     GGC CTA ARA GCG CTG CTC GCG GCA CTT GAG CCA AGC ARA ATT ARA TTA CTT GCT ATG TTC TCA TCT G L K A L L A A L E P S K I K L L A M P S S>
                                                 20980
                       20960
      GCA GCA GGT TTT TAC GGT AAT ATC GGC CAA AGC GAT TAC GCG ATG TCG AAC GAT ATT CTT AAC AAG
      A A G F Y G N I G Q S D Y A M S N D I L N K>
                                                  .
                                                                     21060
               21020
                                         21040
     GCA GCG CTG CAG TTC ACC GCT CGC AAC CCA CAA GCT AAA GTC ATG AGC TTT AAC TGG GGT CCT TGG A A L Q F T A R N P Q A K V M S F N W G P W>
                21100
      21080
      GAT GGC GGC ATG GTT AAC CCA GCG CTT AAA AAG ATG TTT ACC GAG CGT GGT GTG TAC GTT ATT CCA
       D G G M V N P A L K K M F T E R G V Y V
                                                             21200
                        21160 21180
      CTA AAA GCA GGT GCA GAG CTA TTT GCC ACT CAG CTA TTG GCT GAA ACT GGC GTG CAG TTG CTC ATT L K A G A E L F A T Q L L A E T G V Q L L I>
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Fig. 4 16/30

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21240
GGT ACG TCA ATG CAA GGT GGC AGC GAC ACT AAA GCA ACT GAG ACT GCT TCT GTA AAA AAG CTT AAT
G T S M Q G G S D T K A T E T A S V K K L N>
                                        21300
GCG GGT GAG GTG CTA AGT GCA TCG CAT CCG CGT GCT GCA CAA AAA ACA CCA CTA CAA GCT GTC A G E V L S A S H P R A G A Q K T P L Q A V>
                                                                  21380
ACT GCA ACG CGT CTG TTA ACC CCA AGT GCC ATG GTC TTC ATT GAA GAT CAC CGC ATT GGC GGT AAC T A T R L L T P S A M V F I E D H R I G G N>
AGT GTG TTG CCA ACG GTA TGC GCC ATC GAC TGG ATG CGT GAA GCG GCA AGC GAC ATG CTT GGC GCT S V L P T V C A I D W M R E A A S D M L G A>
                                             21500
CAA GTT AAG GTA CTT GAT TAC AAG CTA TTA AAA GGC ATT GTA TTT GAG ACT GAT GAG CCG CAA GAG Q V K V L D Y K L L K G I V F E T D E P Q E>
                                                                      21580
                                  21560
TTA ACA CTT GAG CTA ACG CCA GAC GAT TCA GAC GAA GCT ACG CTA CAA GCA TTA ATC AGC TGT AAT L T L E L T P D D S D E A T L Q A L I S C N>
 GGG CGT CCG CAA TAC AAG GCG ACG CTT ATC AGT GAT AAT GCC GAT ATT AAG CAA CTT AAC AAG CAG
G R P Q Y K A T L I S D N A D I K Q L N K Q>
                                                 21700
 TTT GAT TTA AGC GCT AAG GCG ATT ACC ACA GCA AAA GAG CTT TAT AGC AAC GGC ACC TTG TTC CAC F D. L S A K A I T T A K E L Y S N G T L F H>

21740 21760 21780 21800
 GGT CCG CGT CTA CAA GGG ATC CAA TCT GTA GTG CAG TTC GAT GAT CAA GGC TTA ATT GCT AAA GTC G P R L Q G I Q S V V Q F D D Q G L I A K V>
                                                            21840
                            21820
 GCT CTG CCT AAG GTT GAA CTT AGC GAT TGT GGT GAG TTC TTG CCG CAA ACC CAC ATG GGT GGC AGT A L P K V E L S D C G E F L P Q T H M G G S>
                                                                                      21920
                                                   21900
 CAA CCT TTT GCT GAG GAC TTG CTA TTA CAA GCT ATG CTG GTT TGG GCT CGC CTT AAA ACT GGC TCG
Q P F A E D L L L Q A M L V W A R L K T G S>
                                         21960
  GCA AGT TTG CCA TCA AGC ATT GGT GAG TTT ACC TCA TAC CAA CCA ATG GCC TTT GGT GAA ACT GGT A S L P S S I G E F T S Y Q P M A F G E T G>
                                            22040
                              22020
  ACC ATA GAG CTT GAA GTG ATT AAG CAC AAC AAA CGC TCA CTT GAA GCG AAT GTT GCG CTA TAT CGT T I E L E V I K H N K R S L E A N V A L Y R>
                                   22100
  GAC AAC GGC GAG TTA AGT GCC ATG TTT AAG TCA GCT AAA ATC ACC ATT AGC AAA AGC TTA AAT TCA
D N G E L S A M F K S A K I T I S K S L N S>
                                                                               22180
                                              22160
  GCA TTT TTA CCT GCT GTC TTA GCA AAC GAC AGT GAG GCG AAT TAGTGGA ACAAACGCCT AAAGCTAGTG
A F L P A V L A N D S E A N>
                                                22240
   CG ATG CCG CTG CGC ATC GCA CTT ATC TTA CTG CCA ACA CCG CAG TTT GAA GTT AAC TCT GTC GAC M P L R I A L I L L P T P Q F E V N S V D>
                                                                       •
                      22280
                                                       22300
   CAG TCA GTA TTA GCC AGC TAT CAA ACA CTG CAG CCT GAG CTA AAT GCC CTG CTT AAT AGT GCG CCG Q S V L A S Y Q T L Q P E L N A L L N S A P>
                                                22360
    ACA CCT GAA ATG CTC AGC ATC ACT ATC TCA GAT GAT AGC GAT GCA AAC AGC TTT GAG TCG CAG CTA
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T P E M L S I T I S D D S D A N S
                                                22440
AAT GCT GCG ACC AAC GCA ATT AAC AAT GGC TAT ATC GTC AAG CTT GCT ACG GCA ACT CAC GCT TTG
N A A T N A I N N G Y I V K L A T A T H A
                                                                22520
                                       22500 +
TTA ATG CTG CCT GCA TTA AAA GCG GCG CAA ATG CGG ATC CAT CCT CAT GCG CAG CTT GCC GCT ATG
L M L P A L K A A Q M R I H P H A Q L A A M>
                                 22560
                       . .
CAG CAA GCT AAA TCG ACG CCA ATG AGT CAA GTA TCT GGT GAG CTA AAG CTT GGC GCT AAT GCG CTA
22640
                         22620
22600
AGC CTA GCT CAG ACT AAT GCG CTG TCT CAT GCT TTA AGC CAA GCC AAG CGT AAC TTA ACT GAT GTC
S L A Q T N A L S H A L S Q A K R N L T D
                            22700
                  22680
AGC GTG AAT GAG TGT TTT GAG AAC CTC AAA AGT GAA CAG CAG TTC ACA GAG GTT TAT TCG CTT ATT
        N E C F E N L K S E Q Q F T E V
                                  22760
CAG CAA CTT GCT AGC CGC ACC CAT GTG AGA AAA GAG GTT AAT CAA GGT GTG GAA CTT GGC CCT AAA
 Q Q L A S R T H V R K E V N Q G V E L G P
                         22820
CAA GCC AAA AGC CAC TAT TGG TTT AGC GAA TTT CAC CAA AAC CGT GTT GCT GCC ATC AAC TTT ATT Q A K S H Y W F S E F H Q N R V A A I N F I>
                   22880
                                              22900
AAT GGC CAA CAA GCA ACC AGC TAT GTG CTT ACT CAA GGT TCA GGA TTG TTA GCT GCG AAA TCA ATG N G Q Q A T S Y V L T Q G S G L L A A K S M>
                                      22960
CTA AAC CAG CAA AGA TTA ATG TTT ATC TTG CCG GGT AAC AGT CAG CAA CAA ATA ACC GCA TCA ATA L N Q Q R L M F I L P G N S Q Q Q I T A S I>
                               23020
ACT CAG TTA ATG CAG CAA TTA GAG CGT TTG CAG GTA ACT GAG GTT AAT GAG CTT TCT CTA GAA TGC T Q L M Q Q L E R L Q V T E V N E L S L E C>
                       23080
CAA CTA GAG CTG CTC AGC ATA ATG TAT GAC AAC TTA GTC AAC GCA GAC AAA CTC ACT ACT CGC GAT Q L E L L S I M Y D N L V N A D K L T T R D>
                23140
                                         23160
AGT AAG CCC GCT TAT CAG GCT GTG ATT CAA GCA AGC TCT GTT AGC GCA CAA GCA CAA GCA TTA AGC S K P A Y Q A V I Q A S S V S A A K Q E L S>
                               23220
TTA ATC CAA TAC AAA ACA CCG GGG GGC AGT TAC TTA ACC CTA ACA CCG CTT GGC AGC AAC AAT GAC L I Q Y K T P A G S Y L T L T P L G S N N D>
AAC GCC CAA GCG GGT CTT GCT TTT GTC TAT CCG GGT GGG GGA ACG GTT TAC GCC GAT ATG CTT AAT \cdot N A Q A G L A F V Y P G V G T V Y A D M L N>
                                    23420
CTA CAA GCA GAA GAT ATC TAT CAT CTT GAC CCT AAA CAT GCT GCC CAA ATG AGC TTA GGT GAC TTA L Q A E D I Y H L D P K H A A Q M S L G D L>
                                               23560
                                                                        23580
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-ig.4 18/30

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GCC ATT GCT GGC GTG GGG AGC AGC TAC CTG TTA ACT CAG CTG CTC ACC GAT GAG TTT AAT ATT AAG A I A G V G S S Y L L T Q L L T D E F N I R>
                                                23620
  CCT AAT TIT GCA TTA GGT TAC TCA ATG GGT GAA GCA TCA ATG TGG GCA AGC TTA GGC GTA TGG CAA
       N F A L G Y S H G E A S H W A S L G V W Q>
 AAC CCG CAT GCG CTG ATC AGC AAA ACC CAA ACC GAC CCG CTA TTT ACT TCT GCT ATT TCC GGC AAA N P H A L I S K T Q T D P L F T S A I S G K>
23720
  TTG ACC GCG GTT AGA CAA GCT TGG CAG CTT GAT GAT ACC GCA GCG GAA ATC CAG TGG AAT AGC TTT L T A V R Q A W Q L D D T A A E I Q W N. S F>
                                                    23820
                     23800
  GTG GTT AGA AGT GAA GCA GCG CCG ATT GAA GCC TTG CTA AAA GAT TAC CCA CAC GCT TAC CTC GCG
           R S E A A P I E A L L K D Y P H A Y L A>
                                          23880
  ATT ATT CAA GGG GAT ACC TGC GTA ATC GCT GGC TGT GAA ATC CAA TGT AAA GCG CTA CTT GCA GCA I I Q G D T C V I A G C E I Q C K A L L A A>
                                                               23960
                                  23940
  CTG GGT AAA CGC GGT ATT GCA GCT AAT CGT GTA ACG GCG ATG CAT ACG CAG CCT GCG ATG CAA GAG
L G K R G I A N R V T A M H T Q P A M Q E>
                                                      24020
  CAT CAA AAT GTG ATG GAT TTT TAT CTG CAA CCG TTA AAA GCA GAG CTT CCT AGT GAA ATA AGC TTT H Q N V M D F Y L Q P L K A E L P S E I S F>
                24060
  ATC AGC GCC GCT GAT TTA ACT GCC AAG CAA ACG GTG AGT GAG CAA GCA CTT AGC AGC CAA GTC GTT I S A A D L T A K Q T V S E Q A L S S Q V V>
                                    24140
  GCT CAG TCT ATT GCC GAC ACC TTC TGC CAA ACC TTG GAC TTT ACC GCG CTA GTA CAT CAC GCC CAA A Q S I A D T F C Q T L D F T A L V H H A Q>
   CAT CAA GGC GCT AAG CTG TTT GTT GAA ATT GGC GCG GAT AGA CAA AAC TGC ACC TTG ATA GAC AAG H Q G A K L F V E I G A D R Q N C T L I D K>
                                                  24280
   ATT GTT AAA CAA GAT GGT GCC AGC AGT GTA CAA CAT CAA CCT TGT TGC ACA GTG CCT ATG AAC GCA I V K Q D G A S S V Q H Q P C C T V P M N A>
                                                                       24360
                                         24340
   AAA GGT AGC CAA GAT ATT ACC AGC GTG ATT AAA GCG CTT GGC CAA TTA ATT AGC CAT CAG GTG CCA K G S Q D I T S V I K A L G Q L I S H Q V P>
                                                              24420
                               24400
   TTA TCG GTG CAA CCA TTT ATT GAT GGA CTC AAG CGC GAG CTA ACA CTT TGC CAA TTG ACC AGC CAA
    LSVQPFIDGLKRELTLCQLTSQ>
                                                      24480
    CAG CTG GCA GCA CAT GCA AAT GTT GAC AGC AAG TTT GAG TCT AAC CAA GAC CAT TTA CTT CAA GGG
     Q L A A H A N V D S K F E S N Q D H L L Q G>
                                             24540
    GAA GTC TA ATG TCA TTA CCA GAC AAT GCT TCT AAC CAC CTT TCT GCC AAC CAG AAA GGC GCA TCT
     24580
    CAG GCA AGT AAA ACC AGT AAG CAA AGC AAA ATC GCC ATT GTC GGT TTA GCC ACT CTG TAT CCA GAC
             S K T S K Q S K I A I V G L A T L Y P D>
                                                           24680
    GCT AAA ACC CCG CAA GAA TTT TGG CAG AAT TTG CTG GAT AAA CGC GAC TCT CGC AGC ACC TTA ACT
A K T P Q E F W Q N L L D K R D S R S T L T>
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24740 24760
 AAC GAA AAA CTC GGC GCT AAC AGC CAA GAT TAT CAA GGT GTG CAA GGC CAA TCT GAC CGT TTT TAT N E K L G A N S Q D Y Q G V Q G Q S D R F Y>
                                                     24820
                                         24800
 TGT AAT AAA GGC GGC TAC ATT GAG AAC TTC AGC TTT AATIGCT GCA GGC TAC AAA TTG CCG GAG CAA
                                                              24880
                              24860
  AGC TTA AAT GGC TTG GAC GAC AGC TTC CTT TGG GCG CTC GAT ACT AGC CGT AAC GCA CTA ATT GAT S L N G L D D S F L W A L D T S R N A L I D>
                                                    24940
  GCT GGT ATT GAT ATC AAC GGC GCT GAT TTA AGC CGC GCA GGT CTA GTC ATG GGC GCG CTG TCC TTC A G I D I N G A D L S R A G V V M G A L S F>
                                                                               25020
                                            25000
  CCA ACT ACC CGC TCA AAC GAT CTG TTT TTG CCA ATT TAT CAC AGC GCC GTT GAA AAA GCC CTG CAA
   PTTRSNDLFLPIYHSAVEKALQ>
             25060
                                                                   25080
  GAT AAA CTA GGC GTA AAG GCA TTT AAG CTA AGC CCA ACT AAT GCT CAT ACC GCT CGC GCG GCA AAT D K L G V K A F K L S P T N A H T A R A A N>
                                                          25140
                       25120
  GAG AGC AGC CTA AAT GCA GCC AAT GGT GCC ATT GCC CAT AAC AGC TCA AAA GTG GTG GCC GAT GCA E S S L N A A N G A I A H N S S K V V A D A>
                                                25200
  CTT GGC CTT GGC GGC GCA CAA CTA AGC CTA GAT GCT GCC TGT GCT AGT TCG GTT TAC TCA TTA AAG
L G L G G A Q L S L D A A C A S S V Y S L K>
                                                                 25280
                                      25260
  CTT GCC TGC GAT TAC CTA AGC ACT GGC AAA GCC GAT ATC ATG CTA GCA GGC GCA GTA TCT GGC GCG L A C D Y L S T G K A D I M L A G A V S G A>
                                                             25340
   GAT CCT TTC TTT ATT AAT ATG GGA TTC TCA ATC TTC CAC GCC TAC CCA GAC CAT GGT ATC TCA GTA D P F F I N M G F S I F H A Y P D H G I S V>
                                                     25400
  CCG TTT GAT GCC AGC AGT AAA GGT TTG TTT GCT GGC GAA GGC GCT GGC GTA TTA GTG CTT AAA CGT P F D A S S K G L F A G E G A G V L V L K R>
                                         25460
   CTT GAA GAT GCC GAG CGC GAC AAT GAC AAA ATC TAT GCG GTT GTT AGC GGC GTA GGT CTA TCA AAC L E D A E R D N D K I Y A V V S G V G L S N>
25500
                                 25520
   CAC GGT AAA GGC CAG TTT GTA TTA AGC CCT AAT CCA AAA GGT CAG GTG AAG GCC TTT GAA CGT GCT D G K G Q F V L S P N P K G Q V K A F E R A>
                                    25600
    TAT GCT GCC AGT GAC ATT GAG CCA AAA GAC ATT GAA GTG ATT GAG TGC CAC GCA ACA GGC ACA CCG
Y A A S D I E P K D I E V I E C H A T G T P>
    CTT GGC GAT AAA ATT GAG CTC ACT TCA ATG GAA ACC TTC TTT GAA GAC AAG CTG CAA GGC ACC GAT L G D K L Q G T D>
    GCA CCG TTA ATT GGC TCA GCT AAG TCT AAC TTA GGC CAC CTA TTA ACT GCA GCG CAT GCG GGG ATC A P L I G S A K S N L G H L L T A A H A G I>
                                                          25800 •
    ATG AAG ATC ATC TTC GCC ATG AAA GAA GGT TAC CTG CCG CCA AGT ATC AAT ATT AGT GAT GCT ATC M K M I F A M K E G Y L P P S I N I S D A I>
                                                                                   25880
     GCT TCG CCG AAA AAA CTC TTC GGT AAA CCA ACC CTG CCT AGC ATG GTT CAA GGC TGG CCA GAT AAG
A S P K K L F G K P T L P S M V Q G W P D K>
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Fig. 4 20/30

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25920
  25900
CCA TCG AAT AAT CAT TTT GGT GTA AGA ACC CGT CAC GCA GGC GTA TCG GTA TTT GGC TTT GGT GGC P S N N H F G V R T R H A G V S V P G F G G>
                            25980
TGT AAC GCC CAT CTG TTG CTT GAG TCA TAC AAC GGC AAA GGA ACA GTA AAG GCA GAA GCC ACT CAA
C N A H L L L E S Y N G K G T V K A E A T Q>
                                                      26060
GTA CCG CGT CAA GCT GAG CCG CTA AAA GTG GTT GGC CTT GCC TCG CAC TTT GGG CCT CTT AGC AGC
      PRQAEPLK V V G L A S H F G P L S S>
ATT AAT GCA CTC AAC AAT GCT GTG ACC CAA GAT GGG AAT GGC TTT ATC GAA CTG CCG AAA AAG CGC I N A L N N A V T Q D G N G F I E L P K K R>
                                 26180
TGG AAA GGC CTT GAA AAG CAC AGT GAA CTG TTA GCT GAA TTT GGC TTA GCA TCT GCG CCA AAA GGT W K G L E K H S E L L A E F G L A S A P K G>
GCT TAT GTT GAT AAC TTC GAG CTG GAC TTT TTA CGC TTT AAA CTG CCG CCA AAC GAA GAT GAC CGT A Y V D N F E L D F L R F K L P P N E D D R>
TTG ATC TCA CAG CAG CTA ATG CTA ATG CGA GTA ACA GAC GAA GCC ATT CGT GAT GCC AAG CTT GAG L I S Q Q L M L M R V T D E A I R D A K L E>
                                                                         26400
 CCG GGG CAA AAA GTA GCT GTA TTA GTG GCA ATG GAA ACT GAG CTT GAA CTG CAT CAG TTC CGC GGC P G Q K V A V L V A M E T E L E L H Q F R G>
                                                              26460
 CGG GTT AAC TTG CAT ACT CAA TTA GCG CAA AGT CTT GCC GCC ATG GGC GTG AGT TTA TCA ACG GAT R V N L H T Q L A Q S L A A M G V S L S T D>
                                                   26520
 GAA TAC CAA GCG CTT GAA GCC ATC GCC ATG GAC AGC GTG CTT GAT GCT GCC AAG CTC AAT CAG TAC E Y Q A L E A I A M D S V L D A A K L N Q Y>
                                        26580
 ACC AGC TTT ATT GGT AAT ATT ATG GCG TCA CGC GTG GCG TCA CTA TGG GAC TTT AAT GGC CCA GCC T S F I G N I M A S R V A S L W D F N G P A>
TTC ACT ATT TCA GCA GCA GAG CAA TCT GTG AGC CGC TGT ATC GAT GTG GCG CAA AAC CTC ATC ATG F T I S A A E Q S V S R C I D V A Q N L I M>
                   26700
                                                        26720
                                                                                           26740
 GAG GAT AAC CTA GAT GCG GTG GTG ATT GCA GCG GTC GAT CTC TCT GGT AGC TTT GAG CAA GTC ATT E D N L D A V V I A A V D L S G S F E Q V I>
 CTT AAA AAT GCC ATT GCA CCT GTA GCC ATT GAG CCA AAC CTC GAA GCA AGC CTT AAT CCA ACA TCA L K N A I A P V A I E P N L E A S L N P T S>
                                                                      26860
  GCA AGC TGG AAT GTC GGT GAA GGT GCT GGC GCG GTC GTG CTT GTT AAA AAT GAA GCT ACA TCG GGC A S W N V G E G A G A V V L V K N E A T S G>
                                                           26920
  TGC TCA TAC GGC CAA ATT GAT GCA CTT GGC TTT GCT AAA ACT GCC GAA ACA GCG TTG GCT ACC GAC C S Y G Q I D A L G F A K T A E T A L A T D>
                                                           27000
                                                26980
             26960
  AAG CTA CTG AGC CAA ACT GCC ACA GAC TTT AAT AAG GTT AAA GTG ATT GAA ACT ATG GCA GCG CCT K L \dot{S} Q T A T D F N K V K V I E T M A A P>
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GCT AGC CAA ATT CAA TTA GCG CCA ATA GTT AGC TCT CAA GTG ACT CAC ACT GCT GCA GAG CAG CGT

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S Q I Q L A P I V S S Q V T H T A A E
 GTT GGT CAC TGC TTT GCT GCA GCG GGT ATG GCA AGC CTA TTA CAC GGC TTA CTT AAC TTA AAT ACT V G K C F A A A G M A S L L H G L U N L N T>
                                             27180
                                                                             27200
  GTA GCC CAA ACC AAT AAA GCC AAT TGC GCG CTT ATC AAC AAT ATC AGT GAA AAC CAA TTA TCA CAG
V A Q T N K A N C A L I N N I S E N Q L S Q>
                                                                     27260
  CTG TTG ATT AGC CAA ACA GCG AGC GAA CAA CAA GCA TTA ACC GCG CGT TTA AGC AAT GAG CTT AAA L L I S Q T A S E Q Q A L T A R L S N E L K>
                                                           27320
  TCC GAT GCT AAA CAC CAA CTG GTT AAG CAA GTC ACC TTA GGT GGC CGT GAT ATC TAC CAG CAT ATT S D A K H Q L V K Q V T L G G R D I Y Q H I>
                                                  27380
  GTT GAT ACA CCG CTT GCA AGC CTT GAA AGC ATT ACT CAG AAA TTG GCG CAA GCG ACA GCA TCG ACA V D T P L A S L E S I T Q K L A Q A T A S T>
                                       27440
. GTG GTC AAC CAA GTT AAA CCT ATT AAG GCC GCT GGC TCA GTC GAA ATG GCT AAC TCA TTC GAA ACG
         V N Q V K P I K A A G S V E M A N S F E T>
                                                            27520
                              27500
  GAA AGC TCA GCA GAG CCA CAA ATA ACA ATT GCA GCA CAA CAG ACT GCA AAC ATT GGC GTC ACC GCT
  ESSAEPQITIAAQQTANIGV
                                                   27580
                     27560
  CAG GCA ACC AAA CGT GAA TTA GGT ACC CCA CCA ATG ACA ACA AAT ACC ATT GCT AAT ACA GCA AAT Q A T K R E L G T P P M T T N T I A N T A N>
                                    27640
  AAT TTA GAC AAG ACT CTT GAG ACT GTT GCT GGC AAT ACT GTT GCT AGC AAG GTT GGC TCT GGC GAC N L D K T L E T V A G N T V A S K V G S G D>
                                                                  27720
  ATA GTC AAT TTT CAA CAG AAC CAA CAA TTG GCT CAA CAA GCT CAC CTC GCC TTT CTT GAA AGC CGC I V N F Q Q N Q Q L A Q Q A H L A F L E S R>
                      27760
                                                        27780
  AGT GCG GGT ATG AAG GTG GCT GAT GCT TTA TTG AAG CAA CAG CTA GCT CAA GTA ACA GGC CAA ACT S A G M K V A D A L L K Q Q L A Q V T G Q T>
                                               27840
                                                                               27860
              27820
  ATC GAT AAT CAG GCC CTC GAT ACT CAA GCC GTC GAT ACT CAA ACA ACA ACC GAG AAT GTA GCG ATT GCC I D N Q A L D T Q A V D T Q T S E N V A I A>
                                     27900
   GCA GAA TCA CCA GTT CAA GTT ACA ACA CCT GTT CAA GTT ACA ACA CCT GTT CAA ATC AGT GTT GTG A E S P V Q V T T P V Q V T T P V Q I S V V>
                                                            27980
   GAG TTA AAA CCA GAT CAC GCT AAT GTG CCA CCA TAC ACG CCG CCA GTG CCT GCA TTA AAG CCG TGT E L K P D H A N V P P Y T P P V P A L K P C>
                                                   28040
   ATC TGG AAC TAT GCC GAT TTA GTT GAG TAC GCA GAA GGC GAT ATC GCC AAG GTA TTT GGC AGT GAT I W N Y A D L V E Y A E G D I A K V F G 5 D>
                                         28100
                                                                         28120
   TAT GCC ATT ATC GAC AGC TAC TCG CGC CGC GTA CGT CTA CCG ACC ACT GAC TAC CTG TTG GTA TCG
    Y A I I D S Y S R R V R L • P T T D Y L L V S>
                                            29180
   CGC GTG ACC AAA CTT GAT GCG ACC ATC AAT CAA TTT AAG CCA TGC TCA ATG ACC ACT GAG TAC GAC R V T K L D A T I N Q F K P C S M T T E Y D>
                                                      28240
                       28220
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ATC CCT GTT GAT GCG CCG TAC TTA GTA GAC GGA CAA ATC CCT TGG GCG GTA GCA GTA GAA TCA GGC I P V D A V E S G>
                                                                                      28320
                                               28300
CAA TGT GAC TTG ATG CTT ATT AGC TAT CTC GGT ATC GAC TTT GAG AAC AAA GGC GAG CGG GTT TAT Q C D L M L I S Y L G I D F E N K G E R V Y>
                                                                    28380
                                    28360
CGA CTA CTC GAT TGT ACC CTC ACC TTC CTA GGC GAC TTG CCA CGT GGC GGA GAT ACC CTA CGT TAC R L L D C T L T F L G D L P R G G D T L R Y>
                         28420
GAC ATT AAG ATC AAT AAC TAT GCT CGC AAC GGC GAC ACC CTG CTG TTC TTC TCG TAT GAG TGT D I K I N N Y A R N G D T L L F F F S Y E C>
                                                                                           28520
                                                    28500
              28480
TTT GTT GGC GAC AAG ATG ATC CTC AAG ATG GAT GGC GGC TGC GCT GGC TTC TTC ACT GAT GAA GAG F V G D K M I L K M D G G C A G F F T D E E>
                                                                               28580
CTT GCC GAC GGT AAA GGC GTG ATT CGC ACA GAA GAA GAG ATT AAA GCT CGC AGC CTA GTG CAA AAG
L A D C K G V I R T E E E I K A R S L V Q K>
                                          28640
                             28620
CAA CGC TTT AAT CCG TTA CTA GAT TGT CCT AAA ACC CAA TTT AGT TAT GGT GAT ATT CAT AAG CTA
Q R F N P L L D C P K T Q F S Y G D I H K L>

28680 28700 28720
                                                      28700
TTA ACT GCT GAT ATT GAG GGT TGT TTT GGC CCA AGC CAC AGT GGC GTC CAC CAG CCG TCA CTT TGT L T A D I E G C F G P S H S G V H Q P S L C>
                              28760
 TTC GCA TCT GAA AAA TTC TTG ATG ATT GAA CAA GTC AGC AAG GTT GAT CGC ACT GGC GGT ACT TGG F A S E K F L M I E Q V S K V D R T G G T W>
                   28820
                                                                      28840
 GGA CTT GGC TTA ATT GAG GGT CAT AAG CAG CTT GAA GCA GAC CAC TGG TAC TTC CCA TGT CAT TTC G L G L I E G H K Q L E A D H W Y F P C H F>
 AAG GGC GAC CAA GTG ATG GCT GGC TCG CTA ATG GCT GAA GGT TGT GGC CAG TTA TTG CAG TTC TAT

K G D Q V M A G S L M A E G C G Q L L Q F Y> ... 224444 1 28940 28960 28960 London 1 28960
 ATG CTG CAC CTT GGT ATG CAT ACC CAA ACT AAA AAT GGT CGT TTC CAA CCT CTT GAA AAC GCC TCA
M 'L H L G M' H T Q T K N G R F Q P L E N A S>
                                                                           29040
 CAG CAA GTA CGC TGT CGC GGT CAA GTG CTG CCA CAA TCA GGC GTG CTA ACT TAC CGT ATG GAA GTG Q Q V R C R G Q V L P Q S G V L T Y R M E V>
                                                                                                        29120
                                                                 29100
  ACT GAA ATC GGT TTC AGT CCA CGC CCA TAT GCT AAA GCT AAC ATC GAT ATC TTG CTT AAT GGC AAA
T E I G F S P R P Y A K A N I D I L L N G K>
                                                      29160
  GCG GTA GTG GAT TTC CAA AAC CTA GGG GTG ATG ATA AAA GAG GAA GAT GAG TGT ACT CGT TAT CCA
A V V D F Q N L G V M I K E E D E C T R Y P>
                                           29220
                                                                                 29240
  CTT TTG ACT GAA TCA ACA ACG GCT AGC ACT GCA CAA GTA AAC GCT CAA ACA AGT GCG AAA AAG GTA L L T E S T T A S T A Q V N A Q T S A K K V>
                                                                      29300
  TAC AAG CCA GCA TCA GTC AAT GCG CCA TTA ATG GCA CAA ATT CCT GAT CTG ACT AAA GAG CCA AAC
Y K P A S V N A P L M A Q I P D L T K E P N>
                                                           29360
  AAG GGC GTT ATT CCG ATT TCC CAT GTT GAA GCA CCA ATT ACG CCA GAC TAC CCG AAC CGT GTA CCT K G V I P I S H V E A P I T P D Y P N R V P>
                                                                                     29440
                                                29420
          29400
```

GAT ACA GTG CCA TTC ACG CCG TAT CAC ATG TTT GAG TTT GCT ACA GGC AAT ATC GAA AAC TGT TTC
D T V P F T P Y H H F E F A T G N T B N C F >

9480 29500 29520 29500 9480 GGG CCA GAG TTC TCA ATC TAT CGC GGC ATG ATC CCA CGT ACA CCA TGC GGT GAC TTA CAA GTG
G P E F S I Y R G M I P P R T P C G D L Q V>

29540 29560 29580 ACC ACA CGT GTG ATT GAA GTT AAC GGT AAG CGT GGC GAC TTT AAA AAG CCA TCA TCG TGT ATC GGT T T R V I E V N G K R G D F K K P S S C I A> 29620 GAA TAT GAA GTG CCT GCA GAT GCG TGG TAT TTC GAT AAA AAC AGC CAC GGC GCA GTG ATG CCA TAT E Y E V P A D A W Y F D K N S H G A V H P Y>

29660 29700 29720 TCA ATT TTA ATG GAG ATC TCA CTG CAA CCT AAC GGC TTT ATC TCA GGT TAC ATG GGC ACA ACC CTA
S I L N E I S L Q P N G F I S G Y N G T T L> 29740 GGC TTC CCT GGC CTT GAG CTG TTC TTC CGT AAC TTA GAC GGT AGC GGT GAG TTA CTA CGT GAA GTA
G F P G L E L F F R N L D G S G E L L R E V>

29800 29840 GAT TTA CGT GGT AAA ACC ATC CGT AAC GAC TCA CGT TTA TTA TCA ACA GTG ATG GCC GGC ACT AAC
D L R G K T I R N D S R L L S T V M A G T N>

29860 29880 29900 29920 29900 ATC ATC CAA AGC TTT AGC TTC GAG CTA AGC ACT GAC GGT GAG CCT TTC TAT CGC GGC ACT GCG GTA
I I Q S F S F E L S T D G E P F Y R G T A V> 29960 TTT GGC TAT TTT AAA GGT GAC GCA CTT AAA GAT CAG CTA GGC CTA GAT AAC GGT AAA GTC ACT CAG F G Y F K G D A L K D Q L G L D N G K V T Q> 30000 30020 30040 CCA TGG CAT GTA GCT AAC GGC GTT GCT GCA AGC ACT AAG GTG AAC CTG CTT GAT AAG AGC TGC CGT P W H V A N G V A A S T K V N L L D K S C R> 30080 CAC TIT AAT GCG CCA GCT AAC CAG CCA CAC TAT CGT CTA GCC GGT GGT CAG CTG AAC TIT ATC GAC H F N A P A N Q P H Y R L A G G Q L N F I D> 30140 30160 30180 AGT GTT GAA ATT GTT GAT AAT GGC GGC ACC GAA GGT TTA GGT TAC TTG TAT GCC GAG CGC ACC ATT S V E I V D N G G T E G L G Y L Y A E R T I> 30220 GAC CCA AGT GAT TGG TTC TTC CAG TTC CAC TTC CAC CAA GAT CCG GTT ATG CCA GGC TCC TTA GGT D P S D W F F Q F H F H Q D P V M P G S L G> W FAGA GTT GAA GCA ATT ATT GAA ACC ATG CAA GCT TAC GCT ATT AGT AAA GAC TTG GGC GCA GAT TTC AAA V E A I I E T M Q A Y A I S K D L G A D F K> 30360 AAT CCT AAG TTT GGT CAG ATT TTA TCG AAC ATC AAG TGG AAG TAT CGC GGT CAA ATC AAT CCG CTG
N P K F G Q I L S N I K W K Y R G Q I N P L> 30420 AAC AAG CAG ATG TCT ATG GAT GTC AGC ATT ACT TCA ATC AAA GAT GAA GAC GGT AAG AAA GTC ATC N K Q M S M D V S I T S I K D E D G K K V I> 30480 ACA GGT AAT GCC AGC TTG AGT AAA GAT GGT CTG CGC ATA TAC GAG GTC TTC GAT ATA GCT ATC AGC
T G N A S L S K D G L R I Y E V F D I A I S> 10520

30520

30540

30560

30580 ATC GAA GAA TOT GTA T AAATCGGAGT GACTGTCTGG CTATTTTACT CAATTTCTGT GTCAAAAGTG CTCACCTATA 24/30

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30640
                                                                              30660
                            30620
TTCATAGGCT GCGCGCTTTT TTCTGGAAAT TGAGCAAAAG TATCTGCGTC CTAACTCGAT TTATAAGAAT GGTTTAATTG
                                            30720
AAAAGAACAA CAGCTAAGAG CCGCAAGCTC AATATAAATA ATTAAGGGTC TTACAAATA ATG AAT CCT ACA GCA ACT
                                        30780
AAC GAA ATG CTT TCT CCG TGG CCA TGG GCT GTG ACA GAG TCA AAT ATC AGT TTT GAC GTG CAA GTG N E M L S P W P W A V T E S N I S F D V Q V>
                               30840
ATG GAA CAA CAT AAA GAT TTT AGC CGG GCA TGT TAC GTG GTC AAT CAT GCC GAC CAC GGC TTT M E Q Q L K D F S R A C Y V V N H A D H G F>
                                                       30920
                        30900
GGT ATT GCG CAA ACT GCC GAT ATC GTG ACT GAA CAA GCG GCA AAC AGC ACA GAT TTA CCT GTT AGT G I A Q T A D I V T E Q A A N S T D L P V S>
                                              30980
               30960
GCT TTT ACT CCT GCA TTA GGT ACC GAA AGC CTA GGC GAC AAT AAT TTC CGC CGC GTT CAC GGC GTT
 AFTPALGTESLGDNNFRRVHGV>
   31020
                                     31040
AAA TAC GCT TAT TAC GCA GGC GCT ATG GCA AAC GGT ATT TCA TCT GAA GAG CTA GTG ATT GCC CTA
K Y A Y Y A G A M A N G I S S E E L V I A L>
                           31100
GGT CAA GCT GGC ATT TTG TGT GGT TCG TTT GGA GCA GCC GGT CTT ATT CCA AGT CGC GTT GAA GCG
G Q A G I L C G S F G A A G L I P S R V E A>
 GCA ATT AAC CGT ATT CAA GCA GCG CTG CCA AAT GGC CCT TAT ATG TTT AAC CTT ATC CAT AGT CCT A I N R I Q A A L P N G P Y M F N L I H S P>
 AGC GAG CCA GCA TTA GAG CGT GGC AGC GTA GAG CTA TTT TTA AAG CAT AAG GTA CGC ACC GTT GAA S E P A L E R G S V E L F L R H K V R T V E>
                                                         31320
                                 31300
 GCA TCA GCT TTC TTA GGT CTA ACA CCA CAA ATC GTC TAT TAC CGT GCA GCA GGA TTG AGC CGA GAC A S A F L G L T P Q I V Y Y R A A G L S R D>
                                                    31380
                      31360
 GCA CAA GGT AAA GTT GTG GTT GGT AAC AAG GTT ATC GCT AAA GTA AGT CGC ACC GAA GTG GCT GAA
A Q G K V V V G N K V I A K V S R T E V A E>
 AAG TTT ATG ATG CCA GCG CCC GCA AAA ATG CTA CAA AAA CTA GTT GAT GAC GGT TCA ATT ACC GCT
  K F M M P A P A K M L Q K L V D D
                                  31500
                                                                  31520
 GAG CAA ATG GAG CTG GCG CAA CTT GTA CCT ATG GCT GAC GAC ATC ACT GCA GAG GCC GAT TCA GGT
       Q M E L A Q L V P M A D D I T A E A D S G>
                         31560
 GGC CAT ACT GAT AAC CGT CCA TTA GTA ACA TTG CTG CCA ACC ATT TTA GCG CTG AAA GAA GAA ATT G H T D N R P L V T L L P T I L A L K E E I>
                                                31640
  CAA GCT AAA TAC CAA TAC GAC ACT CCT ATT CGT GTC GGT TGT GGT GGC GGT GTG GGT ACG CCT GAT Q A K Y Q Y D T P I R V G C G G G V G T P D>
                                       31700
                                                                      31720
  GCA GCG CTG GCA ACG TTT AAC ATG GGC GCG GCG TAT ATT GTT ACC GGC TCT ATC AAC CAA GCT TGT
  A A L A T F N M G A A Y I V T G S I N Q A C>
                                                             31780
31740
                              31760
  GTT GAA GCG GGC GCA AGT GAT CAC ACT CGT AAA TTA CTT GCC ACC ACT GAA ATG GCC GAT GTG ACT V E A G A S D H T R K L L A T T E M A D V T>
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Fig.4 25/30

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31860
                             31840
ATG GCA CCA GCT GCA GAT ATG TTC GAG ATG GGC GTA AAA CTG CAG GTG GTT AAG CGC GGC ACG CTA
                  31900
TTC CCA ATG CGC GCT AAC AAG CTA TAT GAG ATC TAC ACG CGT TAC GAT TCA ATC GAA GCG ATC CCA
FPMRANKLYEIYTRYDSIEAIP>
                                               31980
                        31960
TTA GAC GAG CGT GAA AAG CTT GAG AAA CAA GTA TTC CGC TCA AGC CTA GAT GAA ATA TGG GCA GGT
 L D E R E K L E K Q V F R S S L D E I W A G>
                                        32040
ACA GTG GCG CAC TTT AAC GAG CGC GAC CCT AAG CAA ATC GAA CGC GCA GAG GGT AAC CCT AAG CGT
T V A H F N E R D P K Q I E R A E G N P K R>
                                 32100
         32080
AAA ATG GCA TTG ATT TTC CGT TGG TAC TTA GGT CTT TCT AGT CGC TGG TCA AAC TCA GGC GAA GTG
    MALIFRWYLGLSSRWSNSGEV>
              32160
GGT CGT GAA ATG GAT TAT CAA ATT TGG GCT GGC CCT GCT CTC GGT GCA TTT AAC CAA TGG GCA AAA
 G R E M D Y Q I W A G P A L G A F N Q W A K>
                                            32240
                  32220
GGC AGT TAC TTA GAT AAC TAT CAA GAC CGA AAT GCC GTC GAT TTG GCA AAG CAC TTA ATG TAC GGC G S Y L D N Y Q D R N A V D L A K H L M Y G>
                                    32300
GCG GCT TAC TTA AAT CGT ATT AAC TCG CTA ACG GCT CAA GGC GTT AAA GTG CCA GCA CAG TTA CTT A A Y L N R I N S L T A Q G V K V P A Q L L>
              32360 32380 32400
CGC TGG AAG CCA AAC CAA AGA ATG GCC TA ATACACTTAC AAAGCACCAG TCTAAAAAAGC CACTAATCTT
                       32440
                                            32460
              32420
GATTAGTGGC TTTTTTTATT GTGGTCAATA TGAGGCTATT TAGCCTGTAA GCCTGAAAAT ATCAGCACTC TGACTTTACA
                                                     32540
AGCAAATTAT AATTAAGGCA GGGCTCTACT CATTTATACT GCTAGCAAAC AAGCAAGTTG CCCAGTAAAA CAACAAGGTA
                                 32600
CCTGATTTAT ATCGTCATAA AAGTTGGCTA GAGATTCGTT ATTGATCTTT ACTGATTAGA GTCGCTCTGT TTGGAAAAAG
                                  32680
 GTTTCTCGTT ATCATCAAAA TACACTCTCA AACCTTTAAT CAATTACAAC TTAGGCTTTC TGCGGGCATT TTTATCTTAT
             32740 32760 32780
 TTGCCACAGC TGTATTTGCC TTTAGGTTTT GGGTGCAACT ACCATTAATT GAGGCCTCAT TAGTTAAATT ATCTGAGCAA
 GAGCTCACCT CTTTAAATTA CGCTTTTCAG CAA ATG AGA AAG CCA CTA CAA ACC ATT AAT TAC GAC TAT GCG M R K P L Q T I N Y D Y A>
 GTG TGG GAC AGA ACC TAC AGC TAT ATG AAA TCA AAC TCA GCG AGC GCT AAA AGG TAC TAT GAA AAA V W D R T Y S Y M K S N S A S A K R Y Y E K>
                      32960
 CAT GAG TAC CCA GAT GAT ACG TTC AAG AGT TTA AAA GTC GAC GGA GTA TTT ATA TTC AAC CGT ACA
H E Y P D D T F K S L K V D G V F I F N R T>
                          33040
                                                              33060
 AAT CAG CCA GTT TTT AGT AAA GGT TTT AAT CAT AGA AAT GAT ATA CCG CTG GTC TTT GAA TTA ACT N Q P V F S K G F N H R N D I P L V F E L T>
                  33100
                                                       33120
 GAC TTT AAA CAA CAT CCA CAA AAC ATC GCA TTA TCT CCA CAA ACC AAA CAG GCA CAC CCA CCG GCA D F K Q H P Q N I A L S P Q T K Q A H P P A>
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Fig. 4 26/30

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33160
                                               33180
33140
AGT AAG CCG TTA GAC TCC CCT GAT GAT GTG CCT TCT ACC CAT GGG GTT ATC GCC ACA CGA TAC GGT S K P L D S P D D V P S T H G V I A T R Y G>
                                         33240
               33220
33300
GTC TTC ATT AGG TTA ATT GAT GAA TGG TTC ATC GCT GAG CTA TCG CAA TAC ACT GCC GCA GGT GTT V F I R L I D E W F I A E L S Q Y T A A G V>
                           33360
GAA ATC GCT ATG GCT GAT GCC GCA GAC GCA CAA TTA GCG AGA TTA GGC GCA AAC ACT AAG CTT AAT
E I A M A D A A D A Q L A R L G A N T K L N>
                                            33440
                   33420
AAA GTA ACC GCT ACA TCC GAA CGG TTA ATA ACT AAT GTC GAT GGT AAG CCT CTG TTG AAG TTA GTG
 K V T A T S E R L I T N V D G K P L L K L V>
                    33500
            33480
CTT TAC CAT ACC AAT AAC CAA CCG CCG CCG ATG CTA GAT TAC AGT ATA ATA ATT CTA TTA GTT GAG L Y H T N N Q P P P H L D Y S I I I L L V E>
             33560
 ATG TCA TTT TTA CTG ATC CTC GCT TAT TTC CTT TAC TCC TAC TTC TTA GTC AGG CCA GTT AGA AAG M S F L L I L A Y F L Y S Y F L V R P V R K>
                                     33640
 CTG GCT TCA GAT ATT AAA AAA ATG GAT AAA AGT CGT GAA ATT AAA AAG CTA AGG TAT CAC TAC CCT
L A S D I K K M D K S R E I K K L R Y H Y P>
                                      33700
 ATT ACT GAG CTA GTC AAA GTT GCG ACT CAC TTC AAC GCC CTA ATG GGG ACG ATT CAG GAA CAA ACT I T E L V K V A T H F N A L M G T I Q E Q T>
                  33760
 AAA CAG CTT AAT GAA CAA GTT TTT ATT GAT AAA TTA ACC AAT ATT CCC AAT CGT CGC GCT TTT GAG
  K Q L N E Q V F I D K L T N I P N R R A F E>
                                                 33840
                        33820
 CAG CGA CTT GAA ACC TAT TGC CAA CTG CTA GCC CGG CAA CAA ATT GGC TTT ACT CTC ATC ATT GCC Q R L E T Y C Q L L A R Q Q I G F T L I I A>
                                         33900
 GAT GTG GAT CAT TIT AAA GAG TAC AAC GAT ACT CTT GGG CAC CTT GCT GGG GAT GAA GCA TTA ATA
  D V D H F K E Y N D T L G H L A G D E A L I>
                                                           33980
                                   33960
  AAA GTG GCA CAA ACA CTA TCG CAA CAG TTT TAC CGT GCA GAA GAT ATT TGT GCC CGT TTT GGT GGT K V A Q T L S Q Q F Y R A E D I C A R F G G>
                                    34040
            34020
   34000
  GAA GAA TIT ATT ATG TTA TTT CGA GAC ATA CCT GAT GAG CCC TTG CAG AGA AAG CTC GAT GCG ATG
  EEFIM LFRDIP DEPLQRKLDAM>
                                              34100
                     34080
  CTG CAC TCT TTT GCA GAG CTC AAC CTA CCT CAT CCA AAC TCA TCA ACC CCT AAT TAC GTT ACT GTG L H S F A E L N L P H P N S S T A N Y V T V>
                                      34160 34180
  AGC CTT GGG GTT TGC ACA GTT GTT GCT GTT GAT GTT GAA TTT AAA AGT GAG TCG CAT ATT ATT
   S L G V C T V V A V D D F E F K S E S H I I>
                               34220
                                                        34240
       34200
  GGC AGT CAG GCT GCA TTA ATC GCA GAT AAG GCG CTT TAT CAT GCT AAA GCC TGT GGT CGT AAC CAG
       S Q A A L I A D K A L Y H A K A C G R N Q>
                                  34300
                    34280
34260
  TTG TCA AAA ACT ACT ATT ACT GTT GAT GAG ATT GAG CAA TTA GAA GCA AAT AAA ATC GGT CAT CAA
```

Fig. 4 27/30

L s	ĸ	т	т і	T	v I	D E I	E Q L	EAN	кі	: н Q>
	•	3434	10			34360		34380		34400
	ACTO		. •	TTTCC	C CTA	AGTCAGA GCT	ATTTGCC ACT	TCAAGAT GTG	GCTACAA GGC	TACTCT
A>		-	24420			34440		34460		34480
mm(1) 1 1 1 0	•		34420	, A C A C ?		•		TTTAGCCTAT	· TAAACAGAGT '	TAATGACAGC
TICAAAAC	C1 (4500	DICACA	iocaa .	34520	111201010	34540		34560
ጥ ር ስጥርር ጥር	•	-	•	ግጉ ልጥ ሞ?	· CTAG	•	• ACTTATCCAT	TAGTAGTAAC	CAATAAAAA	• TATATATOA
ickiddic	GC /		34580			34600		34620		34640
AAAACTAT			*	rtaca(• GATGA	•	* AATTOOAOOO	GCTGGCTATA	TTCGCACTAG	* Aaataaaat
7002107117			34660			34680		34700		34720
CATTAGAT	rcg (\ATTT	ACGAG	* TCTCGTATAA	AATGTACAAT	AATTCACTTA	ATTTAATACT	• GCATATTTTT
			34740			34760		34780		34800
ACAAGTAG	• SAG	AGCGG'	TGATG	AAACA	• ATAAA	CGAAAGGCTT	TACATTAATT	GAATTAGTCA	* TCGTGATTAT	TATTCTCGGT
			34820			34840		34860		34880
ATACTTGO	TG	CTGTG	GCACT (GCCGA	• AATTC	ATCAATGTTC	* AAGATGACGC	TAGGATCTCT	GCGATGAGCG	GTCAGTTTTC
	-		34900			34920		34940		34960
ATCATTTO	SAA	AGTGC	CGTAA .	AACTA	TACCA	TAGCGGTTGG	• TTAGCCAAAG	GCTACAACAC	TGCGGTTGAA	AAGCTCTCAG
			34980			35000		35020		35040
GCTTTGGG	CA	AGGTA	• TTDTA	GCATC	* AAGTG	ACACAGGTTT	TCCGTACTCA	ACATCAGGCA	CGAGTACTGA	TGTGCATAAA
			35060			35080		35100		35120
GCTTGTG	GTG	AACTA	TGGCA	TGGCA	* COATT	GATACAGACT	TCACAATTGG	TGCGGTTAGT	GATGGCGATC	TAATGACTGC
			35140			35160		35180		35200
AGATGTC	• GAT	ATTGC	TTACA	ССТЛІ	CGTGG	TGATATGTGT	ATCTATCGCG	ATCTGTATTT	TATTCAGCGC	TCATTACCTA
			35220			35240		35260		35280
CTAAGGT	+ GAT	GAACT	* ACAAA	TTTA	AACTG	GTGAAATAGA	AATTATTGAT	GCTTTCTACA	ACCCTGACGG	CTCAACTGGT
			35300			35320		35340	_	35360
CAATTAC	CAT	AAATI	TGGCG	CTTAT	* DAATO	TTGTACTTGC	TCTGACCGAC	ACAAATAATG	TCGTTTCTCA	GCATATATCA
			35380			35400		35420		35440
AAATACA	CAG	CAAAA	· Lattig	GGGTT	• PAGCTA	TATAGCTAAC	CCCAAATCAT	ATCTAACTTT	ACACTGCATC	TAATTCCAAA
			35460			35480		35500		35520
CAGTATO	CAG	CCAA	AAGCCT	AAAC	• Ta tt gt	TGACTCAGCG	CTAAAATAT	CGATGCAACA	AACAAGTCTT	GGATCGCAAT
			35540			35560		35580		35600
ACCTGAG	CTA	TCAA	, Dotaa	TCAC	TCATO	AGCACTTTGA	CGTCCTGTTC	CGGACTCGTT	TATCACCTGA	CCAATCTCAA
			35620			35640		35660		35680
TTATCGG	CGT	ATTT	CTGCTA	TCTT	GAAACT	CACCAATAAC	AATAGATTG	GAAGCAAAG1	CGCAAAACAA	GCGAGCATGA
			35700			35720)	35740)	35760
CTATATA	AGGT	CAGT	TGGCAA	CTCT	TGCTTA	CCCACTTAT	CAGCGCCCA	r TGCAGAAATA	TGCGTTCCTC	CTTGTACCCA
	_		35780			35800		35820		35840
CTGCGCT	TCA	ATA	AAGGCG	CTTG	AGCTG1	GGTTGCTGT	ATAATAATA	T CTGCTTGTT	ACAAGCAGCI	TGTGCATCAC
	_		35860			3588		3590		35920
AAGCTT	cGGC	ATTA	ATGCCT	TTTT	Taato	A AACGCTTAA	CAAGTTTTC	A GTTTTGCTA	G CACTACGGC	AACTACCAA
	_		35940	ı		3596	0	3598	0	36000
ACCTTA	GTTA	ATGA	ACGAAC	CTTC	CTCAC	- F GCTAGCACT	T CATATTCAG	C CTGATGACC	G GTACCAAAA	CAGTTAATA
•			36020	•		3604	0	3606	0	3608
	•	-	•			•				

35920 ACCAAT 36000 Fig. 4 TAATAC 36080 28/30

CGTAGCATCT T	CTCTCGCGA	GGTAACTCAC	TGCTACTGCA	TCGGCAGCAC	CAGTGCGGTA	AGCATTAACG	GTAGTGGCAG
	36100		36120		36140		36160
CAATCACCGN C	* TGCAACATA	CCGGTTAATG	GATCGAGTAA	AAATACGTTA	GTGCCCTGGC	ATGGTAAACC	ATGTTTATGG
	36180		36200		36220	•	36240
TTATCAGGCC A	ATAGCTGCC	TGTTTTCCAG	CCGACAAGGT	TTGGCGTTGA	AGCCGACTTT	AATGAGAACA	TTTCATTAAG
	36260		36280		36300		36320
GTTCGCGCCC 1	• AATTADƏTƏT	CTACCGGGAA	CAAGGTTGCT	TTATCATCTA	CGGCAGCGAC	AAACGCTTCT	TTAACAGCGA
	36340		36360	_	36380		36400
TATAAGCCAG	• TCATGGGAG	ATGAGCTTTG	ATGTTTGCGC	TTCAGTTAAA	TAGATCATAT	TACCACCCCT	GCACTCGATT
	36420		36440		36460		36480
CCAGATCTCA	• TACCACCDAT	TATCACCATC	AGTATCAAAT	ACATGGTACT	GAGCGTGCAT	TGAAGCTGTT	GCACAGGCGT
	36500		36520	_	36540		36560
GGTTCGGCAA	• Adtatgtaga	CGACTACCTA	CCGGGAACTG	CGCTAAATCA	ATAACGCCGC	CATCAACTGC	TTCAATAATG
	36580	ı	36600		36620		36640
CCGTGCTCTT	• GATTAACAGT	TATAACCTGT	AGACCTGATA	ACACGTGACC	GCTGTCGTCA	CACACTAAAC	CATAACCACA
	36660	1	36680		36700		36720
ATCTTTTGGC	* TGCTCTGCAG	TACCTCTATO	ACCCGAAAGA	GCCATCCAAC	CCGCATCAAT	GAAAATCCAG	TTTTTATCAG
	36740)	36760	,	36780		36800
GATTATGACC	AATAACACTO	GTCACTACCO	TTGCGGCAAT	ATCAGTTAAC	TGACACACGI	TTAGCCCTG	CATGACTAAA
	36820		36840)	36860)	36880
TCGAAGAAGG	TGTACACAC	CGCTCTAAC	TCGGTGATC	CATCAAGGTT	TTGATAGCT	TGCGCTGTT	G GTGTTGAACC
	3690	0	36920)	36940)	36960
AATACTAACG	ATGTCACAT	r GCATACCCG	TGCGCGAAT	G CGTCAGCAG	TTGTACAGC	GCTGCAACT	T CATTTTGCGC
	3698	0	3700	0	37020		37040
CGCATCAATT	AATTGCTGT	* T TTTCAAAAC	A TTGATATGA	C TCACCAGCG	T GAGTNAGTA	GCCGTGAAA	A CTCGCTGCGC
	3706	0	3708	0	3710	0	37120
CAGACGTTAG	TATCTGAGC	A ATTTCAATC	A ACTTATCGG	TTCCGGTGG	A ATACCACCA	C GATGGCCAT	C ACAATCAATT
	3714	0	3716	0	3718	0	37200
TCAATTAATG	CTGGTATTT	G GCAGTCATA	A GAACCACAG	A AATGATTTA	G CTGATGCGC	T TGCTCAAC	C TATCAAGTAA
	3722	0	3724	0	3726	0	37280
AACTCTTGCA	TTAATACCI	T GGTCCAACA	TTTAGCAAT	A CGCGGCAAC	T TACCATCGG	C AATACCTA	T GCATAAATAA
	3730	00	3732	0	3734	0	37360
TGTCTGTGTA	ACCTTTAGE	T GCTAAGGC	T CGCCTCTI	TACCGTTG	T ACAGTGACT	G GTGAGTTT	TT AGTGGGTAAT
	3736	30	3740	00	3742	0	37440
AAAAACTCGC	CTGCTTCA	AG TGATCTTA	AC GTTTTAAAJ	T GCGGTCTT	G GTTTGCAC	T AATCCTTC	AA TTTTTTGGCG
	374	60	3740	30	3750	00	37520
TAGTTGACT	AGGTTATT	AA TAAATACT	GG CTTATTTA	AAAAATAT AS	CG GTGTATCA	AT TECTTEAT	AC TGACTTTGCT
	375	40	375	60	375	BO .	37600
GAGTCGTGG	A AAGTATTT	GA GTAGATGG	CA TCTTTAAT.	AT CCTAGTTC	AT CAATCAAT	CT AACAAGTI	TG ATGCCTAGCC
	376	20	376	40	376	60	37680
ACAGTGGCT	T GTATTCAT	GA TGCTTTGG	AA AATGCTTA	TA TTCAAAGT	AT TTGAAAGA	CA TCAAACT	CT TGTTTAATGC
	. 377	•	377	•	377	•	37760
TCAGTATCC	A CCAGCACO	CA TTTATTT	ATTAACTA	TT ATCAAGAI	'AT AGATTAGG	TT CAAACCA	AT GATTAGTACT
	377	•	378	•	378	•	37840
GAAGATCTA	C GTTTTATO	CAG CGTAATCO	SCC AGTCATCO	CA CCTTAGCT	GA TGCCGCTA	GA ACACTAA	ATA TCACGCCACC

Fig. 4 29/30 WO 98/55625

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37860 37880
ATCAGTGACA TTAAGGTTGC AGCATATTGA AAAGAAACTA TCGATTAGCC TGATC

Fig. 4 30/30

Fig. 5

Fig. 5

Fig. 5

Fig. 5

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11410	11420	11430	11440	11450	11460
TGTAATTCAATG	TGGAATCGATA	ATTTAATGGC	TTAAAAGTGA	AGATCCATTA	ATTGTGA
			.*		
11470	11480	11490 *	11500	11510	11520
TGGCGAGGTGAT.	AGACCAATGTA	GACCTTAATG	AATAAAGCAG	GCACGATTG	LATCCATT
11530	11540	11550	11560	11570	11580
CAACGCAAAGTG	GTACTAACTAT	TGTTTTAAAC	GTTATAAATA	GTGTTTTAA.	AGGTTATA
11590	11600	11610	11620	11630	11640
AGTAAATAATTT	АААААСААТАА	TAATCCACAT	GCATTAAATT	TATCATGATA	AACCGCT
	11660				11700
ATATCTCAATGG	CAATTTGGGAT	AAGTGTAAAA	TATATGTAAA	ATGAATGAG1	TGACTTG
11710	11720	11730	11740	11750	11760
CTTTTTTTACAC	TAAGTGATGAA	ATTAAAGCT?	GATGTCGTTG	TTAGCATTG	AATAATTA
		11790			11820
CGTACTAAAATA	CGACATCTAG1	TATAGAAATT	PAAAAAAT	TGGTTTTGA:	TAGCATAA
	11840		11860		
CTGCATAAACTA	ATCAGCTTATT	GTCTGTAAT	ATTTTTGTAAT	rttaaatagg'	AATAATTT
11890			11920	11930	11940
AATTATATGTCT	'GATAAATATA	AACCGTACGA(CCTTTCCTTT	AAAAAGACGT	TTTTGCTG
					10000
	11960				
CCTAAGTTTTGG	CCTGTGTGGT'	ICGGGGTGTT'	TGCAATATAC'	PTATTAGCTT	TTATGCCA
12010	12020	12020	12040	12050	12060
12010					
GTAAAGCCGCGT	rgataaattig	CTCGATTCAT.	AGCGAAGAAA	TIGITIAGIC	IMMANIG
12070	12080	12090	12100	12110	12120
12070 ATGGCAAAGCG					
ATGGCAAAGCG	TAAAAAGGTAG	CAAAGAICAA	IIIMICIMIG	1901100010	MARIGORI
12120	12140	12150	12160	12170	12180
. 12130 GATACGGAACA					
GATACGGAACA	HONCCGINIAN	ICAIGGICAA	ICINGIIACI	IIIIGICAM	CIRICIII
12190	12200	12210	12220	12230	12240
AGTTATGCAGA					
NOTINICAGA	3CCAAG1GCGC	GIAGICGIGC	TIMIMICCOL	0.1000111101	
12250	12260	12270	12280	12290	12300
GGTGGCGAGAA'					
GGIGGCGAGAA	111A111CCGC	INCITORACA	MOOTANOOCI	10111011111	
12310	12320	12330	12340	12350	12360
CATAGCTTCGC					-
00011000		C.1.001 1, 11.01.			
12370	12380	12390	12400	12410	12420
ACTATGTTTAA					
12430	12440	12450	12460	12470	12480
ATGTTTGGAGG					
12490	12500	12510	12520	12530	12540
AAGAGCGGTGA					
.2.01.0000101					

AM

Fig. 5

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Fig. 5

Fig. 5

2965	n 29	660	29670	29680	29690	29700
					CAGTTCAAAAC	
2971				29740		29760
TAAATAATA	ATCAAAAT	ACCTTCCG	rgaattacc <i>i</i>	GAACAACGC	rggaaaggcat	GGAAA
2977				29800		29820
GTAACGCTA	ACGTCATG	CAGTCGTT	ACAATTACGC	AAAGCGCCTA	AAAGGCAGTTA	CGTTG
2002	۸ ۵۵	0.40	20050	29860	29870	29880
2983	ע ב דמטדדמידמי	**************************************	Z 9090 TTTTA A AGTI	CCGCCTAAT	GAAAAAGATTG	
AACAGCIAG	AIAIIGAI	11011000				
2989	0 29	900	29910	29920	29930	29940
TCCCGCAAC	AGTTAATG			CAATGCTGCG	AAAGACGGAGG	TCTAG
2995		960		29980		30000
TTGAAGGTC	GTAATGTT	GCGGTATT	AGTAGCGAT	GGCATGGAA	CTGGAATTACA	TCAGT
				2224		20060
3001				30040	30050 TTACAGCAAGG	30060
ATCGTGGTC	GCGTTAA1	TCTAACCAC	CCAAATIGA	AGACAGCIIA	TIACAGCAAGG	INIIN
3007	10 30	080	30090	30100	30110	30120
	-				GACGGTGTTGC	
3013	30 30	0140	30150	30160	30170	30180
CTGCACAGO	TAAATCA	STATACGAC	TTTCATTGG	TAATATTATG	GCGTCACGTAT	TTCGG
3019			30210	30220	30230	30240
CGTTATGG	SATTTTTC	IGGTCCTGC	TATTACCGT	ATCGGCTGAA	GAAAACTCTG:	TTTATC
302	=0 3	0260	30270	30280	30290	30300
					GAAGCCGTTA	
GIIGIGII	JANTINGC	10AMM1C	AIIICALLO		01111000011111	
303:	10 3	0320	30330	30340	30350	30360
CTGCTGTT	GATTTGTC'	TGGTTCAAT	TGAAAACAT	TACTTTACGI	CAGCACTACG	GTCCAG
303	70 3	0380	30390	30400	30410	30420
TTAATGAA	AAGGGATC	TGTAAGTG!	AATGTGGTCC	GGTTAATGA	AGCAGTTCAG	TAACCA
	_					
304		0440	30450			30480
ACAATATT	CTTGATCA	GCAACAAT	SGCTGGTGGG	TGAAGGCGCA	AGCGGCTATTG	TCGTTA
304	00 3	0500	30510	30520	30530	30540
			-		GATGCGGTGA	
12.0001011	100010101	CC.1 OC.1 O.				
305	50 3	0560	30570	30580	30590	30600
CCCCTGGT	AGCAATGC	GAAAGCAA	TTACGATTGO	AGCGGATAA	AGCATTAACAC	TTGCTG
			,			
		0620			30650	
GTATCAGT	GCTGCTGA	TGTAGCTA	GTGTTGAAG	ACATGCAAG	IGGTTTTAGTG	CCGAAA
_						
				30700		30720
ATAATGCT	GAAAAAAC	CGCGTTAC	CGACTTTATI	ACCCAAGCGC	AAGTATCAGTT	CGGTGA
207	30 3	0740	30750	30760	30770	30780
					UTTUE AAAATTATTAI	-
·······			0001040	J.III COCONG		
		<u></u> .	_			

Fig. 5

TGCATATCACTGAGATCGTGAATGACGCTGGTGAAGTGCGAATCGTTGGTGATGCGAATC

35350	35360	35370	35380	35390	35400
ACCGACGATTA	ATTGGTCTGCGTG	ATGAAGTGCA	AGCGAAGTAT	AACTTCTCTC	CTGCATT
35410	35420		35440	35450	35460
ACGTGTTGGTC	CTGGTGGTGGTA	TCGGAACGCC	TGAAGCAGCA	CTCGCTGCAT	TTAACAT
35470		35490	35500	35510	35520
GGGCGCGGCT	TATATCGTTCTGG	GTTCTGTGAA	TCAGGCGTGT	GTTGAAGCG	
35530	35540	35550	35560	35570	35580
TGAATATACT	CGTAAACTGTTAT	CGACAGTTGA	LAATGGCTGAI	'GTGACTATG	
35590			35620	35630	35640
TGCAGATATG'	TTGAAATGGGTG				
35650		35670		35690	35700
GATGCGTGCG	AAGAAACTGTATG				
35710	35720		35740	35750	35760
AGCTGCTGAA	CGTGAGAAGATTC				
35770	35780	35790	35800	35810	35820 GCAACGAG
GGATGGCACT	ATCGCTTTCTTT!	ACTGAACGCG	ATCCAGAAAT	3CTAGCCCGT	GCAACGAG
35830	35840	35850	35860	35870	35880
TAGTCCTAAA	CGTAAAATGGCA	CTTATCTTCC	GTTGGTATCT	rggcctttct	TCACGCTG
35890		35910	35920	35930	35940
GTCAAACACA	GGCGAGAAGGGA	CGTGAAATGG	ATTATCAGAT	TTGGGCAGGC	CCAAGTTT
35950		35970			
AGGTGCATTC	AACAGCTGGGTG	AAAGGTTCTT	ACCTTGAAGA		
36010		36030	36040	36050	36060
TGTAGATGTT	GCTTTGCATATG				
36070		36090	36100	36110	36120
	AGGTGTTAGCTTA				
36130					36180
TTACTTGAT	SATATGTGAATTA	ATTAAAGCGC	CTGAGGGGG	TTTTTTGG.	IIIIIACI
36190	36200	36210	36220	36230	36240
CAGGTGTTG:	raactcgaaatte	CCCCTTTCA	AGTTAGATCG	TTACTCACT	CACAATATG
2525	36260	36270	36280	36290	36300
	CACTTGCCATAT <i>i</i>				
TIGHTHICG	eac i i occai ai i	ici iccieni			
3631		36330	36340	36350	36360
TAGTCTTTA	ATATCCGAGTCT	TTCTTCAGCA	TAATACTAAT	ATAGAGACTC	GACCAATGT
	0 36380				
TAAACACAA	CAAAGAATATAT:	CTTGTGTAC	IGCCTTATTA	ITAACGAGTG	CGAGTACGA
3603	0 36440	36450	36460	36470	36480
	CGCTAAACAATT				
	_	_			

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39910 39920 39930 39940 39950 39960 TATGGCCATCGAATTTGCAAAATCAGGTCATAACTTAGCACTTTGTGCACGTAGACTTGA

39970 39980 39990 40000 40010 40020 TAATTTAGTTGCACTGAAAGCAGAACTCTTAGCCCTCAATCCTCACATCCAAATCGAAAT

40030 40040 40050 40060 40070 40080 AAAACCTCTTGATGCAATGAACATGAACAAGTCTTCACTGTTTTCCATGAATTCAAAGC

40090 40100 40110 40120 40130 TGAATTTGGTACGCTTGATCGTATTATTGTTAATGCTGGATTAGGCAAGGGTGGATCC

Fig. b

Fig.b

tig.6

TTGCGCAAACCTTGTCTGCAATTACCGCTGCTGGTGGCCAAGCTGAATATGTTTCTGCAG

7000	8000	8010	8020	8030	8040
7990 TTAAGCAGATAT					
8050	8060	8070		8090	8100
TTGCAACAGCTA	AGCAGGGATCC	TTCCCGTTAG	CTGACAACAA:	PATCTTTGCC	AATGATT
8110	8120	8130	8140	8150	8160
	CTATGTTGGTC	TGGGTGCGCA	AACAATTTGG:	TTAGGTAGC	TTACCTT
8170	8180	8190		8210	8220
CGGTGACAACG	CTTGGACTGTG	TATCGTGAAG	TGGTTGTAGA	IGAAGTATTI	TATCIGC
8230	8240	8250	8260	8270	8280
AACTTAATGTT	ettgagcatgat	CTATTGGGTT	CACGCGGCAG	TAAAGCCCGI	TGTGATA
				2222	0240
8290	8300 GCTGCTGATATG	8310 CD DURB CUTC	8320 CCGAAGTGAA	8330 atcagggai	
TICAATIGATI	3C1GC1GA1A1G	CAATIACTIG	CCGAAGIGAA	AICAGCGCAP	101CAG10
8350	8360	8370	8380	8390	8400
TCAGTGACATT'	TTGAACGATATG	TCATGATCGA	GTAAATAATA	ACGATAGGC	STCATGGT
244.2	0.420	0.420	0.4.4.0	0.450	8460
8410	8420 CTGCTTTCTTCA	8430	8440	8450 TARTAGCTA	
GAGCATGGCGT	C10C111C11C1		.1 110101011111		
8470	8480	8490	8500	8510	8520
TGCTTTAAACC	AAGTAAACAAGT	GCTTTTAGCT	ATTACTATTC	CAAACAGGA'	TATTAAAG
0.5.2.0	8540	8550	8560	8570	8580
8530 AGAATATGACG	GAATTAGCTGTT				
	0.1		,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,		
8590			8620	8630	8640
TTGACCGTGTG	GAACGCGCTTTC	TATGAAGGT	CTTATGTAGG	STAATGTTAG	CCGCGTTA
8650	8660	8670	8680	8690	8700
- -	AATGTTATTAGO			+	
8710	8720	8730	8740	8750	8760
TTAACTCTGTC	AGTCTACTAGCC	GCAAACGAAT(CAGTTAAATA	ragctgatat	CGCGGTGT
8770	8780	8790	8800	8810	8820
• •	GATGTAAAAAG				
8830	8840	8850	8860	, 8870	8880
TTGAAAAACAG	TGTGCGAGTTG:	rgttgttatt(GCTGATTTAG	SCCAAGCATT	AAATCAAG
8890	8900	8910	8920	8930	8940
	GTTAATAACCA			TTGGCATGAA	TAACTCGG
8950	8960	8970	8980	8990	9000
TTAATTTATCT	CGTCATGATCT	IGAATCTGTA	ACTGCAACAA'	TCAGCTTTGA	TGAAACCT
9010	9020	9030	9040	9050	9060
TCAATGGTTAT					
3	MACAAIGIAGC.	100011000			
	MACAAIGIAGC	1000110000			
9070	9080 SCAATGTTATAT	9090	9100	9110	9120

Fig. b

WO 98/55625 PCT/US98/11639

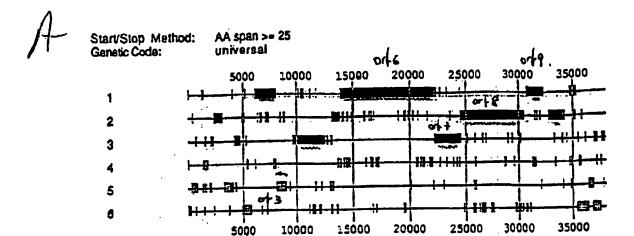
11410	11420	11430	11440	11450	11460
AAGGTGACACA					CAATTTTG
			11500		
ATGCTTCAGGT	TATCAACTCGAT	TAATGATTAT	TAGCCGGIII	AGAIGACCI	MAICAMI
11530	11540	11550	11560	11570	11580
GGGGGCTTTAT	GTTACGAAACA	AGCCCTTACCC	GATGCGGGTTA	TTGGGGCAG	PACTGCAC
			11620	11630	13640
11590 TAGAAAACTGT			11620		11640
INGAAAACIGI	GGIGIGAIIII	NGGIANIIIG.	icmi iccoian		
		11670		11690	11700
TGTTTATGCCT	TTGTATCATCA.	AGTTGTTGAT	aatgccttaa <i>i</i>	AGGCGGTATT	ACATCCTG
	11700	11720	11740	11750	11760
11710	11720 ACGCATTACAC.				
AIIIICAAIIA	ACOCIII IIICIIC				
11770					11820
CAGGTTATCCA	GCTGCATTGAT	CGCGCAAGCG	GCGGGTCTTG	GTGGTTCACA	TTTTGCAC
11830	11840	11850	11860	11870	11880
	TGTGCTTCATC				
11890		11910		11930	11940
CGGGTAAAGCC	CAACATGATGCT	TGCTGGTGCG	GTATCTGCAG	CAGATCCTAT	GTTCGTAA
11950	11960	11970	11980	11990	12000
	CTCGATATTCCA				
12010	12020	12030		12050	
AAAATTCACAA	AGGTCTATTTGC	CGGTGAAGGC	GCGGGCATGA	TGGTATTGAA	ACGTCAAA
. 12070	12080	12090	12100	12110	12120
	ACGTGATGGTGA		GCCATTATTA	AAGGCGGCGC	ATTATCGA
12130	12140	12150	12160	12170	12180
ATGACGGTAA	AGGCGAGTTTG1	ATTAAGCCCC	SAACACCAAGG	GCCAAGIAI	DIAIAIDA
12190	12200	12210	12220	12230	12240
AACGTGCTTA!	TGCCGATGCAG;	TGTTGACCC	GAGTACAGTTC	ACTATATTG!	ATGTCATG
				10000	10200
12250	12260 ACCTAAGGGTG		12280 TTGCGTTCG	12290 ייים או ארכייי	12300 ርምምድር እርሞር
CAACGGGCAC.	ACCIANGGGIGI	CAMIGITOM	11100011001	ii dommeet.	11101010
12310		12330			12360
GCGTAAATAA	CAAACCATTAC	rgggctcggt:	PAAATCTAACO	TTGGTCATT	IGTTAACTG
10070	12200	12200	12400	12410	12420
12370	12380 GCCTGGCATGA	12390			
CCCCIGGIAL	CCC1CCCA1GA	au			
12430					
CAACGATTAA	CTTAAAGCAAC	CACTGCAATC	TAAAAACGĠT	ractttactg(GCGAGCAAA
	10500	10510	10500	10534	12540
12490 TGCCAACGAC	12500 GACTGTGTCTT	· 12510		12530 AAGGCAGATA	
TOCCARCOAC	oc.iGIGICII		20000310001		

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+19.6

Fig. b

Fiq. 6



Page 1

B

Start/Stop Method: Genetic Code: AA span >= 25 universal 1. of 7' of 18 lart 6 åc€4 . JC.

FIG 7

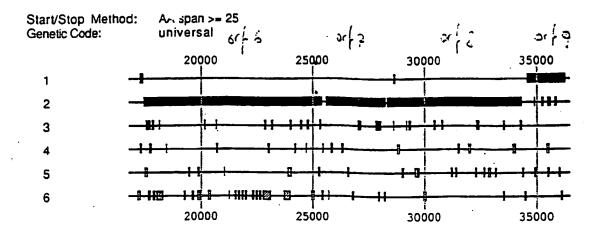
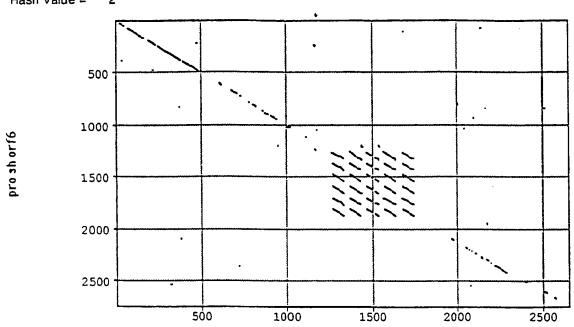


Fig. 8

Window Size = 8 Min. % Score = 60 Hash Value = 2

Scoring Matrix: BLOSUwi 62



Translation of vm6

Fig. 9

Window Size = 8 Min. % Score = 60 Hash Value = 2

pro shorf7

Scoring Matrix: BLOSUM 62

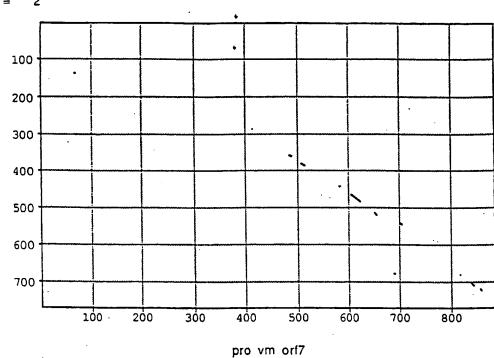
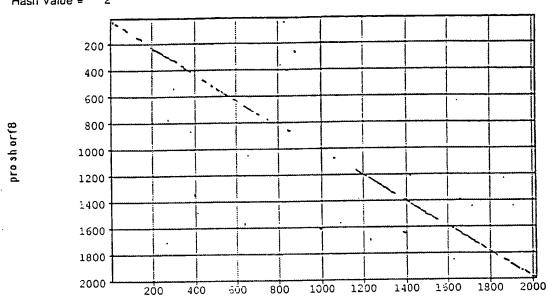


Fig. 10

Page 1

Window Size = 8 Min. % Score = 6 0 Hash Value = 2 Scoring Matrix: BLOS 62



pro vm orf8

rage 1

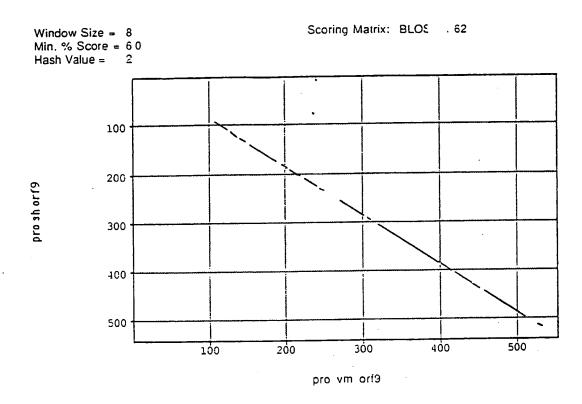


Fig. 12

CO' LEMENTATION Sp / Vm

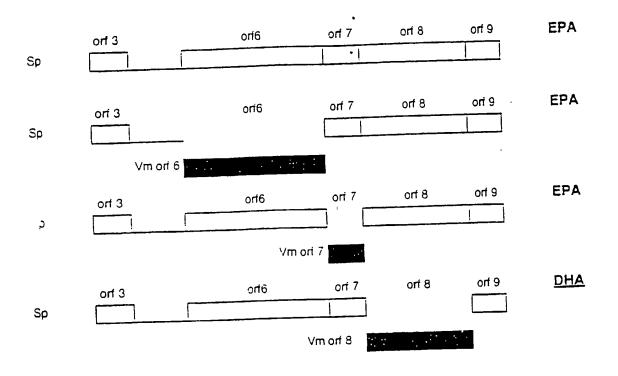
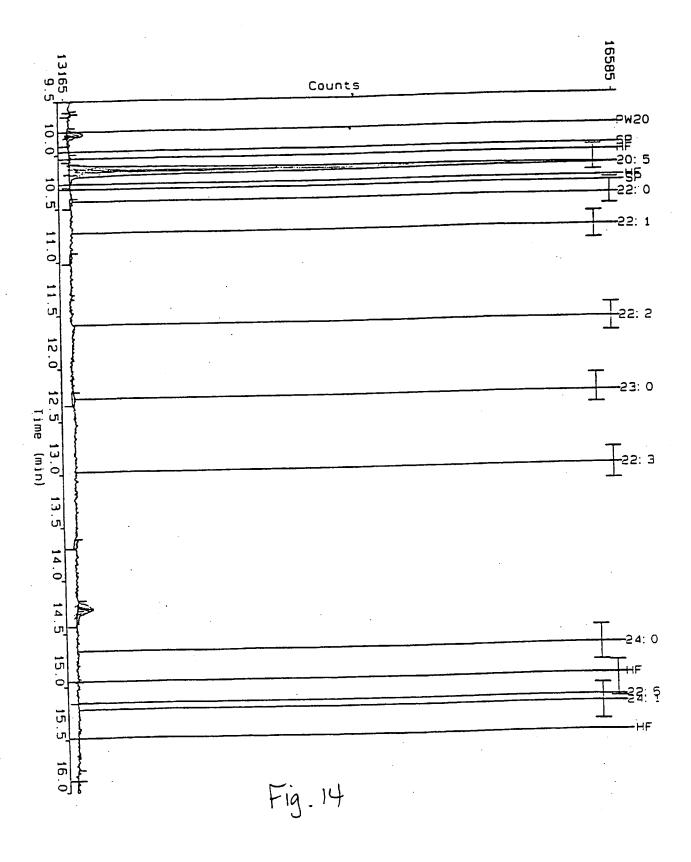
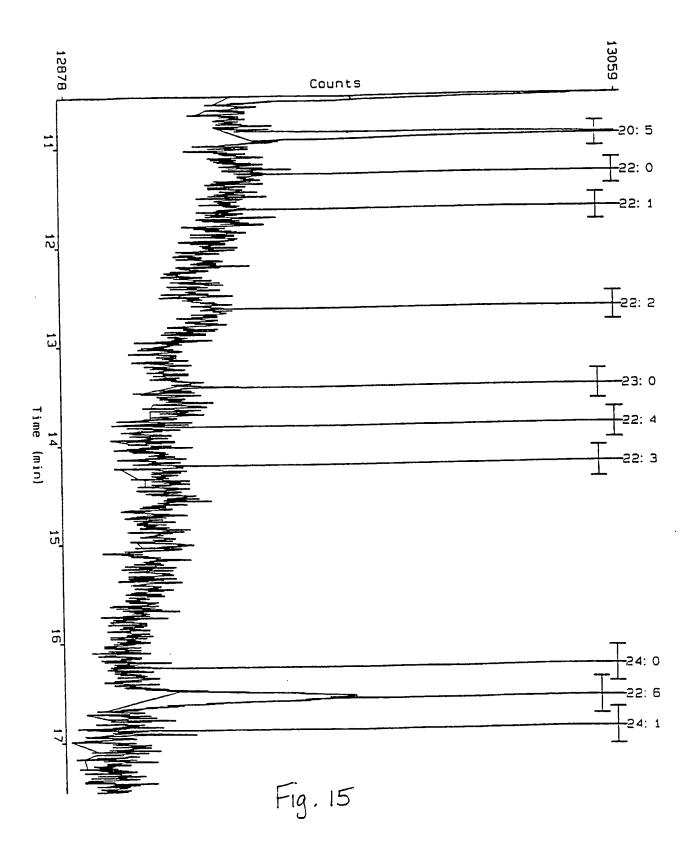


Fig. 13





DHA (% Fatty acids)	<u>20°C</u>
0.06	pEPAD8
0.70	4
0.66	5
0.22	6s
0.59	61
	<u>23°C</u>
0.06	pEPAD8
0.62	4
0.22	6s
0.65	6 1
	0.06 0.70 0.66 0.22 0.59 0.06 0.62 0.22

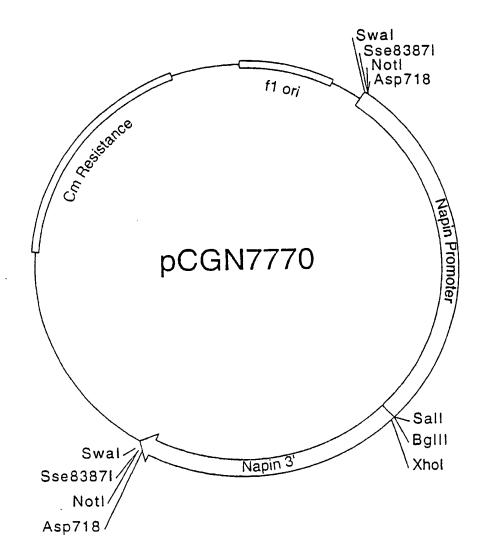
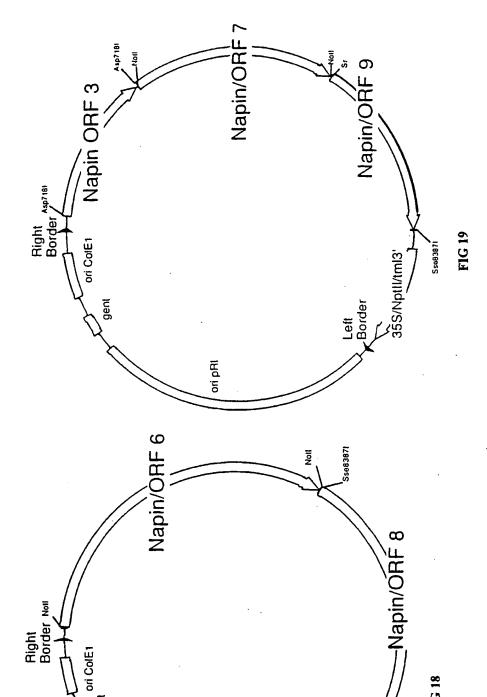


FIG 17

FIG 18

Sse83871

pCGN8537



ori pAl

pCGN8535

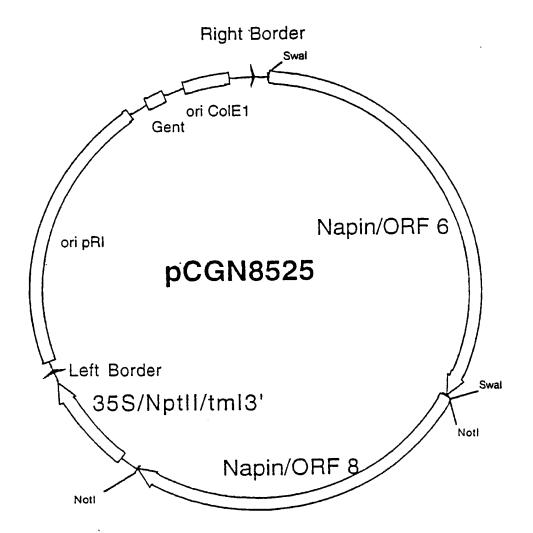
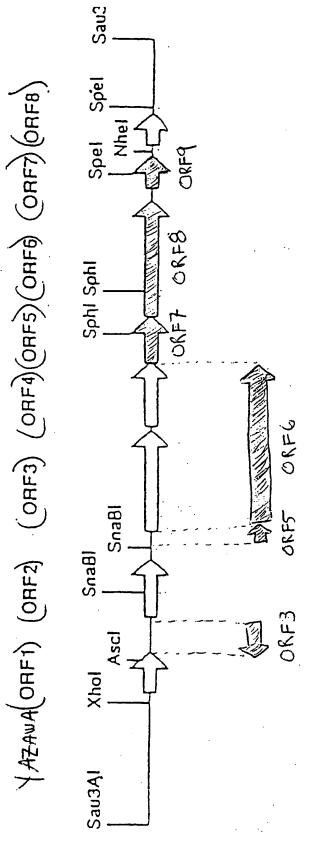
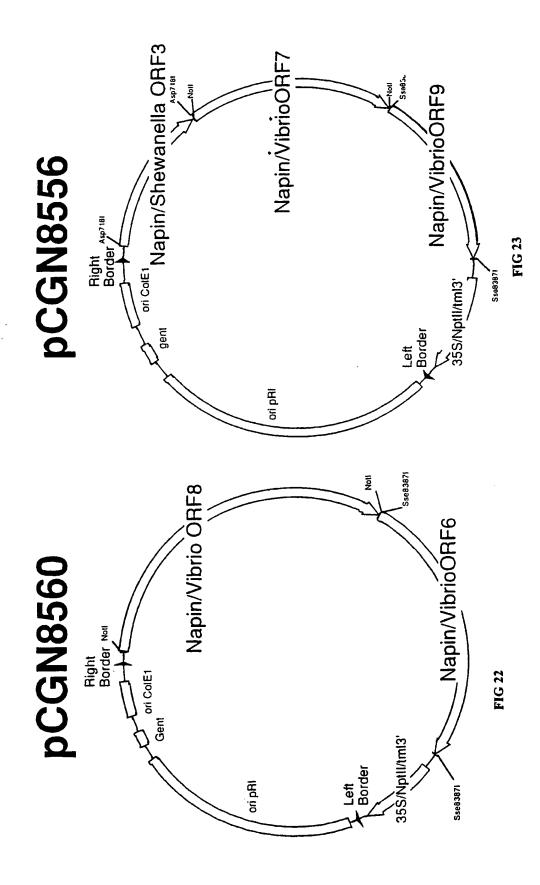


FIG 20



7IG 21



ATT GGT AAA AAT AGG GGT TAT GTT TGT TGC TTT AAA GAG TGT CCT GAA
I G K N R G Y V C C F K E C P E>

AAA TTG CTA ACT TCT CGA TTG ATT TCC TTA TAC TTC TGT CCG TTA ACA
K L L T S R L I S L Y F C P L T>

ATA CAA GAG TGC GAT AAC CAG ACT T T E L V K S W L>

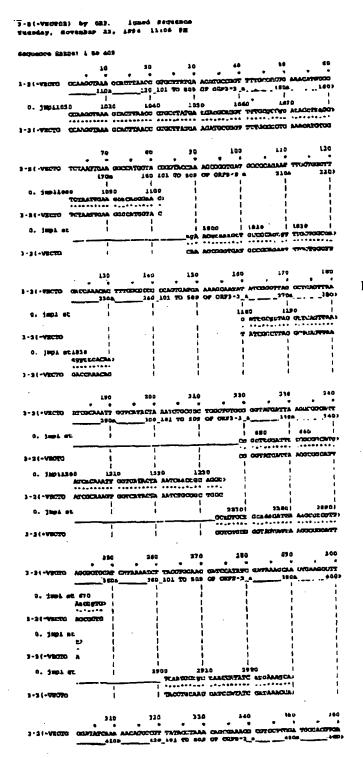
CCT GAA GAT GAG TTA ATT AAG GTT AAA CGC TAC ATT AAA CAA GAA GCT
P E D E L I K V N R Y I K Q E A>

AAA ACT CAA GGT TTA ATG GTA AGA G
K T Q G G L M V R>

FIG 24

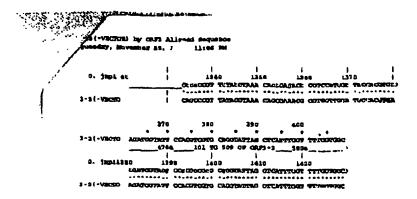
SS9 Photobacter

PCR Product Using Primers Presented in Example I



ORF 6
Probe Resulting from PCR with Primers
Presented in Example I

FIG 26A



INTERNATIONAL SEARCH REPORT

Int tional Application No PC:/US 98/11639

CLASSIFICATION OF SUBJECT MATTER PC 6 C12N15/31 C12N A. CLASS C12N15/52 C12N15/82 C12N15/70 C12N5/10 C12N1/21 C12P7/64 A01H5/00 According to International Patent Classification (IPC) or to both national classification and IPC B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) C12N C12P C07K A01H IPC 6 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) C. DOCUMENTS CONSIDERED TO BE RELEVANT Citation of document, with indication, where appropriate, of the relevant passages Relevant to claim No. Χ NAKAHARA, TORO: "Physiological activity 6,7, of docosahexaenoic acid (DHA) and its 11-13 production by microbial culture" YUKAGAKU (1995), 44(10), 821-7 CODEN: YKGKAM; ISSN: 0513-398X, XP002080682 see abstract 14,32 X NASU M ET AL: "Efficient transformation 25,27, of Marchantia polymorpha that is haploid 28,30 and has very small genome DNA; Agrobacterium tumefaciens-mediated transformation of suspension cell culture, for use in eicosapentaenoic acid. arachidonic acid and antibiotic production" J.FERMENT.BIOENG.; (1997) 84, 6, 519-23 CODEN: JFBIEX ISSN: 0922-338X, XP002080470 see the whole document Further documents are listed in the continuation of box C. χ Patent family members are listed in annex. ° Special categories of cited documents : "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the "A" document defining the general state of the art which is not considered to be of particular relevance invention "E" earlier document but published on or after the international "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such docudocument referring to an oral disclosure, use, exhibition or ments, such combination being obvious to a person skilled in the art. document published prior to the International filing date but later than the priority date claimed "&" document member of the same patent family Date of the actual completion of theinternational search Date of mailing of the international search report 14 October 1998 23/10/1998 Name and mailing address of the ISA Authorized officer European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016 Kania, T

3

INTERNATIONAL SEARCH REPORT

Intra Jonal Application No PCI/US 98/11639

		PC1/US 98/11639
	lation) DOCUMENTS CONSIDERED TO BE RELEVANT	
Category ?	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	KYLE D ET AL: "Long-chain omega-3 polyunsaturated fatty acids: prospects for introduction into horticultural food plants; e.g. alga eicosapentaenoic acid and docosahexaenoic acid gene cloning, expression in transgenic plant oil, crop improvement (conference paper)" HORTSCIENCE;(1990) 25, 12, 1523-26 CODEN: HJHSAR, XP002080471 * see the whole document, esp. p.1524, 2nd par. *	25-28, 30,31
X	EP 0 594 868 A (SAGAMI CHEM RES) 4 May 1994 cited in the application see the whole document	15-17, 19-22,24
X	WO 96 21735 A (SAGAMI CHEM RES) 18 July 1996 cited in the application see the whole document	15-17, 19-22,24
Α	YAZAWA, KAZUNAGA: "Production of eicosapentaenoic acid from marine bacteria" LIPIDS (1996), 31(SUPPL., FATTY ACIDS AND LIPIDS FROM CELL BIOLOGY TO HUMAN DISEASE), S297-S300 CODEN: LPDSAP; ISSN: 0024-4201, XP002080483 cited in the application see the whole document	1-32
A	SOMERVILLE C R: "Future prospects for genetic modification of the composition of edible oils from higher plants; oilseed crop improvement by lipid and fatty acid modification (conference paper)" AM.J.CLIN.NUTR.;(1993) 58, 2, SUPPL., 270S-275S CODEN: AJCNAC, XP002080472 * see esp. p.274S, r. col., lst par. *	1-32

INTERNATIONAL SEARCH REPORT

Information on patent family members

Inte ional Application No PCi/US 98/11639

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